

## Wide Hybridization and Embryo-Rescue for Crop Improvement in *Capsicum*

C Debbarama, VK Khanna, W Tyagi, M Rai and NT Meetei

College of Post-Graduate Studies, CAU, Umiam-793103, Meghalaya, India

### Abstract

Wide-hybridization is an important tool in the hands of the plant breeder and a cytogeneticist. It is the first step to transfer genes of the wild species into the cultivated ones. In Meghalaya, chilli is the third most important spice crop after ginger and turmeric. However, diseases namely, tobacco mosaic virus, root rot, tomato spotted wilt virus, etc. lead to a considerable decline in yield. *Capsicum chinense*, *C. annuum* and *C. frutescens* were crossed with each other and embryo rescue was done between 27-33 days after pollination. The highest percentage of embryo growth was observed in MS medium with 0.5 mg/l GA<sub>3</sub> and 0.05 mg/l NAA. Hybrid plants were obtained and their hybridity was confirmed using both morphological and RAPD markers.

**Keywords:** Interspecific-hybridization; *Capsicum* species; Embryo-rescue; RAPD

### Introduction

North-East India is one of the 12 mega bio-diversity hot spots in the world [1]. In vegetables at least 12 species of *Solanum* are consumed by the local people. Many wild relatives can also contribute as donors in the hybridization programme. Enormous diversities exist within *Capsicum* at the interspecific level and also in their landraces. Wide hybridization is an important tool in the hands of the plant breeder and a cytogeneticist. It is the first step to transfer desirable traits from one species into another.

Successful sexual hybridization involves a series of events including pollen germination, pollen tube growth, fertilization, embryo and endosperm development, and seed maturation. Stebbins [2] divided hybridization barriers into two broad groups, namely pre-fertilization and post-fertilization barriers. The pre-fertilization category includes those mechanisms which prevent fertilization and includes geographical isolation, apomixes, and pollen-pistil incompatibilities. Post-fertilization barriers are a greater hindrance to hybridization and can be a result of ploidy differences, chromosome elimination, seed dormancy and hybrid breakdown of particular importance is the need to get hybrids that are fertile.

To bypass pre-zygotic barriers, the following can be done [3]. Where premature flower abscission takes place, hormone application may prolong the life of the flower. To increase pollen germination, one can apply boric acid, sucrose or abstract of the compatible anthers. We can amputate the stigma and apply sucrose-gelatin paste. One can apply self and other species pollen or bud pollination can be tried. To speed up pollen tube growth, application of growth regulators e.g., IAA, 2,4-D or GA<sub>3</sub> or use of radiation or an immunosuppressant like alpha-amino caproic acid as in *Vigna radiate* x *Vigna umbellate* can help. If there is stylar incompatibility, then one can shorten the style as in maize into *Tripsacum* cross or we can manipulate the ploidy levels of the parents. The use of intra-ovarian pollination has also been used to effect pollination by bypassing the style. This technique which involves injecting a suspension of pollen grains directly into the ovary was used with *Papaver somniferum* to produce normal fruits *in vivo* which contained viable seeds.

To bypass post-fertilization barriers, embryo-rescue, ovule culture and manipulations with protoplasts have been successfully used.

### Interspecific hybridization and embryo rescue in *Capsicum*

Chilli belongs to the genus *Capsicum*, which is among the world's extensively grown spice crops. India ranks first with an average yield of 1.6 mt ha<sup>-1</sup> from the total cultivated area of 7.67 lakhs ha. In Meghalaya, it is the third most important spice crops after ginger and turmeric with an area of 1900 hectares and a production of 2300 tones. However, diseases namely, Tobacco Mosaic Virus (TMV), root rot, Tomato Spotted Wilt Virus (TSWV), etc. leads to a considerable decline in yield. It is therefore necessary to transfer the important genes for various characters from one species to another. But there has been very little success due to problems in crossability. According to Shoemaker [4], in *Capsicum annuum* x *C. frutescens*, in general about two per cent seeds were viable when *C. frutescens* was used as the female parent. The F<sub>1</sub> plants ranged from completely pollen sterile to pollen fertile. In *C. annuum* x *C. chinense* crosses are much easier when *C. annuum* is used as the female parent. Only an occasional successful cross was obtained in the other direction. The F<sub>1</sub> were mostly pollen sterile. Similar results were obtained in the crosses between *C. frutescens* and *C. chinense*. The TMV genes have been transferred from *C. chinensis* to *C. annuum*. Resistance for fruit rot of chilli pepper caused by anthracnose was identified in *C. baccatum* and *C. annuum* by Pae et al. [5] but unfortunately, these species cannot be crossed easily. *C. chinense*, *C. baccatum*, *C. frutescens* and *C. pubescence* are used as genetic resources for disease resistance genes [6].

