Concentration of the CS-846 Epitope in Serum and Synovial Fluid of Horses with Different Grades of Osteochondral Fragments in the Carpal Joints

Hipolito Chavez1, Hugo Folch2, Oscar Araya3, Benjamin Uberti4 and Gabriel Moran4*

1Faculty of Veterinary Medicine, Universidad San Sebastian, Concepción, Chile
2Department of Immunology, Faculty of Medicine, Universidad Austral de Chile, Valdivia, Chile
3Department of Clinical Veterinary Sciences, Faculty of Veterinary Sciences, Universidad Austral de Chile, Valdivia
4Department of Pharmacology, Faculty of Veterinary Sciences, Universidad Austral de Chile, Valdivia, Chile

Corresponding author: Gabriel Moran, Department of Pharmacology, Faculty of Veterinary Sciences, Universidad Austral de Chile, Valdivia, Chile, Tel: +56-63-221936; Fax: +56-63-221444; E-mail: gmoran@uach.cl

Rec date: March 9, 2016; Acc date: May 18, 2016; Pub date: May 24, 2016

Copyright: © 2016 Hipolito C, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

The aim of this study was to determine serum and synovial concentrations of the CS-846 epitope of articular cartilage aggrecan, in horses with a radiological diagnosis of osteochondral fragmentation (OF). For this purpose, 20 thoroughbred horses with unilateral radiocarpal or intercarpal OF were used, and 10 clinically and radiologically healthy Thoroughbred horses were assigned to the control group. Serum and synovial concentrations of the CS-846 epitope were measured in both groups by means of ELISA. The concentration of the CS-846 epitope in synovial fluid was significantly higher in carpal joints with moderate articular damage (grade 2 OF) than in the control group or in horses with severe disease (grade 3 and 4 OF). In serum, non-statistical differences in the concentration of the CS-846 epitope were observed between horses with OF and controls. We conclude that the concentration of CS-846 epitope in synovial fluid suggests an increase in cartilage aggrecan synthesis, which may be associated with the presence of moderate clinical disease. The CS-846 epitope concentration in synovial fluid may be a useful biomarker for the study of carpal OF in horses, providing a measure of the balance between cartilage synthesis and degradation in this equine disease.

Keywords: CS-846; OF; Carpal joint; Horses

Introduction

Lameness is one of the most common and important causes of failure during training of the equine athlete: athletic activity entails repetitive overloading of joints and may lead to articular cartilage damage, often accompanied by synovitis, tendon and ligament lesions and progressive subchondral bone sclerosis [1,2]. Carpal osteochondral fragments, or chip fractures, are a common cause of lameness in race horses. Osteochondral fragments are pieces of osteochondral tissue (cartilage or subchondral bone) that detach from the edges of the bones in a synovial joint. This condition occurs with high incidence in the carpal joint, and it seems most prevalent in young racehorses from 2 to 4 years of age who are undergoing rigorous athletic training [3-5]. The distalarticular aspect of the radius bone, distal aspect of the radial carpal bone and proximal aspects of the intermediate and third carpal bones are common sites of OF; biomechanical loading during hyperextension of highly mobile joints explains the predominance of lesions on the dorsal articular surfaces [3,4,6-13]. Repetitive cyclic loading on the subchondral bone eventually leads to bone sclerosis and focal areas of microfracture, leading to collapse of the weakened tissue and fragmentation of cartilage and bone [14,15]. Such fragments may be a poor prognostic indicator for future athletic performance [15].

Articular cartilage is composed of chondrocytes embedded in an extracellular matrix of type II collagen fibrils. This matrix provides tensile strength and promotes the retention of proteoglycans (PG). Aggrecan is an important part of this matrix; it is a high molecular

