

Research Article

Cloning and Analysis of N-Acetyltransferase 9 Genes in Yellow Catfish *Pelteobagrus fulvidraco*

Ming-Ming Han^{1,2}, Jian-Guo Lu¹, Le-Wang¹, Li-Na-Peng¹, Shahidd Mahboob^{3,4}, Khalid A Al-Ghanim⁴ and Xiao-Wen Sun^{1*}

¹Heilongjiang River Fisheries Research Institute, Chinese Academy of Fishery Sciences, Harbin, China ²Shanghai Ocean University, College of Fisheries and Life Science, Shanghai, China ³Department of Zoology, College of Science, King Saud University, Riyadh, Saudi Arabia ⁴Department of Zoology, GC University, Faisalabad, Pakistan

Abstract

N-acetyltransferase 9 (NAT9) is an important reproduction-related gene in fish. In this study, we cloned the fulllength NAT9 cDNA sequence from yellow catfish *Pelteobagrus fulvidraco*. *P. fulvidraco* NAT9 cDNA is 1253 bp, which including non-coding region (584 bp) and open reading frame (669 bp). NAT9 gene encodes a 222 amino acidsprotein which shares high homology with NAT9 in four species: *letalurus punetaus* (91%), *Astyanax mexicanus* (83%), *Danio rerio* (81%) and *Lepisosteus oculatus* (81%). NAT9 protein is 25.7899 kD, 4.86 theoretical isoelectric point and $C_{1124}H_{1759}N_{311}O_{359}S_{13}$ as revealed by computer-assisted analysis. Tissue transcription profile analysis indicated that the catfish NAT9 gene is generally but differentially expressed in the detected tissues, including gonads, brain, liver, kidney, spleen, intestine, blood, gills, muscle and heart. Identification and tissue distribution of yellow catfish NAT9 genes provided initial step towards understanding their biological roles in yellow catfish.

Keywords: N-acetyltransferase 9; *Pelteobagrus fulvidraco*; Gene clone; Bioinformatics analysis

Introduction

N-acetyltransferases (NATs) are well characterized xenobioticmetabolizing enzymes which catalyze the acetyl Coenzyme A (AcCoA)dependent N-, O- and N,O-acetylation of aromatic amines and hydrazines. They played important endogenous roles, as well as having potential as novel targets for pharmacological intervention. NAT9 is a member of NATS which has N-acetyltransferase, acyltransferase and transferase activities. NAT9 has important biological function, due to the neutral protein electropositivity [1]. NAT acetylates wide range of drugs and xenobiotics into prominence, as acts in the polymorphic metabolism of the anti-tubercular agent isoniazid [2].

Latest studies reported that NAT9 was highly expressed in human brain, gonad, and specifically expressed in the heart, spleen, gonad of adult mouse. Whole-mount in situ hybridization showed that NAT9 specifically expressed in E7.5 and E8.5 mouse embryo brains and in HH10 stage chicken embryo brain. These results suggest that NAT9 may play important roles in the development of embryonic brain and adult brain and gonad's function [3]. As above reports, NAT9 is an important reproduction related gene. Although NAT9 gene was found has expression in human [4], rat [5], cattle [6] and other animals, this gene expression in yellow catfish has not been reported yet. Whether NAT9 gene is associated with the reproductive traits in yellow catfish is still unknown.

Yellow catfish *Pelteobagrus fulvidraco*, an omnivorous freshwater fish, is regarded as a good candidate for freshwater culture in China for its delicious meat and high market value [7]. In this study, we obtained the full-length cDNA sequence of NAT9 gene, and analyzed tissue expression and putative proteins through bioinformatics approaches. The results of our study provide the basic information of NAT9, and illuminate the molecular mechanism of this gene in *P. fulvidraco*.

Materials and Methods

RNA isolation

The P. fulvidraco were obtained from the Chinese Academy of

Fishery Sciences *P. fulvidraco* Breeding Engineering Center. The yellow catfish were 2 years old, 60.6 ± 2.7 g weight and 18.5 ± 3.4 cm length. The extraction of total RNAs from the yellow catfish was conducted using the Trizol (Invitrogen, China) technique following the manufacturer's instructions. The total RNA concentration was determined by measuring the absorbance at OD260. RNA integrity was checked by electrophoresis. The total RNA was reverse-transcribed into cDNA using the aM-MLV RTase cDNA Synthesis Kit (TaKaRa, Japan).

