

Enterococcus in Water, Sediment and Clams in a Tropical Environment, Maracaibo Lake, Venezuela

Marynes Montiel*, Ricardo Silva, Jesús Núñez, Félix Morales, Hector Severeyn and Yajaira García

University of Zulia, Maracaibo, Venezuela

Abstract

The genus *Enterococcus* has widespread extra enteric sources and reservoirs. It has been suggested that coastal and Great Lakes States adopt enterococci as an alternative indicator for the monitoring of recreational water quality. Limited information, however, is available about the presence of enterococci in Lake Maracaibo, which is an important estuary in Venezuela, with the income and interchange of the Caribbean Sea, and is used by the people for recreational purposes and the culture of marine organisms. In this study, the density and species composition of enterococci in sediment, clams and water were examined at Lake Maracaibo. Enterococci was enumerated by the Most Probable Number Technique (MPN), and isolated by standard methods. Results obtained by MPN analyses indicated that enterococci were present in all samples, and their densities were generally higher in clams than sediment and water with means of 1.0×10^5 MPN/100 g, 2.1×10^3 MPN/100 g, and 6.0×10^1 MPN/100 ml, respectively. Dominant *Enterococcus* species were *E. faecalis* (65%), *E. casseliflavus* (20%), *E. sanguinicola* (5%), *E. faecium* (5%), and unidentified strains (5%). Results suggest that some enterococci are able to persist in Lake Maracaibo, especially in clams and sediment, for a prolonged amount of time after being introduced.

Keywords: *Enterococcus*; Water sediment; Clams; Tropical environment

Introduction

Enterococci are spherical or ovoid cells arranged in pairs or chains, Gram positive non-spore-forming, obligately fermentative chemoorganotrophs. They are important members of gut communities in many animals, and opportunistic pathogens that cause millions of infections annually. They can be found in soil, water, and plants [1]. When they are found in water it is associated with fecal matter. In the last two decades, the genus *Enterococcus* has been believed to be one of the most common causative agents of nosocomial diseases in humans following staphylococci [2-4]. Additionally, some vancomycin-resistant enterococci belonging to the group of zoonoses (Slovak Regulation No.626/2004) can be transmitted to humans via food chain, and are employed as vectors in dissemination of resistance genes inside as well as outside of genus. This feature shifts them to the group of potential reservoirs of resistance genes with significant risk of resistance transfer to obligate pathogens, such as *Staphylococcus* spp. or *Listeria* spp. [5-7]. Also, *Enterococcus* has been proposed as fecal indicator in some countries, especially in recreational waters. The vast majority of clinical enterococcal infections in humans are caused by *Enterococcus faecalis* (around 80% of clinical isolates) and *Enterococcus faecium* (most of the remainder), with occasional infections being caused by *E. durans*, *E. gallinarum*, *E. casseliflavus*, *E. avium*, *E. hirae*, *E. mundtii*, and *E. raffinosus* [8,9]. The emergence of *Enterococcus faecalis* and *Enterococcus faecium* was paralleled by increases in glycopeptide and high-level aminoglycoside resistance, both important compounds for the treatment of human infections [10]. Although *Enterococcus faecalis* is the causative agent in most enterococcal infections, a partial replacement of *Enterococcus faecalis* by *Enterococcus faecium* has been noted in the last years, and presently, up to one-third of enterococcal infections in some countries is attributed to this species [11,12].

The presence of enterococci as an indicator of fecal contamination has been used in management of recreational water quality standards as it correlates best with the incidence of swimming-related illnesses [13,14].

Studies conducted by EPA to determine the correlation between

different bacterial indicators and the occurrence of digestive system illness at swimming beaches suggest that the best indicators of health risk from recreational water contact in fresh water are *E. coli* and enterococci. For salt water, enterococci are the best [15].

However, little is known about the behavior of FIB in tropical environments, particularly in estuaries, since most studies have been performed in temperate to cold locations [16].

