

What Role do Circadian Rhythms Play in Learning and Memory?

Xiang Mou*

Department of Molecular and Cellular Biology, Baylor College of Medicine, Houston, USA

*Corresponding author: Xiang Mou, Department of Molecular and Cellular Biology, Baylor College of Medicine, Houston, USA, Tel: 713-798-5809; Fax: 713-798-5765; E-mail: xiangmou@gmail.com

Received date: March 08, 2016; Accepted date: April 12, 2016; Published date: April 19, 2016

Copyright: © 2016 Mou X. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

Circadian rhythmicity in gene expression and physiological process has been observed both in the central nervous system and in the periphery, including the hippocampus. As a center for memory formation and storage, the hippocampus shows circadian rhythmicity in clock gene expression and synaptic plasticity. Circadian variation in performance of hippocampus-dependent memory task suggests a link between clock gene oscillation and behavioral response. Yet the discrepancy in time scale between fast information encoding during memory acquisition and much slower circadian oscillation in cellular processes casts doubt on the underlying mechanism of circadian regulation of learning and memory. This short review suggests that instead of being a modulator of learning process and memory formation, the time-of-day information itself could be integrated as a component of episodic memory for later consolidation and retrieval.

Keywords: Circadian rhythm; Suprachiasmatic nucleus; Learning; Memory; Hippocampus

Introduction

Circadian rhythms are basic biological phenomena conserved in a variety of organisms. Endogenous circadian oscillators modulate a range of physiological and behavioral activities observed throughout phylogeny. Animals need intact learning ability and well-maintained memory to survive varying environmental conditions. However, the relationship between circadian rhythm biology and memory formation is far from well understood. This short review first summarizes the basic organization and functionality of circadian machineries in the central nervous system and in the periphery. It then reviews current knowledge about circadian modulation of learning and memory. Finally, it raises the question: Is circadian rhythmicity a modulator of learning and memory, or it is a component integrated into episodic memory during learning? I suggest that the time-of-day information could be encoded as a component of episodic memory rather than being an active modulator of learning and memory.

Circadian clocks exist in the central nerve system and in the periphery

Circadian rhythms are biological activities that oscillate under constant environmental conditions with a ~24 hour period. Circadian rhythms regulate the biological processes of diverse organisms ranging from prokaryotes to mammals [1-4]. Patterns of brain activity, hormone production, cell regeneration and many other biological activities are linked to this 24-hour cycle [5-7]. Circadian rhythms are endogenously driven: organisms maintain their behavioral and physiological rhythms even under constant environmental conditions.

In mammals, the master circadian center is located in the suprachiasmatic nucleus (SCN) of the hypothalamus [8,9]. In individual SCN cells, circadian machinery is self-sustained and maintained by a cellular feedback loop. In the positive feedback phase,

two transcription activators, CLOCK and BMAL1, dimerize and bind to E-box motifs in promoter segments and stimulates transcription of clock genes [1,10]. In the negative feedback phase, two clock gene (Per and Cry) products, Period and Cryptochrome proteins, form a heterodimer complex and inhibit CLOCK:BMAL1-mediated transcription through direct protein-protein interaction [11]. This inhibition down-regulates the transcription of clock genes including their own. The ensuing decline in Period and Cryptochrome protein levels eventually leads to reactivation of CLOCK/BMAL1-induced clock gene transcription and reinitiation of the cycle. Individually oscillating SCN cells use neuropeptide communication (primarily vasoactive intestinal peptide (VIP) [12-14]) and gap junctions [15,16] to promote synchronization and coordination among them.

Rhythmic clock gene expression underlies the rhythmic electrical activity, such as spontaneous firing rate, of SCN neurons [17,19]. In nocturnal rodents, spontaneous firing rate of SCN neurons show circadian rhythmicity with an elevated firing rate in subjective daytime [20-22]. It is possible that there is direct transcriptional regulation of ion channel proteins or regulatory factors affecting channel activity by CLOCK/BMAL1. Clock genes may also regulate electrical activity indirectly via clock controlled genes (CCGs), such as vasopressin, which appears to augment the magnitude of the electrical activity rhythm in the SCN through a receptor-mediated excitation of SCN neurons [23,24].