The present investigation was carried out on three species viz., *Capsicum annuum*, *Capsicum chinense*, and *Capsicum frutescens*, to determine whether this low success in crossability is due to pre-fertilization problems and to find out the optimum timing for embryo rescue. Hybrid plants were obtained and their hybridity was confirmed using both morphological and molecular markers (RAPD).

**\*Corresponding author:** V. K. Khanna, College of Post-Graduate Studies, CAU, Umia793103, Meghalaya, India, E-mail: [khannavk@rediff.com](mailto:khannavk@rediff.com)

**Received** December 06, 2012; **Accepted** December 25, 2012; **Published** December 31, 2012

**Citation:** Debbarama C, Khanna VK, Tyagi W, Rai M, Meetei NT (2013) Wide Hybridization and Embryo-Rescue for Crop Improvement in *Capsicum*. Agrotechnol S11: 003. doi:10.4172/2168-9881.S11-003

**Copyright:** © 2013 Debbarama C, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

## Material and Methods

The experimental material used in the present study comprised of three species including two varieties and two accessions of *Chilli*, respectively i.e. *Capsicum annuum* namely Kashi Anmol and Pusa jwala and *C. chinense* (COO-304) and *C. frutescens* (COO-904).

Experimental strategies were developed by adopting various standard and self developed techniques. They were suitably modified as per the need of the experiment. The complete course of the investigations was divided into the following parts.

### A. Crossability studies

1. Selfing and inter-specific crosses
2. Pollen viability, pollen germination, pollen tube growth and fruit set.

### B. Correlation studies for various characters in inter-specific crosses of *Capsicum*

#### C. *In vitro* studies: embryo rescue

1. Effect of the age of the hybrid embryos, on development, when cultured on media
2. Effect of media

### D. Confirmation of hybridity by morphological and molecular markers

## Results and Discussion

### Viable pollen

The average percentage of viable pollen showed little differences among the different species (Figure 1). There seems to be no correlation between viable pollen with the fruit set. Pollen adhesion on the stigmatic surface involves both impaction and the subsequent formation of attachment bonds [7]. According to Clarke et al. [8] on initial contact, the adhesive components could be contributed by either or both the partners, to enhance mutual adhesion. The specificity needed for foreign pollen discrimination might be provided by adhesive base or the thickening agents, both high molecular weight components (protein and carbohydrates) (Figure1).

### Pollen germination

In the present study there was regular increase in percent pollen germination after 10 hours, 20 hours and 30 hours in both selfing and in reciprocal crosses. After 30 hours pollen germination was more in

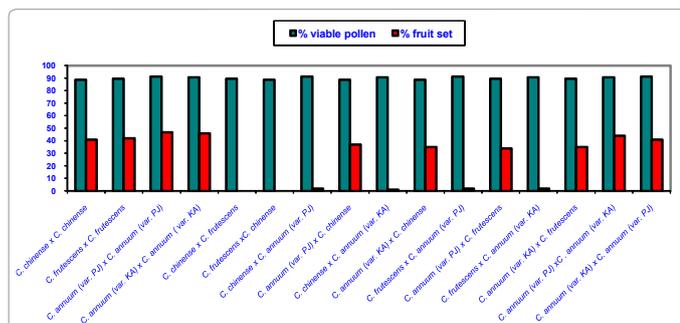


Figure 1: Percent viable pollen and percent fruit set on selfing and reciprocal crosses in different species of *Capsicum*.