Although some countries required enterococci test for recreational bacterial water quality standards, the requirement in Venezuela are only total and fecal coliforms [17]. Previous studies in Venezuela have been showed the presence of enterococci in the environment but not much is available about the presence of enterococci in Lake Maracaibo [18]. The objective of this research was to analyze and compare the density and species composition of enterococci in water, sediment and clams at Curarire beach, located at Lake Maracaibo, Venezuela.

Materials and Methods

Study area. Lake Maracaibo is a tropical lake and remains warm (28-32.5°C; mean 30°C) throughout the annual seasonal cycle; it is the largest lake in South America [19], located at Zulia State, Venezuela. This estuarine lake is connected to the Gulf of Venezuela via the Strait of Maracaibo and El Tablazo Bay, and is brackish due to intrusion of salty water from the strait [19]. Lake Maracaibo receives large loads of nutrients from tributaries, sewage discharges, and agricultural sources [20].

*Corresponding author: Marynes Montiel, University of Zulia, Maracaibo, Venezuela, Tel: 2617416786; Fax: 2617483724; E-mail: montielmarynes@gmail.com

Received September 04, 2013; Accepted September 23, 2013; Published September 28, 2013

Citation: Montiel M, Silva R, Núñez J, Morales F, Severeyn H, et al. (2013) *Enterococcus* in Water, Sediment and Clams in a Tropical Environment, Maracaibo Lake, Venezuela. J Marine Sci Res Dev 3: 133. doi: [10.4172/2155-9910.1000133](https://doi.org/10.4172/2155-9910.1000133)

Copyright: © 2013 Montiel M, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Samples collection

Samples were collected from three different points at Curarire Beach, during a period of a year (October 2008 to October 2009). Water, sediment and clams samples were, using a sterile 1 L screw-top bottle or sterile Whir-Pak bags (Nasco), respectively. Water samples were collected from 0.3 m below the surface of the water. Samples were transported in a cooler on ice packs to the laboratory and tested within 6 h of collection [21].

Samples analysis

The seafood and sediment samples were prepared for testing by blending 25 g in 225 mL of 0.1% sterile peptone water for 2 minutes [21,22].

Microbial density: Enterococci in water, sediment, and clams homogenate, were enumerated using the standard Most Probable Number Technique using Azide dextrose broth (Himedia, India) and KF-enterococcus agar (Himedia, India). Strains grown on KF-streptococcus agar were identified as *Enterococcus* with the following biochemical tests: Gram stain, catalase, growth with 6.5% of NaCl, growth at 45°C, bile esculine hydrolysis.

Species identification: Enterococcal strains were further identifies to the species level by using conventional physiological tests which are based on carbohydrate fermentation using 1% solution of the following sugars: manitol, sorbitol, sucrose, raffinose, and arabinose; by arginine decarboxilation in Moellers decarboxylase broth; tellurite; motility test, pigment production detected on tryptic soy agar (TSA) (Difco, USA) [23].

Physicochemical parameters: Temperature, pH, Turbidity and salinity were measures in the water using convectional techniques [21].

Results and Discussion

Water, sediment and clam samples were taken at Curarire Beach, Lake Maracaibo. The water temperature ranged between 23.0°C to 35.0°C, with a mean of 30.6°C; pH 8.74 to 10.5 (mean 9.42); salinity 2 PSU to 6 PSU (mean 3.85), and turbidity 20.7 NTU to 94.7 NTU (mean 44.48). *Enterococcus* densities in water, sediment and clams ranged from <2-50 MPN 100 mL⁻¹, <2-130 MPN 100 g⁻¹, and <2-70 MPN 100 g⁻¹, respectively with a geometric mean between 2 and 130 (Figure 1). *Enterococcus* prevalence rate was 71%. One of the water sample exceeded EPA guidelines, being the sample with the highest fecal coliform value (700 MPN 100 mL⁻¹) (data not shown). Previous studies have shown the highest value of fecal coliforms in shellfish *Rangiacuneata* evaluated from this environment exceeding the

Venezuelan guidelines [24].