The SCN outputs are thought to synchronize a number of circadian oscillators in the periphery, such as lungs [25], pancreas [26,27], adipose tissue [28], adrenal glands [29,30] and ovaries [31,32]. Furthermore, independent circadian oscillators exist in the periphery, including the food-entrainable oscillator in the liver [33-35] and oscillator in the olfactory bulb [36,37].

Circadian fluctuations in clock gene expression and synaptic plasticity exist in the hippocampus

The hippocampus is a region in the brain critical for learning and memory formation. Interestingly, some clock genes are expressed in

the hippocampus and their expressions oscillate in a robust circadian manner. For instance, the *Per1* and *Per2* genes are reported to oscillate in both the dentate gyrus and CA1 regions of the hippocampus [38]; the oscillation of the latter in the dentate gyrus can be abolished by lesion of the SCN [39]. In addition to circadian fluctuations in gene expressions at the cellular level, time-dependent synaptic strengthening has been observed in rodent SCN. Stimulation of the optic nerve elicited long-term potentiation (LTP) only during the day [40]. This time-dependent LTP finds its parallel in the hippocampus. In both hamsters and mice, LTP was elevated in slices containing hippocampal CA1 prepared during light phase but tested during dark phase [41,42]. Conversely, in hippocampal slices prepared during dark phase but tested during light phase, LTP exhibited opposing activities [42]. These data raise the possibility that hippocampal LTP is dependent on the time of testing; they also support the hypothesis that an independent circadian center controls the hippocampal plasticity since the possible association between the SCN and the hippocampus was abolished in vitro.

Circadian modulation of learning and memory

The influence of circadian rhythm on learning and memory has long been studied. Circadian effects on different stages of memory formation have been postulated in various paradigms [43,44]. Chaudhury and Colwell showed that recall of contextual and cued fear memory in mice peaks in the early daytime [45]. They also showed that mice displayed the same periodicity of peak memory when housed under constant darkness condition and this peak memory is independent of the time of training. In contrast, other studies suggest that memory formation is dependent upon time of training [46-49]. Rats demonstrate better acquisition and performance on an operant task when trained during the dark phase. Spatial learning ability assessed by Morris water maze was not affected by the time of training; but better long term spatial memory was achieved if animals were originally trained during the dark phase [47]. These studies indicate that the time of training, rather than an endogenous circadian system, appears to be the critical factor for memory formation.

Given that some clock genes have been found in hippocampus, it is natural to ask whether the circadian regulation of learning and memory formation are attributed to hippocampus-specific clock gene fluctuation; or one might think that circadian machineries may exert their functions through other neuronal processes and, therefore, affect learning and memory indirectly. Most manipulations of the circadian clock also lead to sleep disruption, which is a major cause of learning deficits and memory dysfunctions. As a result, the cellular mechanisms of circadian regulation of learning and memory remain vague. Using a non-invasive means in Siberian hamsters, Ruby et al. recently showed that circadian system is involved in memory function independent of sleep [50].

It is worth noting that the presence of clock genes in the hippocampus does not necessitate them for hippocampus-dependent learning and memory. To elucidate the link between circadian clock and hippocampus-dependent learning and memory, region-specific alterations of circadian clock machinery in the hippocampus would be necessary. Rather than the coarse ablation of the SCN area, spatially restricted manipulations of clock genes in subregions of the hippocampus would allow us to determine whether disruptions of circadian clock affect learning and memory at cellular as well as behavioral level. With the advent of novel biotechnologies, including cell type-specific genetic manipulation and optogenetics [51,52],

questions such as which group of cells that express clock genes undergo activation after learning can be addressed.

Mechanisms underlying circadian regulation of learning and memory

The aforementioned evidence along with other experimental results point to the notion that different stages of memory formation along with their cellular substrates are under the modulation of circadian rhythms. However, whether the time-dependent LTP observed in hippocampus and/or the circadian modulated memory formations are SCN-dependent are questions to which answers remains largely elusive. Anatomically, the SCN has connections with many brain regions [53-55]. Of particular interest is its projections to the hippocampus, both directly [55] and indirectly via the locus coeruleus [56], which in turn mediates hippocampal activation [57]. Whether these connections are responsible for the circadian expressions of clock genes in hippocampus, or hippocampal synaptic plasticity is not known. Since the LTP has been thought the cellular basis of long-term memory formation [58,59], it would be premature to propose that hippocampus-dependent learning and memory is modulated by the SCN pacemakers until it can be confirmed that disruptions of SCN-hippocampus connection lead to memory dysfunctions.