weight proteoglycan which bestows compressive resistance to cartilaginous tissue [16]. Molecular markers of proteoglycan origin can be released into synovial fluid (SF) and serum, and may be useful for monitoring metabolic activity (anabolism or catabolism) of cartilage over time. For example, keratan sulfate increases in synovial fluid as a marker of articular cartilage catabolism [17]. The 846 epitope of chondroitin sulfate (CS-846) is another indicator of changes in cartilage glycosaminoglycan content, specifically of aggrecan turnover, and it can be measured in SF by immunoassay [18-21]. In humans, the release of CS-846, normally present in juvenile articular cartilage but almost completely absent from adult cartilage, is closely correlated with aggrecan synthesis [22]. A previous study in horses found that CS-846 epitope increases not only in SF but also in serum of joints with OF [19]. However, CS-846 expression has not been correlated with the varying severity of osteochondral fragmentation. We hypothesize that CS-486 concentration varies depending on the severity of osteochondral lesions in joints. Thus, the objective of the present study was to determine serum and synovial concentrations of CS-846 in horses with a radiological diagnosis and different grades of disease, compared to healthy horses.

Material and Methods

Horses

Thirty thoroughbred horses were included in this study (16 females and 14 males), with a mean age of 3 years (range 2-5 years old) and a mean weight of 428 kg (range 390-466 kg). Twenty animals had different degrees of carpal OF, and ten healthy animals represented the
control group. The time of pre-existence was not determined for this study.

Owner’s consent was obtained in all cases before inclusion in the study. All procedures were approved by the Bioethical Committee of the Universidad Austral de Chile.

**Clinical and radiographic examinations**

Physical and lameness examinations were performed on all horses before inclusion in the study. Localized lameness, heat, pain and effusion of the carpal joints was recorded in all cases. Stationary (standing) and dynamic (trotting) flexion tests were used to subjectively assess joint pain and synovial effusion. All included horses underwent carpal radiographic examinations, consisting of five radiographic views (dorsopalmar, lateromedial, dorsopalmar lateromedial oblique, dorsopalmar mediolateral oblique and flexed lateromedial) of each carpus were examined. Radiographic lesions were classified depending on the presence of osteochondral fragments and underlying bone lesions, in the following manner: grade 0: joint with no evidence of osteochondral fragmentation (n = 10); grade 1: minimal continuity loss of the dorsal bone surface (less than 5 mm) (n = 8); grade 2: 45% loss of the articular surface (n = 8); grade 3: 60% loss of the articular surface (n = 8); grade 4: significant subchondral loss with loss of support of the dorsal surface (n = 8).

**Synovial fluid and serum samples**

Each subject was sampled once. Horses were restrained manually during the collection of all samples, using a twitch when needed; sedation was not used. Whole blood samples were collected by jugular venipuncture into plain sterile tubes without anticoagulant. For serum extraction from whole blood, the tubes were stored vertically for 30 min at 37°C until centrifuged for 7 minutes at 650 × g. The serum was kept frozen at −70°C until used.

Samples of synovial fluid were obtained by arthrocentesis using 21G or 18G sterile needles. Samples were obtained from the radiocarpal or intercarpal joints, depending on the location of the osteochondral fragments in each case, using a dorsal or dorsolateral approach. Samples were collected in dry tubes, centrifuged at 690 × g for 30 minutes at room temperature to remove cells, and the supernatant was stored at −70°C until analyzed.

**Immunooassay of the CS-486 epitope**

The concentration of the CS-846 epitope in serum and synovial fluid was assessed by competitive ELISA. For this assay, we used a commercial kit (IBEX Pharmaceuticals Inc., Canada) using a mouse monoclonal anti-bovine aggrecan, which cross-reacts with equine aggrecan as described previously [19]. The serum and synovial fluid were diluted 1:5 and 1:30, respectively. The samples were added to each well of a microtiter plate that was pre-coated with goat anti-mouse immunoglobulin. Later, CS-846 epitope labeled with biotin was added, followed by a mouse IgM antibody specific for CS-846. A calibration curve was generated using a standard solution of known concentrations of bovine aggrecan. Streptavidin-peroxidase conjugate was added to each well, and then tetramethylbenzidine (TMB) substrate was added. The reaction was stopped with H$_2$SO$_4$ (0.2 M). Finally, the absorbance was measured in a spectrophotometer to obtain the optical density (OD), which is inversely proportional to the concentration of the epitope in the samples. The OD was converted to the concentration of CS-846 epitope (µg/ml) using the computer program GraphPadPrism 4.0 for non-linear calibration curves.