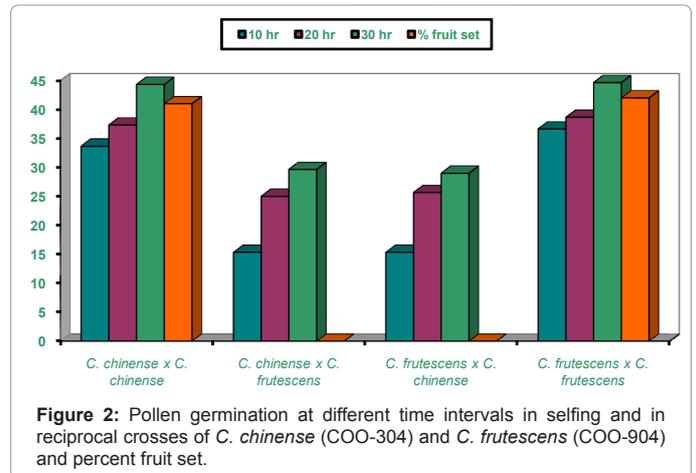


Figure 2: Pollen germination at different time intervals in selfing and in reciprocal crosses of *C. chinense* (COO-304) and *C. frutescens* (COO-904) and percent fruit set.

selfings than in crosses (Figure 2). Percent pollen germination after 30 hours of pollination showed significant positive correlation with fruit set. When *C. annuum* was used as the female, higher percent fruit set was recorded than in *C. frutescens* and *C. Chinense* where the fruit set was exceptionally as low as 1-2 percent.

Rehydration of mature pollen grains is an essential prelude to germination [6]. Since the water potential of the dry grain is lower than that of the stigma surfaces, it is readily apparent that the water of hydration will move in the direction of decreasing water potential from the stigma cell to the pollen grain. In a natural pollination, hydration is from a dry stigma. The water uptake, although rapid, is thus controlled and probably self regulatory [9,10]. Unregulated hydration leads to the bursting of the grain. However, tube growth in the secondary stigma branch is accompanied by shrinkage of hydrated grain, indicating that water is abstracted from the grain during the early extension. This conclusion provides the explanation that when there is competition between tubes some may fail to enter and will then continue growth on the surface of the branch or even in the air [11]. The rapid transition of the male gametophyte state of comparative inactivity in the grain to vigorous growth for the germination of pollen implies rapid re-establishment of a normal metabolism, including notably the synthetic capacity required for the synthesis of the pollen tube wall. The activation of pollen grain depends on rehydration which depends on the inflow of water from the stigma after attachment of the grain. Hydration in the suitable medium leads to the extrusion of the intine of the aperture [10]. In the viable grains azonation is quickly developed, generally within 5 minutes from the beginning of hydration (Figure 2).

### Pollen tube growth

Pollen tube growth increased with the passage of time (Figure 3). At 10 hours of pollination a number of pollen grains had germinated and the tubes had penetrated into the stigma hairs. Some of the pollen tubes had just started their growth in the hairs and a few had already entered the transmitting tissue of the style. After 20 hours of pollination there was a greater differentiation and elongation in the pollen tubes whereas some had just started to elongate and all intermediate stages were observed. After 30 hours of pollination, very long pollen tubes were recorded and since the styles were quite long, the pollen tubes could not be traced up to the base of the style as their staining became faint as they worked their way towards the ovary (Figure 3).

Fruit set showed a highly significant positive correlation with pollen tube growth (0.885) (Table 1).



Media	Casein hydroly-sate (mg/l)	Yeast extract (mg/l)	GA3 (mg/l)	NAA (mg/l)	Number of embryos cultured	Number of embryos showing growth	Embryos showing growth (%)
MS-1	500	500	0.25	0.025	20	0.0	0.0
MS-2	500	500	0.25	0.050	20	0.0	0.0
MS-3	500	500	0.25	0.100	20	0.0	0.0
MS-4	500	500	0.50	0.025	20	2.0	10.0
MS-5	500	500	0.50	0.050	20	16.0	84.0
MS-6	500	500	0.50	0.100	20	3.0	15.0
MS-7	500	500	0.75	0.025	20	0.0	0.0
MS-8	500	500	0.75	0.050	20	0.0	0.0
MS-9	500	500	0.75	0.100	20	0.0	0.0

- i. MS- 1, 2, 3, 7, 8 and 9 not showing any signs of embryo formation.
- ii. MS- 4 and 6 shows embryo formation but growth inhibited/growth improved; occasional sparse proliferation.
- iii. MS- 5 showed healthy plantlets; frequent formation of multiple roots (plate 2,3,4,5,6).

**Table 3:** Effect of media composition on embryo formation.



**Plate 1:** Immature seed, ovary and embryo 30 days after pollination in *C. chinense* x *C. annuum* (Kashi Anmol).



**Plate 2:** 28 days old plantlets of *C. chinense* x *C. annuum* (Kashi Anmol) after embryo rescue in culture tube.

by embryo culture [17], ovule culture [18] and plant regeneration from ovule-derived callus [19]. Secondly, normal seed development has been obtained by adjusting the environment and crossing factors during the crossing period and fruit growth [20], pollination with gamma-ray irradiated pollen grains [21], use of polyploidy [22], bridge crossing [23] and the selection of self-compatible species [24]. However, the plants obtained are very few (plate 1) (Table 2).