The circadian clock could regulate learning and memory through hormonal signalling. For instance, melatonin administration inhibits LTP in the hippocampus [60,61]. In rats, melatonin modulates memory in a phase-specific manner [62]. Genetic deletions of melatonin receptors in mice lead to enhanced cognitive performance [63]. Glucocorticoid is another hormone that has been shown to exert rhythmic regulation of neuronal plasticity. High glucocorticoid levels increase learning-induced spine formation, while low glucocorticoid levels do not [64]. Meanwhile, prolonged exposure to high glucocorticoid levels disrupts previous memories and eliminates new spines induced by learning [64].

But is the link between hormones and learning and memory direct? Or do hormones regulate learning and memory indirectly via affecting neuronal plasticity in general? Most learning processes take place at a time scale much smaller than 24-h period, rendering the former possibility less likely. Recent findings of time cells in hippocampus [65,66] suggest that a subset of hippocampal cells can keep track of temporal elapses independent of pacemaker cells in the SCN. During the acquisition phase, the time-of-day information could be encoded as a contextual component, therefore becoming a time "tag" of episodic memory. This would ensure that during later test phases, animals demonstrate significantly better memory performance at the same time of training [67,68]. This encoding process can be SCN-independent so that animals with lesioned SCN demonstrate similar performance in memory tasks as their wild-type counterparts [69].

Conclusion

Circadian rhythmicity in cell physiology is conserved across a wide variety of organisms. The presence of circadian variations in gene expression and synaptic plasticity in hippocampal cells, as well as in learning and memory formation, indicates an inherent link between cellular activities and behaviors of the whole animal. However, the mechanism by which the activity of a group of pacemaker cells is translated into behavioral responses is still poorly understood. Anatomical and physiological evidence summarized above and many other studies indicate that pacemaker cells in the SCN may modulate

circadian variations in learning and memory formation directly and/or indirectly. However, contemporary theories of memory consolidation suggest that newly-acquired episodic memories are replayed during ripples for consolidation at tens of millisecond time scale, both immediately after learning and remotely during sleep [70-72]. How could cellular processes oscillating at ~24 h cycles influence the memory encoding at such finer time scales? It is more plausible that during learning, the time-of-day information is embedded as a component of episodic memory which is consolidated along with other memory components during subsequent replays. This time “tag” of episodic memory could be encoded independent of the SCN or local circadian center in the hippocampus. However, the elucidation of the relationship between circadian rhythms and memory formation warrants further research.