NAT9 cloning

Every tissue total RNA used cDNA reverse treasure article number for RR047A transcription kit and used minus 20 degrees to put on cDNA template. According to *Ietalurus punetaus* in Genbank madtoms NAT9 (serial number DQ353801.1) cloning NAT9 cDNA partial sequence. After the sequence by Heilongjiang River Fisheries Research Institute of products research, development of genomics and molecular breeding research of yellow catfish are found in ninety-nine percent of similarity in the method of the transcriptome gene sequence and gene sequence designed primers. We designed the PCR primers NAT9-F1, NAT9-R1, NAT9-F2, NAT9-R2, NAT9-F3, NAT9-R3, NAT9-F4, NAT9-R4, NAT9-F5 and NAT9-R5 to clone the NAT9 cDNA partial sequence (Table 1).

PCR was conducted for 5 min at 94°C followed by 35 cycles at

*Corresponding author: Xiao-Wen Sun, Heilongjiang River Fisheries Research Institute, Chinese Academy of Fishery Sciences, Harbin, China, Tel: +86-451-84869349; Fax: +86-451-84604803; E-mail: sunxw2002@163.com

Received February 13, 2016; Accepted March 03, 2016; Published March 10, 2016

Citation: Han MM, Lu JG, Wang L, Peng LN, Mahboob S, et al. (2016) Cloning and Analysis of N-Acetyltransferase 9 Genes in Yellow Catfish *Pelteobagrus fulvidraco*. Biochem Physiol 5: 202. doi: 10.4172/2168-9652.1000202

Copyright: © 2016 Han MM, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Citation: Han MM, Lu JG, Wang L, Peng LN, Mahboob S, et al. (2016) Cloning and Analysis of N-Acetyltransferase 9 Genes in Yellow Catfish *Pelteobagrus fulvidraco*. Biochem Physiol 5: 202. doi: 10.4172/2168-9652.1000202

Page 2 of 5

Primer	Sequence (5'-3')	°C T _m	bp size	
NAT9-F1	ATTGGTTCCCTACAACGCTG	56 56 544		
NAT9-R1	CAGCAGCTCTTCCCATGATG			
NAT9-F2	ATGCGGCTGAATGAAGACAC	58.5 58.5 561		
NAT9-R2	CCATGATGCCTCGTTCACAG			
NAT9-F3	GGGTCCCCTGTACCAAGATG	56		
NAT9-R3	CCAACACAACATTGCTGCCT	56	56 366	
NAT9-F4	CCCATCTACAGAGGCAGAGG	59 59 59		
NAT9-R4	TCAGTGGTGAAAACTTGCTTCT			
NAT9-F5	CTGGAAGGCAGCAATGTTGT	56 56 125		
NAT9-R5	AGGGTTAGAGGTTCAGACGC			
Actin-F	CCACCTTCAACTCCATCA	60	60 60 223	
Actin-R	TACCACCAGACAGAACAG	60		

Table 1: Primer used for the study of the P. fulvidraco NAT9 gene.

94°C for 30 s, 56°C for 30 s, and 72°C for 30 s. The final extension step was performed at 72°C for 2 min. Each of the 20 μ L reaction mixtures contained 1.5 mM of MgCl₂, 0.2 mM of dNTP, 0.3 mM of each of the primers, 1 U of Taq DNA polymerase (TaKaRa, Japan), and 5 ng of cDNA. Amplicons of expected sizes were purified using an Agarose Gel DNA Purification Kit (TaKaRa, Japan), and then subcloned into the pMD-18T cloning vector (TaKaRa) connection at 16°C for the night. Tiangen Top 10 into competent *E. coli* in LB Amp tablet on 37°C, or 230 r/min, Pick the bacteria, expand training and gas-bacilli PCR detection, expand training and gas-bacilli PCR detection, expand training inserts of an expected size were sequenced using M13 primers and sequenced at BGI (Shanghai, China). The sequencing results on NCBI homology retrieval.