**Statistical analysis**

All statistical analyses were done with GraphPad Prism version 4.0 for Windows (GraphPad Software, La Jolla California USA). Data homoscedasticity was evaluated by Kolmogorov-Smirnov test. Differences between the mean values of the variables in horses with carpal OF and controls were evaluated by analysis of variance (ANOVA). When significant differences were obtained, we applied the method of Bonferroni-Dunn’s multiple comparison test. A value of P < 0.05 was considered statistically significant.

**Results**

All data was not normally distributed. The observed concentrations of the CS-846 epitope were significantly higher (P < 0.05) in synovial fluid than in serum, both in horses with carpal joint OF and in controls. The mean serum concentrations of the CS-846 epitope were 0.64 ± 0.50, 0.40 ± 0.33, 0.26 ± 0.33 and 0.60 ± 0.27 (µg/ml ± SD) for horses carrying carpal OF grades 1, 2, 3 and 4, respectively, and 0.17 ± 0.13 (µg/ml ± SD) for control horses. On the other hand, the SF concentrations of the CS-846 epitope were 3.80 ± 1.77, 5.05 ± 2.63, 1.98 ± 2.92, and 1.65 ± 0.79 (µg/ml ± SD) for horses with carpal OF grades 1, 2, 3 and 4, respectively, and 2.86 ± 0.51 (µg/ml ± SD) for control horses. In the synovial fluid of the radiocarpal or intercarpal joints, the concentrations of the CS-846 epitope in horses with grade 2 OF were significantly higher (Figure 1).

**Figure 1:** CS-846 epitope concentration in synovial fluid from horses with varying grades of osteochondral fragmentation (OF, grades 0–4) in radiocarpal or intercarpal joints. Each box plot is composed of five horizontal lines displaying the 10th, 25th, 50th (median), 75th and 90th percentiles of the epitope concentration (µg/ml) obtained from different groups of equines. A * indicates (P < 0.05).
the SF and serum have been observed in human arthropathies as well. Its concentration in the SF may indicate the rate of aggrecan synthesis because the CS-846 epitope is expressed only in new aggrecan molecules [18]. These same authors show that the CS-846 epitope is a good indicator of aggrecan synthesis, which suggests that this protein may be considered a marker of cartilage anabolic activity. Additionally, both the rate of release of the CS-846 epitope (on fragments of PG) and its concentration in the SF may indicate the rate of aggrecan synthesis [22]. In contrast, more severe joint damage (grade 4 OF) is associated with low biosynthetic activity by chondrocytes [23].

In horses with OF, the concentration of the CS-846 epitope was higher in the SF than in the serum. The results of this study are consistent with investigations of Frisbie et al. [19] in horses with carpal joint OF, which found higher concentrations of the CS-846 epitope in the SF than in the serum of the same animals. In turn, Matyas et al. [24], in an experimental model of osteoarthritis (OA) in dogs, also found significantly higher levels of the CS-846 epitope in the SF than in the serum. Differences in the concentration of the epitope between the SF and serum have been observed in human arthropathies as well. A study by Poole et al. [22] also found that the CS-846 epitope reached higher levels in the SF than in the serum of patients with OA or rheumatoid arthritis (RA). Thus, it seems clear that although the CS-846 epitope can occasionally be detected in blood/serum, it typically seems to remain exclusively in the synovial cavity. Indeed, the epitope is exclusive of specific cartilage PG aggrecans, and has not been found in tissues other than articular cartilage, with the sole exception of a study by Antoniou et al. [25] which identified this epitope in the nuclei of human intervertebral discs.