References

1. Bell-Pedersen D, Cassone VM, Earnest DJ, Golden SS, Hardin PE, et al. (2005) Circadian rhythms from multiple oscillators: lessons from diverse organisms. *Nat Rev Genet* 6: 544-556.
2. Kondo T, Ishiura M (2000) The circadian clock of cyanobacteria. *Bioessays* 22: 10-15.
3. Partch CL, Green CB, Takahashi JS (2014) Molecular architecture of the mammalian circadian clock. *Trends Cell Biol* 24: 90-99.
4. Sharma VK (2003) Adaptive significance of circadian clocks. *Chronobiol Int* 20: 901-919.
5. Mong JA, Baker FC, Mahoney MM, Paul KN, Schwartz MD, et al. (2011) Sleep, rhythms, and the endocrine brain: influence of sex and gonadal hormones. *J Neurosci* 31: 16107-16116.
6. Plikus MV, Van Spyk EN, Pham K, Geyfman M, Kumar V, et al. (2015) The circadian clock in skin: implications for adult stem cells, tissue regeneration, cancer, aging, and immunity. *J Biol Rhythms* 30: 163-182.
7. Rivkees SA, Hao H (2000) Developing circadian rhythmicity. *Semin Perinatol* 24: 232-242.
8. Moore RY, Eichler VB (1972) Loss of a circadian adrenal corticosterone rhythm following suprachiasmatic lesions in the rat. *Brain Res* 42: 201-206.
9. Stephan FK, Zucker I (1972) Circadian rhythms in drinking behavior and locomotor activity of rats are eliminated by hypothalamic lesions. *Proc Natl Acad Sci U S A* 69: 1583-1586.
10. Reppert SM, Weaver DR (2002) Coordination of circadian timing in mammals. *Nature* 418: 935-941.
11. Okamura H, Yamaguchi S, Yagita K (2002) Molecular machinery of the circadian clock in mammals. *Cell Tissue Res* 309: 47-56.
12. Aton SJ, Colwell CS, Harmar AJ, Waschek J, Herzog ED (2005) Vasoactive intestinal polypeptide mediates circadian rhythmicity and synchrony in mammalian clock neurons. *Nat Neurosci* 8: 476-483.
13. Itri J, Colwell CS (2003) Regulation of inhibitory synaptic transmission by vasoactive intestinal peptide (VIP) in the mouse suprachiasmatic nucleus. *J Neurophysiol* 90: 1589-1597.
14. Piggins HD, Cutler DJ (2003) The roles of vasoactive intestinal polypeptide in the mammalian circadian clock. *J Endocrinol* 177: 7-15.
15. Colwell CS (2005) Bridging the gap: coupling single-cell oscillators in the suprachiasmatic nucleus. *Nat Neurosci* 8: 10-12.
16. Rash JE, Olson CO, Pouliot WA, Davidson KG, Yasumura T, et al. (2007) Connexin36 vs. connexin32, “miniature” neuronal gap junctions, and limited electrotonic coupling in rodent suprachiasmatic nucleus. *Neuroscience* 149: 350-371.
17. de Jeu M, Hermes M, Pennartz C (1998) Circadian modulation of membrane properties in slices of rat suprachiasmatic nucleus. *Neuroreport* 9: 3725-3729.
18. Pennartz CM, De Jeu MT, Geurtsen AM, Sluiter AA, Hermes ML (1998) Electrophysiological and morphological heterogeneity of neurons in slices of rat suprachiasmatic nucleus. *J Physiol* 506: 775-793.
19. Schaap J, Bos NP, de Jeu MT, Geurtsen AM, Meijer JH, et al. (1999) Neurons of the rat suprachiasmatic nucleus show a circadian rhythm in membrane properties that is lost during prolonged whole-cell recording. *Brain Res* 815: 154-166.
20. Kononenko NI, Kuehl-Kovarik MC, Partin KM, Dudek FE (2008) Circadian difference in firing rate of isolated rat suprachiasmatic nucleus neurons. *Neurosci Lett* 436: 314-316.