Sequence analysis of NAT9

Deduced amino acid sequences of NAT9 cDNA were analyzed using the software Vector NTI Advance 11. NAT9 sequences from different organisms were obtained using the NCBI BLAST search program. The cDNA sequence and deduced amino acid sequence from Yellow catfish were analyzed using the BLAST algorithm (http://www.ncbi. nlm.nih.gov/blast). Translation and protein analyses were performed using ExPASy tools (http://us.expasy.org/tools/). A multiple sequence alignment was created with ClustalW (http://www.ebi.ac.uk/clustalw/). Protein sequences retrieved from the public database were used for ORF and domain searches; alignment; and phylogenetic reconstruction. ORFs were predicted using Open Reading Frame Finder (http://www. ncbi.nlm.nih.gov/gorf/gorf.html), and signal peptides and fibrinogenlike domain (FBG) were identified by the NCBI conserved domain feature of blastp (http://www.ncbi.nlm.nih.gov/BLAST) and by the Simple Modular Architecture Research Tool (SMART) (http://smart. emblheidelberg.de).

Tissue expression of NAT9 gene

NAT9 gene was in accordance with full design relative fluorescence quantitative PCR primers for NAT9-F5 and NAT9-R5 (Table 1). Primers were 125 bp amplification length. Primers respectively had actins-F and actins-R. The length of amplified fragment is 200 pb. It can enter the yellow catfish beta actin genes for internal reference. RT-PCR reverse transcription using the reverse transcription kit ABI. The PCR reaction was performed in a 25 lL volume with a SYBR Premix Ex TaqTM Kit (TaKaRa, Japan). Experiment according to the instructions, the reaction fluorescence quantitative instrument (ABI 7500) on the conduct for analysis using 2 μ M of each specific primer, and 1 lL of cDNA using the following procedure: initial denaturation at 95°C for 2 min followed by 40 cycles of amplification (95°C for 10 s and 55°C for 30 s). Experimental analysis need to $_{\Delta\Delta}$ CT multiplied by 100%.

To determine NAT9 gene expression in, samples of 9 tissues including brain, muscle, kidney, gill, intestine, heart, brain, spleen and blood from various healthy male and female yellow catfishes were 2 years old, 60.6 ± 2.7 g weight, 18.5 ± 3.4 cm length, a total of 30 and some channel catfishes as control were isolated, pooled, and flash-frozen in liquid nitrogen. The yellow catfish Tissues were homogenized under liquid nitrogen using a mortar and pestle, and stored at -80° C until RNA extraction.

Phylogenetic analysis

The amino acid sequence and coding regions of NAT9 gene were deduced using the Primer 5 software, NAT9 gene were excerpts from GenBank, number for KT778803 GenBank landed. Sequences of NAT9: *Ietalurus Punetaus, Astyanax mexicanus zebra, Danio rerio, Esox Lucius, Oncorhynchus mykiss, Lepisosteus oculatus, Reochromis spp., Clupea harengus, Astatotilapia burtoni, Oryzias latipes, Neolamprologus brichardi, Fundulus heteroclitus, Xiphophorus maculatus, Fugu rubripes* and *Poecilia Formosa* retrieved from databases were aligned using the ClustalW2 program (http://www.ebi.ac.uk/Tools/clustalw2/). Phylogenetic trees were constructed using the neighbour-joining (NJ) method based on the deduced full-length amino acid sequences with 1,000 bootstrapping replications within the Molecular Evolutionary Genetics Analysis (MEGA 4.0) package.

