

Optimization of Fertilization Success in the Bivalve Mollusk *Tivela mactroides* under Laboratory Conditions

Yajaira García de Severeyn^{1*}, Ana Villasmil¹, Héctor Severeyn², Félix Morales³ and Marynes Montiel⁴

¹Universidad del Zulia, Facultad Experimental de Ciencias, Dpto. de Biología, Laboratorio de Cultivo de Invertebrados Acuáticos, Venezuela

²Universidad del Zulia, Facultad Experimental de Ciencias, Dpto. de Biología Laboratorio de Sistemática de Invertebrados Acuáticos, Venezuela

³Universidad del Zulia, Facultad Experimental de Ciencias, Dpto. de Biología Laboratorio de Oceanografía, Venezuela

⁴Universidad del Zulia, Facultad Experimental de Ciencias, Dpto. de Biología, Unidad de investigaciones de Microbiología Ambiental Maracaibo, Zulia, Venezuela

Abstract

With the objective to produce seed in an aquaculture facility it was studied under laboratory conditions, the effect of sperm concentration and contact time between gametes to optimized fertilization success of the bivalve mollusk *Tivela mactroides*. Mature oocytes and spermatozooids gotten through gonads dissection of adult mature male and female clams were artificially combined to produce fertilization. Solutions of 5.5×10^6 oocytes were fertilized with 10 different sperm concentrations. Results indicate that the percentage of succeed fertilization was low, highly variable and relying on the sperm concentration. The maximum mean percentage of fertilization was 22% between 1.3×10^7 and 1.7×10^7 spermatozoid/ml. A strong fall in fertilization success was observed at sperm concentrations lower than 10^6 spermatozoid/ml. The time lapsed after the first contact for fertilization that produced the highest fertilization rate was three hours, at sperm concentrations higher than 107 spermatozooids/ml. The highest fertilization percentage found in this study falls between the ranges reported for other bivalve mollusk species.

Keywords: Bivalve; Mollusks; *Tivela*; Veneridae; Fertilization; Sperm concentration; Oocytes; Contact time; Gamete

Introduction

The success of fecundation of the mature oocytes is a key factor to maximize the reproduction. This process in aquatic invertebrates depends on several factors such as the distribution, density and the individuals' behavior, the necessary density of sperms to fecundate the eggs, the longevity and the behavior of the gametes [1,2]. There are not only complex interactions among these biological factors but also with the physical properties and the dynamics of the aquatic environment [3]. Before a wide interval of sperm concentrations, the fecundation rates can diminish quickly when the sperms are diluted in the water column and because they also become less active with time. To counteract the dilution of the gametes and to maximize the fecundation rates, the aquatic animals, after a natural spawn, have developed strategies which increase the probabilities of a successful fertilization [1].

However, the artificial fecundation in aquaculture facilities allows the control of the concentration of sperms therefore the oocytes can be inseminated to an appropriate concentration to maximize the fecundation rate. This procedure avoids the problems associated with dropping fecundation rates and the development of polyspermy [4].

Until the publication of the classic study of Levitan [1], the literature on the study of the factors that could maximize the fecundation in aquatic invertebrates was scarce and dispersed. Nonetheless it is surprising that his paper did not mention any mollusk species when almost 30 years before started this type of studies in pectinid bivalve mollusks [5]. These authors studied the best sperm and oocyte concentrations to get viable larvae of *Pecten maximus*.

In the present investigation we studied the marine clam *Tivela mactroides*, a marine bivalve mollusk, and candidate to be cultivated in order to recover its natural banks in the southwest of the Venezuelan Gulf. These banks were decimated by a spill of petroleum in 1997 [6]. The objective was to optimize the production of embryos using, as high-priority factors, the concentration of sperms and the time lapsed after having been induced the contact among the gametes. The final

objective of this and following investigations is to obtain viable larvae that can be used to repopulate the area affected by the oil spill.

Materials and Methods

Collection of adults of *Tivela mactroides*

The clams were gathered in the beach Miramar, an area of about two Km² of coast which occupies an arid-coastal plain. It is a high energy beach and strong surf located to the north in the capital of the Municipality Buchivacoa, Capatárida Falcon. (11°13 '53 '' North and 70°36 '51 '' West). This population of *T. mactroides* is located to 300 Km. east of the area affected by the spill and it still remains as a pristine area, only being exploited by artisanal fishermen.