### Hybridity test by morphological markers

When the morphological parameters of the parents and the hybrids were compared, it was seen that there were clear cut differences in the height or fruit shape (Plates 2-6) of the hybrids as compared to the parents. The method of hybrid identification based on morphological characters can be used but these are influenced by environmental

factors and frequently lack the resolving power to identify hybrids at the juvenile stages, so one has to wait for the plants to reach maturity.

### Hybridity Test by RAPD markers

A total of 8 10-mer primers were used to screen polymorphism between the two pairs of parents. Out of the primers tested, all 8 produced clear amplification products. On an average each primer amplified 6.62 scorable bands (Table 4). To test the conformity of hybrid progeny, one must be able to distinguish, within a random sample of plants, those resulting from crossing the female and male parents obtained through embryo rescue. To check both possibilities with our markers, a first selection was done among the 8 clear primers and 16 (30.2%) showed polymorphism between the parents. Data on those 8 primers were used to select further for the presence of bands specific to the male parent of each hybrid. Only those primers which amplified bands specific to the male parent might reveal a proper pattern of a true hybrid as opposed to that of a selfed seed of the female parent. In the latter case, the band pattern would reflect the absence of the male band and the presence



**Plate 3:** F<sub>1</sub> of *C. chinense* x *C. annuum* (Kashi Anmol) obtained through embryo rescue.



**Plate 4:** F<sub>1</sub> of *C. frutescens* x *C. annuum* (Kashi Anmol) obtained through embryo rescue.



**Plate 5:** Fruits of parents and the hybrid.



**Plate 6:** Fruits of parents and the hybrid.

of the female band. The reproducibility of the pattern was verified: the primers that generated bands specific to the male parent were repeated at least two more times with the male, the female and the hybrid. Those primers giving the same pattern in the two replicates were chosen and were thereafter called RAPD markers. The rest were discarded. It was found that five primers were useful in determining the two hybrids. The efficiency to find a RAPD marker useful for hybrid determination was about 8%. This value is similar to that found for tomato hybrids, another Solanaceous species, in which 13 primers showed good polymorphism between parents out of 160 primers tested [25] and the value also was similar to that found for hybrid seed purity analysis in which 53 primers showed good polymorphism between parents out of 100 primers tested [26]. In our study, these primers (8) generated 10 RAPD markers (Table 4). Six primers (OPE-02, OPG-19, OPP-08, OPV-12, OPZ-04 and OPZ-06) gave one marker and the other two (OPQ-07, OPV-06) showed two and three useful bands, respectively. On the other hand, two of the primers (OPV-12, OPZ-4) were good to confirm the hybridity. In total, the number of RAPD markers useful for checking hybridity of the two *Capsicum* embryo rescued hybrids ranged from zero (OPQ-7) to four (OPV-12).

The development of improved cultivars through hybridization has made a major contribution to increased productivity and quality of plants in different crop plants. Hybridization of genetically different parents is followed for hybrid cultivar development and molecular marker techniques are often used for fastening plant improvement [27]. One of the problems faced today by the breeders is the difficulty in identifying true hybrids from the crossed progenies before planting. Traditional method of hybrid identification based on morphological characters is influenced by environmental factors and lacks the resolving power to identify hybrids at juvenile stage, which makes it necessary to grow the plants up to maturity to confirm hybridity. Molecular markers used to detect DNA polymorphism are the most direct answer to it. Molecular analysis of genomes can be made with RAPD approach, where PCR allows exploration of large genomic portions.

Out of the 35 decamer primers used in RAPD analysis, 13 primers, viz., OPE-01, OPE-02, OPE-05, OPE-08, OPE-11, OPE-13, OPE-16, OPE-17, OPE-18, OPE-19, OPE-20 and OPC-16 yielded the best amplification products [28]. Amplified products were scored on the basis of presence or absence of bands. RAPD banding pattern among parents and their hybrid population was compared to assess hybridity at the DNA level. Non-parental bands observed in some progenies might have resulted from DNA recombination or mutation. Chromosomal crossing-over during meiosis may have resulted in the loss of priming sites and thus markers are present in parents but not in offspring.