Bioinformatics analysis

The physicochemical properties, protein domains, secondary structures, secretory proteins, subcellular localization signals, transmembrane regions, and 3D structure models of the putative proteins of NAT9 were predicted and analyzed using online bioinformatics tools, as listed in Table 2. To strengthen the conclusions, more than one bioinformatics tool was used in the prediction of

Item of analysis	Website	Online tool	
Physicochemical property	http://web. expasy. org/protparam/	ExPASy proteomics	
Domain	http://pfam. xfam. org/	Pfam	
Secondary structure	http://www.cbs.dtu.dk/services/SignalP	Signalp 3.0	
Three-dimensional structure mode	http://www.sbg.bio.ic.ac.uk/phyre/html/index.html	Phyre website	
Secretary protein	http://www.cbs.dtu.dk/services/SignalP/	Signal P4. 0 Server	
	Targetp 1.1 server	Targetp 1.1 server	
Subcellular localization signal	http://www.genscript.com/psort/wolf_psort.	WOLF PSORT	
	https://www.predictprotein.org/	https://www.predictprotein.org/	
Transmembrane regions	http://www.enzim.hu/hmmtop/html/adv_submit.	НММТОР	
5.00	http://www.enzim.hu/hmmtop/html/adv_submit.	TMHMM Server v. 2.0	

Table 2: The tools and websites for bioinformatics analysis.

Citation: Han MM, Lu JG, Wang L, Peng LN, Mahboob S, et al. (2016) Cloning and Analysis of N-Acetyltransferase 9 Genes in Yellow Catfish *Pelteobagrus fulvidraco*. Biochem Physiol 5: 202. doi: 10.4172/2168-9652.1000202

secretory proteins, subcellular localization signals, and transmembrane regions for the protein. The sequence alignment of amino acids was performed by using the BLASTX program, which is available on the NCBI website. The amino acid sequences of HsHDR1 and HsHDR2 were aligned, and the phylogenetic tree was constructed with neighborjoining criteria by the MEGA 6.0 software.

Statistical analysis

All data obtained from the qRT-PCR were calculated to the value of $2^{-\Delta\Delta CT}$ and differences were evaluated statistically using a t-test in SPSS 20.0 software (IBM, Chicago, USA). The relative expression levels of each gene in different tissues were calculated to the $2^{-\Delta\Delta CT}$ of the mean and the standard error (SE) from the three replications. Statistical significance was set with a P value less than 0.01.

Results

Phylogenetic analysis of the NAT9 gene

A phylogenetic tree was constructed based on the amino acid sequences of selected NAT9 using the neighbour-joining (NJ) method (Figures 1 and 2). Data were analyzed by Poisson's correction, and gaps were removed by complete deletion. *Neolamprologus brichardi, Astatotilapia burtoni* and *Reochromis spp.* belong to Perciformes; *Fundulus heteroclitus, Oryzias latipes, Xiphophorus maculatus* and *Poecilia formosa* belong to Cyprinodontiformes; *Fugu rubripes* of Tetraodontiformes and yellow catfish NAT9 amino acids have different branches. The results showed that the molecular NAT9 evolutionary relationships and the corresponding species are known similar evolutionary relationships. The NAT9 gene encodes a protein of 222 amino acids which shares high homology with the NAT9 from four species: *letalurus punetaus* (91%), *Astyanax mexicanus* (83%), *Danio rerio* (81%) and *Lepisosteus oculatus* (81%) (Figure 2).

Tissue expression of NAT9 gene

Quantitative real-time PCR was used to determine tissue distribution of NAT9 gene expression in both male and female *P. fulvidraco*. NAT9 genes were widely expressed in all 10 tissues (gonads, brain, liver, kidney, spleen, intestine, blood, gills, muscle and heart). NAT9 gene expression in the male gut and liver were higher than in female. NAT9 gene expression in the female heart, gill, muscle and spleen were higher than males (Figure 3).



Figure 1: Alignment of amino acid sequence of NAT9 in 6 species fish. The identical residues are represented by ^(**), the highly conserved residues are represented by ^(:) and less conserved amino acid residues are represented by ^(:). The conservative substitutions are represented by grey shading.