Densities were generally higher in sediment than clams and water with means of 25.9 MPN/100 g, 11.4 MPN/100 g, and 5.6 MPN/100 mL, respectively. Higher concentration of enterococci in sediment and clams than water has been reported before. LeFevre and Lewis 2003, defined a transport pathway for enterococci that begins with the primary source, followed by deposition to a sink, in which enterococci are temporarily sequestered (e.g., sediments), and eventually it can become a secondary source when organisms reenter the water column following a disturbance [25] and can also be filtered by shellfish as clams. Culturable enterococci cells persist longer in sand than water and some studies have documented growth of this microorganism rather than loss, in beach sand [26].

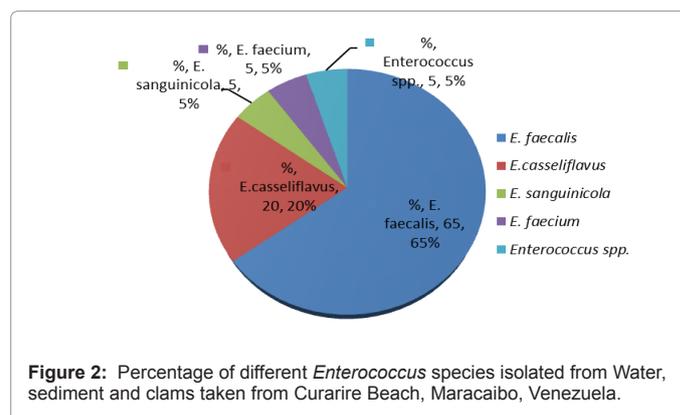
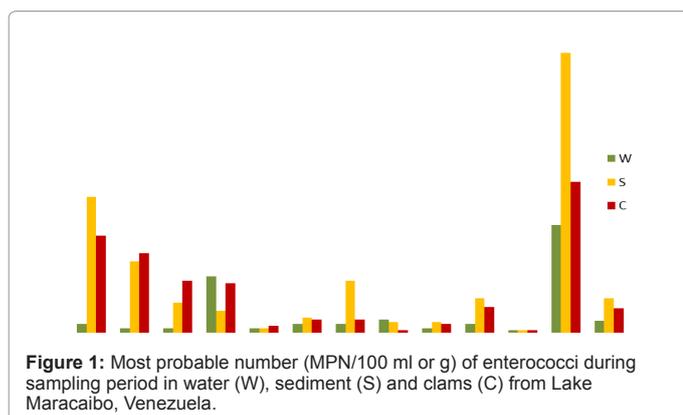
Survival in sediment may be enhanced relative to water because of protection from sunlight/UV inactivation, buffered temperatures, and availability of nutrients accumulated from algae, debris and plankton [27]. Bacteria may also be protected within biofilms on moist sand grains [28]. In some geographical regions, highly favorable conditions may be encountered outside the host. In relatively warm, nutrient-rich, pristine tropical soils and waters, EC have been found at densities far exceeding the concentrations found in highly-polluted temperate waters [29]. *Enterococcus* densities were higher during November to March related with dry season in Venezuela (Figure 1). Other research reported higher values during summer months.

Sixty strains of enterococci were isolated from ninety samples collected form water, sediment and clams at Curarire beach, Lake Maracaibo. The most frequent species isolated was *E. faecalis* (65%) followed by *E. casseliflavus* (20%) (Figure 2). The water samples obtained from different points, *E. faecium* and *E. faecalis* as the most frequent species isolated followed by *E. gallinarum* and *E. casseliflavus* [30]. *E. faecalis* is the most common specie isolated from clinical samples (80-90%), interacts with many other organisms and has effects on the environment. In the other hand, *E. casseliflavus* has been reported in 20% of beach samples and it is associated with natural source, such as plants [31-33]. This is consistent with the results found in this research.

Results showed the importance of surveillance enterococci in tropical water especially in Venezuela in order to determine the ability to survive in Maracaibo Lake and if it's necessary to include it in the Venezuelan guidelines.

Acknowledgement

This research was supported by the Consejo Cientificoy Humanistico (CONDES) University of Zulia, Maracaibo, Venezuela.