Black pepper being heterozygous and propagated through cuttings, segregation of characters can be expected in the hybrids progenies, which makes the bands from parents to be absent in hybrids. RAPD marker technique has potential application in the identification, registration and protection of pepper accessions. It will also help in screening of duplicates, assessing genetic diversity and monitoring the genetic stability of conserved germplasm, which has shown successful results in genus *Piper*.

Black pepper being heterozygous, markers diagnostic of each male parent is to be determined for each cross. Confirmation of hybrid nature of seedlings at the juvenile stage by screening with RAPD markers would be practical and of economic significance in a perennial crop like black pepper. It will enable elimination of all doubtful seedlings and can save labor, space and cost. RAPD can also be applied to broad array

of cultivars and wild accessions for precise determination of genetic diversity within genus *Capsicum*.

Interspecific hybridization is essential to introgress resistance genes from *Capsicum baccatum*, a related species of cultivated pepper (*C. annuum*), since reliable genetic resources resistant to anthracnose have recently been identified within the *C. baccatum* germplasm but unfortunately in our case the seeds did not germinate.

To overcome post-fertilization barriers we studied the regeneration potential by taking immature crossed embryos as explants and succeeded in getting plantlets. We could confirm the hybridity by morphological and RAPD markers. One can study further by screening for the genes for resistance to diseases, etc. and make use of these hybrids for crossing programs and develop high yielding, disease resistant varieties.

### Confirmation of hybridity by morphological and molecular markers

Sl. No.	Primer Code	Total number of bands	Number of polymorphic bands	(Polymorphic bands/ Total bands)x100
1	OPE-2	7	4	57.14
2	OPG-19	5	2	40.00
3	OPP-8	5	1	20.00
4	OPQ-7	7	0	00.00
5	OPV-12	9	4	44.44
6	OPV-6	5	0	00.00
7	OPZ-4	6	2	33.33
8	OPZ-6	9	3	33.33
<b>Total</b>		<b>53</b>	<b>16</b>	<b>228.24</b>
<b>Average</b>		<b>6.62</b>	<b>2</b>	<b>30.21</b>

Table 4: Total number of RAPD loci detected using 8 RAPD primers.

### Conclusion

There were little differences in the average pollen viability among the species but showed huge differences in fruit set in reciprocal crosses. On selfing there was more pollen germination percentage compared to the crosses. Pollen germination had significant positive correlation with fruit set. Fruit set was high on selfing as compared to reciprocal crosses. Maximum fruit set was recorded on selfing of *C. annuum*, variety Pusa jwala (47 per cent). Both the species (*C. frutescens* and *C. chinense*) responded to embryo germination by MS+Yeast extract+casein hydrolysate+GA<sub>3</sub> at 0.5 mg/l+NAA at 0.05 mg/l. NAA (0.025 mg/l and 0.10 mg/l) showed poor percent of embryo germination. Average number of plantlets was from the medium supplemented with 0.5 mg/l GA<sub>3</sub> and NAA 0.05 mg/l. Primers which amplified bands in the putative hybrids and were specific to the male parent confirmed the hybridity.

### Acknowledgments

The authors would like to acknowledge funding received from College of PG Studies, Central Agricultural University (CAU), Imphal, Manipur for conducting the experiments. Mr. C. Debbarama was supported by funding from State Agricultural Department, Tripura. This work is part of MSc. Thesis.

### References

- Hore DK, Nagachan SV, Mishra A, Kadirvel G, Das A, et al. (2010) Important plant genetic resources of North Eastern India: their sustainable utilization and conservation. In: Conservation of Natural Resources for Sustainable Hill Agriculture. ICAR Res pp: 85-92. Complex for NEHR, Umiam, Meghalaya.
- Stebbins GL (1950) Variation and evolution in plants. Columbia Univ. Press, New York.
- Srivastava M, Eidelman O, Torosyan Y, Jozwik C, Mannon RB, et al. (2011)