Analysis of the physicochemical properties

We analyzed the physicochemical properties of the putative NAT9 protein using the ProtParam (http://www.au.Expasy.Org/tools/ protparam.html). The molecular weight of NAT9 protein was 25.7899 kD and 4.86 theoretical isoelectric point. The negatively charged amino acid residues (Asp + Glu) was 40, and positively charged amino acid residues (Arg + Lys) was 26. The instability coefficient of NAT9 protein was 48.15, indicating that the protein was unstable. Theoretical derivation half-life of NAT9 protein was about 30 h. The fat coefficient was 0.669 and grand average of hydrophobic was 71.94. Amino acid content: Glu 12.2%, Leu 9.0%, Thr 7.7%, Arg 7.2%, Gly 6.8%, Ser 6.3%, Asp 5.9%. The hydropathicity were 0.669 (Table 3). The other detailed results of the physicochemical properties for the protein, including the

Protein characteristic Website	NAT9	
The number of amino acids	222	
Formula	C ₁₁₂₄ H ₁₇₅₉ N ₃₁₁ O ₃₅₉ S ₁₃	
Molecular weight	25.7899 kD	
Isoelectric point (pl)	SWISS-MODEL	
Secretary protein Positively charged residues (Arg+Lys)	40	
Negatively charged residues (Asp+Glu)	26	
Instability index	48.15	
Aliphatic index.	0.669	
Grand average of hydropathicity	71.94	

Table 3: Physicochemical properties of the putative NAT9 proteins.

Citation: Han MM, Lu JG, Wang L, Peng LN, Mahboob S, et al. (2016) Cloning and Analysis of N-Acetyltransferase 9 Genes in Yellow Catfish Pelteobagrus fulvidraco. Biochem Physiol 5: 202. doi: 10.4172/2168-9652.1000202

Measure	Position	Value	Cutoff	Signal peptide	Secretory protein
Max.C	30	0.565	0.32	YES	YES
MaX.Y	30	0.690	0.33	YES	YES
Max.S	12	0.989	0.87	YES	YES
Mean S	1-29	0.852	0.48	YES	YES
D	1-29	0.771	0.43	YES	YES

Table 4: Results of signal peptide sequence prediction.

number of amino acids, formula and isoelectric point were summarized in Table 3.

Prediction of signal peptide and transmembrane domain

We analyzed the signal peptides in NAT9 by using the SignalP 3.0 Server (http:// www.cbs.dtu.dk/ervices/SignalP/). Significant signal peptides were predicted based on the putative protein sequence, which indicated that NAT9 was secretory proteins and might play an important role (Table 4). The TMHMM Server v. 2.0 (http://www.cbs. dtu.dk/services/TMHMM/) were also used to analyze transmembrane domains. No transmembrane domains were found in this protein, which indicated thatNAT9 may locate in the cellular cytoplasm.

Prediction of secondary structure and construction of 3D structure model

The secondary structure of the putative protein was predicted by Predictprotein server software (http://www.predictprotein.org/). The percentage of α -helix, β -sheet, and random coil was 45.05%, 4.95%, and 50.00%, respectively. The 3D structures of the putative NAT9 protein were predicted by the SWISS-MODEL, using the GANT (Gcn5-related N-acetyltransferase) as the template. Disorder prediction and fold recognition showed that *P. fulvidraco* NAT9 protein was similar with c3tcvB genetic code template structure (Figure 4). Compared the template and P. fulvidraco NAT9 PDF files by Rasmol view fish; we found two monomer structure were very similar (Figure 4). Gcn5-related N-acetyltransferase super family is a comprehensive super family. They have a huge number of homologous region and acetylated related motif similar kind of material [8], including general control nonderepressible-5', Hat1, Elp3, Hpa2, other eukaryotic and prokaryotic acetyltransferase of different substrates. The P. fulvidraco NAT9 code guarantees may be as an auxiliary activation factor with endogenous type A activity of HATs.

Discussion

In the current study, we firstly cloned the full-length cDNA of P. fulvidraco NAT9 gene. Through sequence analysis, it can be seen that



A - NAT9 Monomer 3D structure B - c3tcvB_ Monomer 3D structure

ISSN: 2168-9652 BCP, an open access journal

Biochem Physiol

the encoding protein of P. fulvidraco NAT9 gene is highly homologous with NAT9 in other fishes. This implied that the NAT9 genes were highly conserved in some fishes, and NAT9 of yellow catfish might have similar functions as in other fishes. NAT9 protein of yellow catfish has new feature compared with other fishes' NAT9, hints that P. fulvidraco NAT9 gene has some unique functions than other fishes.