References

1. Brtkova M, Filipova H, Drahovska H, Bujdakova (2010) Characterization of enterococci of animal and environmental origin using phenotypic methods and comparison with PCR based methods. *Veterinarni Medicina* 55: 97-105.
2. DeLisle S, Perl TM (2003) Vancomycin-resistant enterococci: a road map on how to prevent the emergence and transmission of antimicrobial resistance. *Chest* 123: 504-518.
3. Marothi YA, Agnihotri H, Dubey D (2005) Enterococcal resistance – an overview. *Indian J Med Microbiol* 4: 214-219.
4. Hayden MK, Bonten MJ, Blom DW, Lyle EA, van de Vijver DA, et al. (2006) Reduction in acquisition of vancomycin-resistant enterococci after enforcement of routine environmental cleaning measures. *Clin Infect Dis* 42: 1552-1560.
5. Dzidic S, Bedekovic V (2003) Horizontal gene transfer-emerging multidrug resistance in hospital bacteria. *Acta Pharmacol Sin* 24: 519-526.
6. Belicova E, Krizkova L, Krajcovic J, Jurkovic D, Sojka M, et al. (2007) Antimicrobial susceptibility of *Enterococcus* species isolated from Slovak bryndza cheese. *Folia Microbiol (Praha)* 52: 115-119.
7. Paoletti C, Foglia G, Princivalli MS, Magi G, Guaglianone E, et al. (2007) Co-transfer of vanA and aggregation substance genes from *Enterococcus faecalis* isolates in intra and interspecies matings. *J Antimicrob Chemother* 59: 1005-1009.
8. Devriese LA, Pot B, Collins MD (1993) Phenotypic identification of the genus *Enterococcus* and differentiation of phylogenetically distinct enterococcal species and species groups. *J Appl Bacteriol* 75: 399-408.
9. Patel R, Piper KE, Rouse MS, Steckelberg JM, Uhl JH, et al. (1998) Determination of 16S rRNA sequences of enterococci and application to species identification of nonmotile *Enterococcus gallinarum* isolates. *J Clin Microbiol* 36: 3399-3407.
10. Shepard BD, Gilmore MS (2002) Antibiotic-Resistant Enterococci: The Mechanisms and Dynamics of Drug Introduction and Resistance. *Microbes Infect* 4: 215-224.
11. Iwen PC, Kelly DM, Linder J, Hinrichs SH, Dominguez EA, et al. (1997) Change in Prevalence and Antibiotic Resistance of *Enterococcus* Species Isolated from Blood Cultures Over an 8-Year Period. *Antimicrob Agents Chemother* 41: 494-495.
12. Damborg P, Top J, Hendrickx APA, Dawson S, Willems RJL, et al. (2009) Dogs area Reservoir of Ampicillin-Resistant *Enterococcus faecium* Lineages Associated with Human Infections. *Appl Environ Microbiol* 75: 2360-2365.
13. (2003) U.S. EPA: Bacterial Water Quality Standards for Recreational Waters (Freshwater and Marine Waters).
14. Cabelli V, Dufour AP, McCabe LJ, Levin MA (1982) Swimming-associated gastroenteritis and water quality. *Am J Epidemiol* 115: 606-616.
15. EPA Emergency preparedness for incidents affecting drinking water and wastewater Systems. *Ground Water and Drinking Water*.
16. <http://water.epa.gov/type/rs/monitoring.vms511.cfm>
17. Bordalo AA, Onrassami R, Dechsakulwatana (2002) Survival of faecal indicator bacteria in tropical estuarine waters (Bangpakong River, Thailand). *J Appl Microbiol* 93: 864-871.
18. Gaceta de Venezuela (1995) Gaceta oficial de la República de Venezuela.
19. Herrera A, Suarez, P (2005) Indicadores bacterianos como herramientas para medir la calidad ambiental del agua costera. *Interciencia* 30: 171-176.
20. Parra-Pardi G (1986) La conservación del Lago de Maracaibo. Diagnostico Ecológico y Plan Maestro. 1ra ed. Lagoven. Caracas. Venezuela.