The animals were collected by means of a trail with a mesh diameter of 20 mm. In some occasions the fishing was made manually. During the samplings, the quantity and size of the gathered clams was made at random, since the size of the animal is independent of the sexual maturity [7]. Of the gathered clams, 100 were placed into 20 L. plastic containers, with oxygen and water of the collecting size cooled with pack of ice. This was done to avoid as much as possible stress during their transfer by car to the laboratory thus avoiding that clams may have spontaneous spawning.

***Corresponding author:** Yajaira García de Severeyn, Universidad del Zulia, Facultad Experimental de Ciencias, Dpto. de Biología, Laboratorio de Cultivo de Invertebrados Acuáticos, Venezuela, Tel: 58-261-7434136; Fax: 58-261-7434136; E-mail: ygsevereyn@yahoo.com

Received September 04, 2013; **Accepted** December 23, 2013; **Published** December 30, 2013

Citation: Severeyn YG, Villasmil A, Severeyn H, Morales F, Montiel M (2013) Optimization of Fertilization Success in the Bivalve Mollusk *Tivela mactroides* under Laboratory Conditions. J Marine Sci Res Dev 4: 142. doi: [10.4172/2155-9910.1000142](http://dx.doi.org/10.4172/2155-9910.1000142)

Copyright: © 2013 Severeyn YG, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Obtaining of the gametes

To obtain the gametes, the clams were opened up by dissection, and their gonads observed under the microscope to identify the sex. Only mature male and female gametes were used to produce fertilization. For this procedure, gonads were carefully perforated with a sterile micro-scissor to liberate the gametes. Then gonads were washed out and collected into Erlenmeyer. Ovocites were filled with 50 ml and sperm with 180 ml of artificial seawater (Instant Ocean) prepared to 40 ups to create the stock solution.

Obtaining of the experimental concentrations of the gametes

To estimate the concentration of sperms and ovocites in the stock solutions there were taken, 3 times, 5 ml of each solution, which were preserved with five drops of 10% formalin solution. Later on, with a camera of New bauer, the gametes were counted. The mean of the three replicates was taken as the concentration of each solution: 5.8×10^9 spermatozoids/ml, and 5.5×10^6 ovocites/ml. Starting from the stock solutions, there were carried out the dilutions to obtain the concentration series of sperms to study. In order to do them, there were prepared 10 dilutions adding to each rehearsal tube from 9 to 1 ml of artificial seawater to 40 ups and a series from 1 to 9 ml of the solution stock of sperms. Two controls of 1 ml of ovocites solution were set, the first one without sperms to discard that this species does not present parthenogenesis, and the second one with sperms previously inactivates through freezing at 0°C during 4 hours, with the purpose of observing if the sperm contains some protein that intervenes or stimulates the fecundation process.

Induction of the fecundation

After carrying out the dilutions of sperms, 1 ml of the ovocites solution stock was placed in each one of the rehearsal tubes with the respective concentrations to induce the fecundation. The experiments stayed to $28 \pm 1^\circ\text{C}$ in an environmental cabinet, during five hours starting from the mixture of the gametes. Each experiment was carried out with three replicates. Once united the gametes, there were taken samples of 0.05 ml at 30 minutes, three and five hours, placed in an excavated slide and examined under microscope. In this observations there were recorded the total number of ovocites, fecundated ovocites, those not fecundated and the embryos in division.

Results

Effect of the concentration of sperms

A notable result of this investigation was the high variability that was observed in the percentage of fecundation among the studied concentrations. Indeed, although the observed maximum value was of 50% in two single replicates, 33% of all experimental concentrations there was not fecundation of all. For this reason, the non-parametric analysis of variant (Kruskal-Wallis) did not detect significant differences among the studied concentrations ($p = 0.8854$). The maximum mean fertilization percentage, 22%, was observed between 1.3×10^7 and 1.7×10^7 spermatozoids/ml (Figure 1). Above or below these values, the fecundation decayed notably.

Effect of the time lapsed over fecundation

The time lapsed after the contact of gametes affected the percentage of fecundated eggs. Indeed, the percentage of fecundation after three hours was significantly smaller ($p=0.0210$) that during the first half hour or after five hours (Figure 1).