21. Kuhlman SJ, McMahon DG (2004) Rhythmic regulation of membrane potential and potassium current persists in SCN neurons in the absence of environmental input. *Eur J Neurosci* 20: 1113-1117.
22. Pennartz CM, de Jeu MT, Bos NP, Schaap J, Geurtsen AM (2002) Diurnal modulation of pacemaker potentials and calcium current in the mammalian circadian clock. *Nature* 416: 286-290.
23. Ingram CD, Snowball RK, Mihai R (1996) Circadian rhythm of neuronal activity in suprachiasmatic nucleus slices from the vasopressin-deficient Brattleboro rat. *Neuroscience* 75: 635-641.
24. Mihai R, Juss TS, Ingram CD (1994) Suppression of suprachiasmatic nucleus neurone activity with a vasopressin receptor antagonist: possible role for endogenous vasopressin in circadian activity cycles in vitro. *Neurosci Lett* 179: 95-99.
25. Haspel JA, Chettimada S, Shaik RS, Chu JH, Raby BA, et al. (2014) Circadian rhythm reprogramming during lung inflammation. *Nat Commun* 5: 4753.
26. Gale JE, Cox HI, Qian J, Block GD, Colwell CS, et al. (2011) Disruption of circadian rhythms accelerates development of diabetes through pancreatic beta-cell loss and dysfunction. *J Biol Rhythms* 26: 423-433.
27. Marcheva B, Ramsey KM, Buhr ED, Kobayashi Y, Su H, et al. (2010) Disruption of the clock components CLOCK and BMAL1 leads to hypoinsulinaemia and diabetes. *Nature* 466: 627-631.
28. Zvonic S, Ptitsyn AA, Conrad SA, Scott LK, Floyd ZE, et al. (2006) Characterization of peripheral circadian clocks in adipose tissues. *Diabetes* 55: 962-970.
29. Chung S, Son GH, Kim K (2011) Circadian rhythm of adrenal glucocorticoid: its regulation and clinical implications. *Biochim Biophys Acta* 1812: 581-591.
30. Son GH, Chung S, Choe HK, Kim HD, Baik SM, et al. (2008) Adrenal peripheral clock controls the autonomous circadian rhythm of glucocorticoid by causing rhythmic steroid production. *Proc Natl Acad Sci U S A* 105: 20970-20975.
31. Sellix MT (2015) Circadian clock function in the mammalian ovary. *J Biol Rhythms* 30: 7-19.
32. Yoshikawa T, Sellix M, Pezuk P, Menaker M (2009) Timing of the ovarian circadian clock is regulated by gonadotropins. *Endocrinology* 150: 4338-4347.
33. Fuller PM, Lu J, Saper CB (2008) Differential rescue of light- and food-entrainable circadian rhythms. *Science* 320: 1074-1077.
34. Mistlberger RE (2011) Neurobiology of food anticipatory circadian rhythms. *Physiol Behav* 104: 535-545.
35. Stephan FK (2002) The “other” circadian system: food as a Zeitgeber. *J Biol Rhythms* 17: 284-292.
36. Granados-Fuentes D, Ben-Josef G, Perry G, Wilson DA, Sullivan-Wilson A, et al. (2011) Daily rhythms in olfactory discrimination depend on clock genes but not the suprachiasmatic nucleus. *J Biol Rhythms* 26: 552-560.
37. Miller JE, Granados-Fuentes D, Wang T, Marpegan L, Holy TE, et al. (2014) Vasoactive intestinal polypeptide mediates circadian rhythms in mammalian olfactory bulb and olfaction. *J Neurosci* 34: 6040-6046.
38. Feillet CA, Mendoza J, Albrecht U, Pévet P, Challet E (2008) Forebrain oscillators ticking with different clock hands. *Mol Cell Neurosci* 37: 209-221.
39. Lamont EW, Robinson B, Stewart J, Amir S (2005) The central and basolateral nuclei of the amygdala exhibit opposite diurnal rhythms of expression of the clock protein Period. *Proc Natl Acad Sci U S A* 102: 4180-4184.