Tissue expression analysis shown yellow catfish NAT9 gene has huge differentially expressed in detected tissues. The suitable explanation for this under current conditions is that at the same time those biological activities related to the mRNA expression of yellow catfish NAT9 gene were presented diversely in different tissues. This indicates that NAT9 gene has diverse roles in different tissues because of the expression difference. To further understand the function of this novel gene, more research based on these primary results is needed.

The structure of P. fulvidraco NAT9 protein is similar to Gcn5related N-acetyltransferases. Gcn5 have auxiliary activation factor with endogenous type activity of HATs [9]. NAT9 may be as an auxiliary activation factor with endogenous type A activity of histone acetyltransferases (HATs). HATs maintains cell dynamic equilibrium from histone acetylation/deacetylation [10]. The steady-state level of histone acetylation is a balance between the action of HATs and histone deacetylases. Obviously, as a result of histone acetylation plays an important role in the regulation of gene expression. So the unbalance of gene expression regulation will lead to cells disorder. It has been found that many diseases incidence relate to histone acetylation/deacetylation balance disorder.

In summary, based on the sequence from full-length cDNA of the NAT9 belong to NAT family in P. fulvidraco, we analyzed and detected tissue transcription profile and also predicted the putative protein using bioinformatics tools. These results of our study will pave the way to furthers researches the reproduction and disease resistance of NAT9 in P. fulvidraco.

Acknowledgment

The research was funded and supported by grants from National Science Foundation of China (31302177), and The authors would like to extend their sincere appreciation to the Deanship of Scientific Research at King Saud University for funding this Research No. RG 1435-012.

References

- 1. Xi S, Chuang K, Fang K, Lee Y, Chung J, et al. (2014) Effect of berberine on activity and mRNA expression of N-acetyltransferase in human lung cancer cell line A549. J Tradit Chin Med 34: 302-308
- 2. Sugamori KS, Brenneman D, Sanchez O, Doll MA, Hein DW, et al. (2012) Reduced 4-aminobiphenyl-induced liver tumorigenicity but not DNA damage in arylamine N-acetyltransferase null mice. Cancer Lett 318: 206-213.
- 3. Zou X, Ci HL, Chen W, Li YP (2006) [Cloning and expression analysis of human N-acetyltransferase doman containing gene hNATL]. Fen Zi Xi Bao Sheng Wu Xue Bao 39: 22-28.
- Watahiki A, Waki K, Hayatsu N, Shiraki T, Kondo S, et al. (2004) Libraries 4. enriched for alternatively spliced exons reveal splicing patterns in melanocytes and melanomas. Nat Methods 1: 233-239.
- 5. Helms C, Cao L, Krueger JG, Wijsman EM, Chamian F, et al. (2003) A putative RUNX1 binding site variant between SLC9A3R1 and NAT9 is associated with susceptibility to psoriasis. Nat Genet 35: 349-356.
- Zimin AV, Delcher AL, Florea L, Kelley DR, Schatz MC, et al. (2009) A wholegenome assembly of the domestic cow, Bos taurus. Genome Biol 10: R42.
- Tan XY, Luo Z, Xie P, Liu XJ (2009) Effect of dietary linolenic acid/linoleic acid ratio on growth performance, hepatic fatty acid profiles and intermediary metabolism of iuvenile vellow catfish Pelteobaarus fulvidraco. Aquaculture 296: 96-101.

Page 4 of 5

Citation: Han MM, Lu JG, Wang L, Peng LN, Mahboob S, et al. (2016) Cloning and Analysis of N-Acetyltransferase 9 Genes in Yellow Catfish *Pelteobagrus fulvidraco*. Biochem Physiol 5: 202. doi: 10.4172/2168-9652.1000202

- Yu M, de Carvalho LP, Sun G, Blanchard JS (2006) Activity-based substrate profiling for gcn5-related n-acetyltransferases: the use of chloroacetylcoenzyme a to identify protein substrates. J Am Chem Soc 128: 15356– 15357.
- Turner BM (1998) Histone acetylation as an epigenetic determinant of longterm transcriptional competence. Cell Mol Life Sci 54: 21-31.
- 10. Kundu TK, Dasgupta D (2007) Chromatin and Disease. Springer, Netherlands.