Discussion

In aquatic invertebrates, the fecundation success depends on many factors both internal as external to the gametes. To this respect, Levitan [1] points out four groups of factors: environmental, poblacional, individual and specifically internal (genetics) to the gametes. Products of the interaction of the first three categories are generated three conditions that have to do directly with the gametes: the concentration of these in the aquatic means, their dispersion rate (ability to swim longer distances) and the grade of mixture of the gametes of different animals. To these three conditions, the same author adds the intrinsic factors to the gametes: variation in the individual acting (for example longevity), the variation in the size and the form of the eggs, the morphology of the sperms and the rate of collisions among gametes.

However, it is of consensus that the concentration of sperms is one of the most critical factors. The success of the fecundation and therefore the percentage of fecundated ovocites it is highly dependent of the good concentration of sperms. [4,8-11]. When the fecundation takes place with excess of sperms, polispermia increase, generating a decrease of viable embryos [12,13].

In the present study, only two of the mentioned factors were investigated: The concentration of spermatozoids and the time lapsed after having induced the contact among the gametes (an indirect way to measure the effect of sperm longevity).

With regard to the first factor, it was already indicated that the results showed a lot of variability, so statistically speaking, no significant differences was detected among the studied sperm concentrations. These results coincide with other studies in several aspects.

First, high concentrations of sperms are required in the case of clams. Some authors [8] have reported that concentrations of more than 10 million spermatozoids/ml are necessary to successful fecundation. Our successful sperm concentrations were between 13 and 17 million. However, it is clear that the optimal concentration is a factor related to the species, so it is not surprising to find successful ranges very below respect to the reported here. Indeed, *Spisula sollidissima* [14] has optimal sperm concentrations from 800.000 up to 4.000.000 spermatozoids/ml.

The second aspect evaluated in this investigation with regard to the success of the fecundation was the necessary time to obtain the maximum fecundation percentage. As demonstrated above, three hours was the necessary time to maximize the quantity of fecundated ovocites. This time of maximum fecundation coincides with studied in other bivalve where the range goes from 10 minutes to six hours [2,14-16], but it is evident that this wide range should be product of other factors not evaluated in these researches.

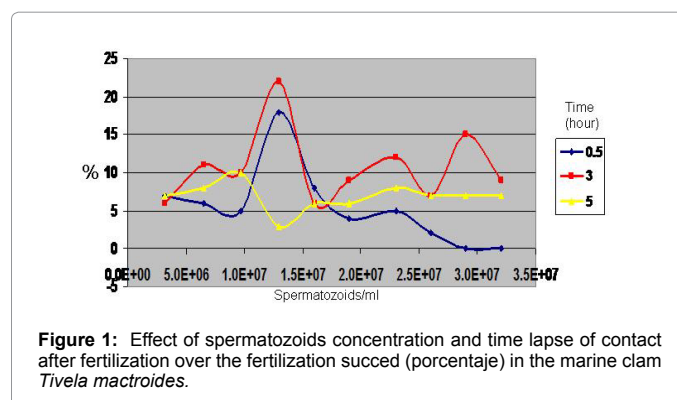


Figure 1: Effect of spermatozoids concentration and time lapse of contact after fertilization over the fertilization succed (percentaje) in the marine clam *Tivela mactroides*.

Conclusions

Considering the main objective of the present study, is not conclusive which is the optimal concentration of sperms to maximize embryos production. However it is also clear that it is not the only necessary factor to achieve high percentages of fecundation in *Tivela mactroides*. Time of contact (longevity of spermatozoid) is also important as we saw in the present results. Both factors together but they do not work alone to affect the success of the fecundation measured as formation of viable embryos. Additional research is necessary to clear this situation.

Considering the necessity to produce the biggest possible number of viable embryos, the range of concentrations of sperms that maximized the fecundation in this investigation was between 1.3 and 1.7×10⁷ spermatozooids/ml after three hours of contact among gametes. Half hour turned out to be little time, but five hours they did not improve the success of fecundation above three hours.