- Elevated expression levels of ANXA11, integrins  $\beta$ 3 and  $\beta$ 3, and TNF- $\alpha$  contribute to a candidate proteomic signature in urine for kidney allograft rejection. *Proteomics Clin Appl* 5: 311-321.
4. Shoemaker JS (1955) Small fruit culture. 3rd edn. Mc Graw- Hill, London.
  5. Pae D, Cho M, Jung S (1995) Breeding for resistance in chilli pepper. NHRI Annual Report Pp 19-28.
  6. Caranta C, Pflieger S, Lefebvre V, Daubèze AM, Thabuis A, et al. (2002) QTLs involved in the restriction of cucumber mosaic virus (CMV) long-distance movement in pepper. *Theor Appl Genet* 104: 586-591.
  7. Dumas C, Gaude T (1981) Stigma- pollen recognition: a new look. In : Int Symp Adv Plant cytoembryology, *Acta Soc Bot Pol* 50: 235-247.
  8. Clarke A, Gleeson P, Harrison S, Knox RB (1979) Pollen-stigma interactions: Identification and characterization of surface components with recognition potential. *Proc Natl Acad Sci U S A* 76: 3358-3362.
  9. Heslop-Harrison J (1978) Recognition and response in the pollen-stigma interaction. *Symp Soc Exp Biol* 32: 121-138.
  10. Heslop HJ (1979) Aspects of the structure, cytochemistry and germination of the pollen of rye (*Secale cereale* L.) *Adv Bot* 44: 47.
  11. Heslop HJ, Heslop HY (1981) The pollen-stigma interaction in the grasses. *Acta Botanica Neerlandica* 30: 289-307.
  12. AVRDC (1986) AVRDC adds pepper as new principal crop. *Centre point* 5: 1-3.
  13. Erickson AN, Markhart AH (1999) Mechanisms of fruit set reduction in *Capsicum annuum* at high temperatures. American Society of Plant Biologists.
  14. Kodali S, Khanna VK (1994) Standardization of the best timing of growth hormone application in wheat-barley crosses to increase seed set. *Cer Res Comm* 22: 309-312.
  15. Raghavan V (1976) Experimental Embryogenesis in Vascular Plants. New York Academic Press.
  16. Hossain MA, Minami M, Nemoto K (2003) Immature Embryo Culture and Interspecific Hybridization between *Capsicum annuum* L. and *C. frutescens* L. via Embryo Rescue. *Japanese Journal of Tropical Agriculture* 47: 9-16.
  17. Fienup DM, Hamelin J, Reyes-Giordano K, Falcomata TS (2011) College-level instruction: derived relations and programmed instruction. *J Appl Behav Anal* 44: 413-416.
  18. Imanichi S, Watanabe Y, Hiura, I (1985) A simple and efficient method for the interspecific hybridization between *Lycopersicon esculentum* and *L. peruvianum*. *J. Yamagata. Agr For Soc* 42: 13-15.
  19. Thomas BR, Pratt D (1981). Efficient hybridization between *L. esculentum* and *L. peruvianum* via embryo callus. *Theor Appl Genet* 59: 215-219.
  20. Kuriyama T, Kuniyasu K, Mochizuki H (1971) Studies on the breeding of disease- resistant tomato by interspecific hybridization. Influence of the environment and crossing factors on the frequency of interspecific hybrids. *Bull Hort Res Station* 10: 51-90.
  21. Yamakawa K (1971) Effect of chronic gamma radiation on hybridization between *Lycopersicon esculentum* and *L. peruvianum*. *Gamma Field Symposia* 10: 11-30.
  22. Kuriyama T, Mochizuki H (1971) Studies on the breeding of disease- resistant tomato by interspecific hybridization. I Fertility and disease resistance of the progenies of interspecific hybridization. *Bull Hort Res Station* 11: 33-60.
  23. Poysa V (1990) The development of bridge lines for interspecific gene transfer between *Lycopersicon esculentum* and *L. peruvianum*. *Theor Appl Genet* 79: 187-192.
  24. Hogenboom NG (1972) Breaking breeding barriers in *Lycopersicon*. The genus *Lycopersicon*, its breeding barriers and the importance of breaking these barriers. *Euphytica* 21: 221-227.
  25. Paran I, Aftergoot E, Shiffriss C (1998) Variation in *Capsicum annuum* revealed by RAPD and AFLP markers. *Euphytica* 99: 167-173.
  26. Ballester MJ, Vicente, MC (1998) Determination of F1 hybrid seed purity in pepper using PCR-based markers. *Euphytica* 103: 223-226.
  27. Winter P, Kahl G (1995) Molecular marker technologies for plant improvement. *World J Microbiol and Biotech* 11: 438-448.
  28. George KJ, Ganga G, Varma RS, Sasikumar B, Saji KV (2005) Identification of hybrids in black pepper (*Piper nigrum* L.) using male parent specific RAPD markers. *Current Science* 88: 216-218.