The current study found a significant degree of synovitis in horses with OF. Several mechanisms could explain the different CS-846 epitope concentrations found in the SF and serum of horses with carpal joint OF. Simkin et al. [26] suggest that the number of molecules from normal cartilage detected in the SF (including fragments of PG) is determined not only by the rate of release of these molecules but also by factors affecting the clearance of molecules from the synovial cavity. These factors include the volume of the SF in the joint space, the blood flow and inflammation of the synovial membrane, joint mobility and the composition of the synovial fluid, particularly its leukocyte content [27]. As mentioned above, Myers [27] suggests that joint inflammation could alter the clearance of biological markers from joint cavities. In a follow-up study, Myers et al. [28] found an accelerated clearance of albumin-1131 in a model of canine OA with moderate synovitis, indicating that the inflamed synovial membrane might favor the clearance of low molecules but not the large PG molecules expressing the CS-846 epitope. On the other hand, older studies by Page-Thomas et al. [29] in rabbits with experimental arthritis found no significant differences between PG clearance from the inflamed synovial cavity versus controls.

Our findings show an increased concentration of CS-846 epitope in SF of horses with moderate (grade 2) OF. This suggests either reduced clearance or augmented release from cartilage, as explained above, and may well be related to mechanical damage of chondrocytes. Studies by Elder et al. [30] indicate that chondrocytes are sensitive to compressive forces that accumulate over time. D’Lima et al. [31] found 47% apoptosis in cell cultures of bovine chondrocytes that received pressure loads of 23 megapascals (MPa) for 48 hours. If we assume that 7 MPa is the upper limit of physiological stress of cartilage [32], this might suggest that a stress load that causes osteochondral fracture, such as grade 4 OF, may be sufficient to induce apoptosis of these cells, which could significantly compromise the synthesis of PG. Other studies suggest that the dislocation of results in a lower loading of the cartilage, which could influence the synthetic activity of the chondrocytes [33]. Those authors showed that chondrocytes in the cartilage from the part of the fractured bone that continued to bear a load did not express cartilage oligomeric matrix protein (COMP) mRNA, which suggests that the reduced mechanical compression of the dislocated OF stimulates chondrocyte synthetic activity, possibly contributing to the up-regulation of the CS-846 epitope.

The low concentrations of the CS-846 epitope that were recorded in the SF of horses with grades 3 and 4 carpal OF may be associated with the marked inflammatory process occurring in these degrees of injury. Inflammation may inhibit the synthesis of aggrecan through excessive production of pro-inflammatory cytokines [34]. Chondrocytes express several chemokines and have receptors that enable responses associated with cartilage catabolism [35-37]. Both in vitro and in vivo studies indicate that interleukin-1 (IL-1) and tumor necrosis factor (TNF)-α are the predominant catabolic cytokines involved in the initiation and progression of articular cartilage destruction and that they have an adverse effect on the synthesis of aggrecan by chondrocytes [38,39]. In addition to inducing the synthesis of metalloproteinases and other proteinases by chondrocytes, IL-1 and TNF-α increase the synthesis of prostaglandin E2 (PGE2) by stimulating the expression or activities of COX-2, microsomal PGE synthase-1 (mPGES-1) and soluble phospholipase A2 (sPLA2). They also up-regulate the production of nitric oxide via inducible nitric oxide synthetase (iNOS, or NOS2), other proinflammatory cytokines, such as IL-6 [40], leukemia inhibitory factor (ILF), IL-17 and IL-18, and chemokines, such as IL-8 [38]. The extent of inflammation can vary widely depending on the amount of mechanical damage to chondrocytes, and thus, it seems that marked inflammation in severe clinical cases may actually lead to reduced synthesis of aggrecan and less CS-846 concentration in SF of affected joints.

In conclusion, our findings showed increased concentration of CS-846 epitope in synovial fluid of horses with moderate (grade 2),
sugesting an increase in cartilage aggrecan synthesis compared to healthy joints [41,42]. The lack of a significant increase of CS-846 epitope concentration in joint fluid of horses with more severe (grade 3 and 4) may suggest decreased aggrecan synthesis due to severe inflammation. The CS-846 epitope in synovial fluid may be a useful biomarker in the study of cartilage OF in horses. Further studies are needed to determine whether this epitope may provide a measure of the balance between cartilage synthesis and degradation in osteochondral disease in horses [43,44].

Acknowledgments

This work was supported by Graduate School, Faculty of Veterinary Science, Universidad Austral de Chile.

References