References

1. Levitan D (1998) Sperm Limitation, Gamete Competition, and Sexual Selection 175-202. In External Fertilizers, (1stedn), Academic Press, New York, USA.
2. Song Y, Suquet M, Quéau I, Lebrun I (2009) Setting of a procedure for experimental fertilization of Pacific oyster (*Crassostrea gigas*) oocytes. Aquaculture 287: 311-314.
3. Hodgson A, Le Quesne W, Hawkins S, Bishop J (2007) Factors affecting fertilization success in two species of patellid limpet (Mollusca: Gastropoda) and development of fertilization kinetics model. Mar Biol 150: 415-426.
4. Liu W, Alabi A, Pearce C (2008) Fertilization and embryonic development in the basket cockle, *Clinocardium nuttallii*. J Shellf Res 27: 393-397.
5. Gruffydd L, Beaumont A (1970) Determination of the optimum concentration of eggs and spermatozoa for the production of normal larvae in *Pecten maximus* (Mollusca, lamellibranchia). Helgolander wiss Moeresunters 20 486-497.
6. Severeyn H, Delgado J, Godoy A, García de Severeyn Y (2003) Efecto del derrame de petróleo del buque Nissos Amorgos sobre la fauna macro invertebrada bentónica del Golfo de Venezuela: cinco años después. Ecotrópicos 16: 83-90.
7. Reverol Y, Delgado J, García de Severeyn Y, Severeyn H (2004) Embrionary and larval development of the marine clam *Tivela mactroides* (Bivalvia:Veneridae)in Zulia State,Venezuela. Revista de Biología Tropical.
8. Powell D, Tyler P, Peck LL (2001) Effect of sperm concentration and sperm ageing on fertilization success in the Antarctic soft-shelled clam *Laternula elliptica* and the Antarctic limpet *Nacella concinna*. Mar Ecol Prog Ser 215: 191-200.
9. Luttkhuizen P, Drent J (2003) Fertilization in broadcast spawning marine bivalves is a small scale process: the Baltic clam *Macoma balthica*. J Evol Biol 119-126.
10. Luttkhuizen P, Honkoop P, Drent J, Van Der Meer J (2003) optimal egg size for external fertilization: theory on the role of sperm limitation and field observations for a marine bivalve. J Evol Biol 83-102.
11. Kupriyanova E (2006) Fertilization success in *Galeolaria caespitosa* (Polychaeta: Serpulidae): gamete characteristics, role of sperm dilution, gamete age, and contact time. Scientia Marina 70: 309-317.
12. Narvate M, Pascual M (2003) Fertilization, larval rearing and post-larval growth of the Tehuelche scallop *Aequipecten tehuelchus* D'Orb., 1846. Aquaculture 217: 259-274.
13. FARIAS J (2006) Cultivo de moluscos. 1era Edic.. Editorial Alfaomega. México, D.F., México.
14. Clotteu G, Dube F (1993) Optimization of fertilization parameters for rearing surf clams (*Spisula solidissima*). Aquaculture 114: 339-353.
15. Dos Santos A, Nascimiento I (1985) Influence of gamete density, salinity and temperature on the normal embryonic development of the mangrove oyster *Crassostrea rhizophorae* Guilding, 1828. Aquaculture 47:335-352.
16. O'connor W, Heasman M (1995) Spawning induction and fertilization in the doughboy scallop *Chlamys (Mimachlamys) asperima*. Aquaculture 136: 117-129.

Citation: Severeyn YG, Villasmil A, Severeyn H, Morales F, Montiel M (2013) Optimization of Fertilization Success in the Bivalve Mollusk *Tivela mactroides* under Laboratory Conditions. J Marine Sci Res Dev 4: 142. doi: [10.4172/2155-9910.1000142](https://doi.org/10.4172/2155-9910.1000142)

Submit your next manuscript and get advantages of OMICS Group submissions

Unique features:

- User friendly/feasible website-translation of your paper to 50 world's leading languages
- Audio Version of published paper
- Digital articles to share and explore

Special features:

- 300 Open Access Journals
- 25,000 editorial team
- 21 days rapid review process
- Quality and quick editorial, review and publication processing
- Indexing at PubMed (partial), Scopus, EBSCO, Index Copernicus and Google Scholar etc
- Sharing Option: Social Networking Enabled
- Authors, Reviewers and Editors rewarded with online Scientific Credits
- Better discount for your subsequent articles

Submit your manuscript at: www.editorialmanager.com/environsci