40. Nishikawa Y, Shibata S, Watanabe S (1995) Circadian changes in long-term potentiation of rat suprachiasmatic field potentials elicited by optic nerve stimulation in vitro. *Brain Res* 695: 158-162.
41. Chaudhury D, Wang LM, Colwell CS (2005) Circadian regulation of hippocampal long-term potentiation. *J Biol Rhythms* 20: 225-236.
42. Raghavan AV, Horowitz JM, Fuller CA (1999) Diurnal modulation of long-term potentiation in the hamster hippocampal slice. *Brain Res* 833: 311-314.
43. Moura PJ, Gimenes-Júnior JA, Valentinuzzi VS, Xavier GF (2009) Circadian phase and intertrial interval interfere with social recognition memory. *Physiol Behav* 96: 51-56.
44. Valentinuzzi VS, Menna-Barreto L, Xavier GF (2004) Effect of circadian phase on performance of rats in the Morris water maze task. *J Biol Rhythms* 19: 312-324.
45. Chaudhury D, Colwell CS (2002) Circadian modulation of learning and memory in fear-conditioned mice. *Behav Brain Res* 133: 95-108.
46. Decker S, McConnaughey S, Page TL (2007) Circadian regulation of insect olfactory learning. *Proc Natl Acad Sci U S A* 104: 15905-15910.
47. Gritton HJ, Kantorowski A, Sarter M, Lee TM (2012) Bidirectional interactions between circadian entrainment and cognitive performance. *Learn Mem* 19: 126-141.
48. Lyons LC, Rawashdeh O, Katzoff A, Susswein AJ, Eskin A (2005) Circadian modulation of complex learning in diurnal and nocturnal *Aplysia*. *Proc Natl Acad Sci U S A* 102: 12589-12594.
49. Valentinuzzi VS, Neto SP, Carneiro BT, Santana KS, Araújo JE, et al. (2008) Memory for time of training modulates performance on a place conditioning task in marmosets. *Neurobiol Learn Mem* 89: 604-607.
50. Ruby NE, Hwang CE, Wessells C, Fernandez F, Zhang P, et al. (2008) Hippocampal-dependent learning requires a functional circadian system. *Proc Natl Acad Sci U S A* 105: 15593-15598.
51. Kohara K, Pignatelli M, Rivest AJ, Jung HY, Kitamura T, et al. (2014) Cell type-specific genetic and optogenetic tools reveal hippocampal CA2 circuits. *Nat Neurosci* 17: 269-279.
52. Redondo RL, Kim J, Arons AL, Ramirez S, Liu X, et al. (2014) Bidirectional switch of the valence associated with a hippocampal contextual memory engram. *Nature* 513: 426-430.
53. Krout KE, Kawano J, Mettenleiter TC, Loewy AD (2002) CNS inputs to the suprachiasmatic nucleus of the rat. *Neuroscience* 110: 73-92.
54. Moga MM, Weis RP, Moore RY (1995) Efferent projections of the paraventricular thalamic nucleus in the rat. *J Comp Neurol* 359: 221-238.
55. Morin LP (2013) Neuroanatomy of the extended circadian rhythm system. *Exp Neurol* 243: 4-20.
56. Markov D, Goldman M (2006) Normal sleep and circadian rhythms: neurobiologic mechanisms underlying sleep and wakefulness. *Psychiatr Clin North Am* 29: 841-853.
57. Berridge CW, Foote SL (1991) Effects of locus coeruleus activation on electroencephalographic activity in neocortex and hippocampus. *J Neurosci* 11: 3135-3145.
58. Bliss TV, Collingridge GL (1993) A synaptic model of memory: long-term potentiation in the hippocampus. *Nature* 361: 31-39.
59. Shors TJ, Matzel LD (1997) Long-term potentiation: what's learning got to do with it? *Behav Brain Sci* 20: 597-614.
60. El-Sherif Y, Tesoriero J, Hogan MV, Wieraszko A (2003) Melatonin regulates neuronal plasticity in the hippocampus. *J Neurosci Res* 72: 454-460.
61. Wang LM, Suthana NA, Chaudhury D, Weaver DR, Colwell CS (2005) Melatonin inhibits hippocampal long-term potentiation. *Eur J Neurosci* 22: 2231-2237.
62. Takahashi Y, Sawa K, Okada T (2013) The diurnal variation of performance of the novel location recognition task in male rats. *Behav Brain Res* 256: 488-493.
63. O'Neal-Moffitt G, Pilli J, Kumar SS, Olcese J (2014) Genetic deletion of MT₁/MT₂, melatonin receptors enhances murine cognitive and motor performance. *Neuroscience* 277: 506-521.
64. Liston C, Cichon JM, Jeanneteau F, Jia Z, Chao MV, et al. (2013) Circadian glucocorticoid oscillations promote learning-dependent synapse formation and maintenance. *Nat Neurosci* 16: 698-705.
65. Kraus BJ, Robinson RJ, White JA, Eichenbaum H, Hasselmo ME (2013) Hippocampal "time cells": time versus path integration. *Neuron* 78: 1090-1101.
66. MacDonald CJ, Lepage KQ, Eden UT, Eichenbaum H (2011) Hippocampal "time cells" bridge the gap in memory for discontinuous events. *Neuron* 71: 737-749.
67. Cain SW, Chou T, Ralph MR (2004) Circadian modulation of performance on an aversion-based place learning task in hamsters. *Behav Brain Res* 150: 201-205.
68. Cain SW, McDonald RJ, Ralph MR (2008) Time stamp in conditioned place avoidance can be set to different circadian phases. *Neurobiol Learn Mem* 89: 591-594.
69. Cain SW, Ralph MR (2009) Circadian modulation of conditioned place avoidance in hamsters does not require the suprachiasmatic nucleus. *Neurobiol Learn Mem* 91: 81-84.
70. Atherton LA, Dupret D, Mellor JR (2015) Memory trace replay: the shaping of memory consolidation by neuromodulation. *Trends Neurosci* 38: 560-570.
71. Bendor D, Wilson MA (2012) Biasing the content of hippocampal replay during sleep. *Nat Neurosci* 15: 1439-1444.
72. Carr ME, Jadhav SP, Frank LM (2011) Hippocampal replay in the awake state: a potential substrate for memory consolidation and retrieval. *Nat Neurosci* 14: 147-153.