21. Rivas Z, Márquez R, Trocone F, Sánchez J, Colina M, et al. (2005) Contribución de los principales ríos tributarios a la contaminación y eutrofización del Lago de Maracaibo 13: 68-77.
22. (2009) Standard methods for the examination of water and wastewater. American Public Health Association, Washington, DC, USA.
23. (2001) U.S. Food and Drug Administration. Bacteriological Analytical Manual Online
24. Murray P, Baron E, Jorgensen J, Landry M, Pealler M (2007) Manual of clinical Microbiology 9th edition" Editorial ASM press. Washington DC, USA.
25. Montiel M, Silva R, Nuñez J, Espinoza N, Morales F (2011) Indicadores bacterianos y materia organica en la almeja *Rangia cuneata* y su relación con el agua y sedimento. *Revista de la Universidad del Zulia* 2: 66-78.
26. Le Fevre NM, Lewis GD (2003) The role of resuspension in enterococci distribution in water at an urban beach. *Water Sci Technol* 47: 205-210.
27. Halliday E, Gast RJ (2011) Bacteria in beach sands: an emerging challenge in protecting coastal water quality and bather health. *Environ Sci Technol* 45: 370-379.
28. Davies CM, Long JA, Donald M, Ashbolt NJ (1995) Survival of fecal microorganisms in marine and freshwater sediments. *Appl Environ Microbiol* 61: 1888-1896.
29. Priester JH, Horst AM, Van De Werfhorst LC, Saleta JL, Mertes LAK, et al. (2007) Enhanced visualization of microbial biofilms by staining and environmental scanning electron microscopy. *J Microbiol Methods* 68: 577-587.
30. Carrillo M, Estrada E, Hazen TC (1985) Survival and enumeration of the fecal indicators *Bifidobacterium adolescentis* and *Escherichia coli* in a tropical rain forest watershed. *Appl Environ Microbiol* 50: 468-476.
31. Ferguson D, Griffith JF, McGee CD, Weisberg SB, Hagedorn C (2013) Comparison of *Enterococcus* Species Diversity in Marine Water and Wastewater Using Enterolert and EPA Method 1600. *Journal of Environmental and Public Health* 2013: 6.
32. Ott EM, Müller T, Müller M, Franz CM, Ulrich A, et al. (2001) Population dynamics and antagonistic potential of enterococci colonizing the phyllosphere of grasses. *J Appl Microbiol* 91: 54-66.
33. Aarestrup M, Butaye P, Wolfgang W (2002) Nonhuman reservoirs of enterococci, in *The Enterococci: Pathogenesis, Molecular Biology and Antibiotic Resistance*, M. S. Gilmore, D. B. Clewell, P. Courvalin, G. M. Dunny, B. E. Murray, and L. B. Rice, Eds., pp. 104–105, ASM Press, Washington, DC, USA.

Citation: Montiel M, Silva R, Núñez J, Morales F, Severeyn H, et al. (2013) *Enterococcus* in Water, Sediment and Clams in a Tropical Environment, Maracaibo Lake, Venezuela. *J Marine Sci Res Dev* 3: 133. doi: [10.4172/2155-9910.1000133](https://doi.org/10.4172/2155-9910.1000133)

Submit your next manuscript and get advantages of OMICS Group submissions

Unique features:

- User friendly/feasible website-translation of your paper to 50 world's leading languages
- Audio Version of published paper
- Digital articles to share and explore

Special features:

- 200 Open Access Journals
- 15,000 editorial team
- 21 days rapid review process
- Quality and quick editorial, review and publication processing
- Indexing at PubMed (partial), Scopus, EBSCO, Index Copernicus and Google Scholar etc
- Sharing Option: Social Networking Enabled
- Authors, Reviewers and Editors rewarded with online Scientific Credits
- Better discount for your subsequent articles

Submit your manuscript at: www.editorialmanager.com/environsci