Commentry   open access

Current Concepts of the Mechanisms in Age-Related Hearing Loss

Takashi Sakamoto * and Tatsuya Yamasoba

Department of Otolaryngology and Head and Neck Surgery, Graduate School of Medicine, Faculty of Medicine, University of Tokyo, Tokyo, Japan

*Correspondence author:** Sakamoto T, Department of Otolaryngology and Head and Neck Surgery, Graduate School of Medicine, Faculty of Medicine, University of Tokyo, Hongo 7-3-1, Bunkyo-ku, Tokyo 113-8655, Japan, Tel: 81358008924; Fax: 81338149486; E-mail: ta-saka@nifty.com

Commentary

Age-related hearing loss (AHL), also known as presbycusis, is one of the most prevalent chronic degenerative conditions; it progresses with age and affects tens of millions of the elderly worldwide. It is characterized by a decline in auditory function, which is reflected by higher hearing thresholds and poor frequency resolution [1], resulting in the inability to understand words and making it difficult to understand everyday language. The primary pathology of AHL includes the loss of sensory hair cells and spiral ganglion neurons, and strial atrophy [2], in addition to degeneration of the central auditory pathways. AHL is caused by the interaction of multiple factors and shows large variations in the onset and extent of hearing loss. These multiple factors complicate the interpretation of basic and clinical research in AHL [3]. Human epidemiological studies have identified four risk factor categories for AHL, including cochlear aging (individual age), environment (occupational and leisure noise exposure, ototoxic medications, socioeconomic status), genetic predisposition (sex, race, specific genetic loci/genes), and health comorbidities (hypertension, diabetes, stroke, cigarette smoking) [4-6]. Genetic investigation has identified several putative gene associations, including with genes related to antioxidant defense systems, such as glutathione S-transferase and atherosclerosis.

It has been postulated that reactive oxygen species (ROS) play a major role in the degeneration of cochlear cells during aging [7,8]. Oxidative stress in the cochlea may result in enhanced hypoxia owing to a poor cochlear blood supply in atherosclerosis, which could be accelerated by genetic and comorbid factors.

Mitochondria are believed to be a major source of ROS [9-11]. ROS generated inside the mitochondria can damage nuclear DNA, mitochondrial DNA (mtDNA), membranes, and proteins, and further, accumulate such damage. In particular, the level of a 4977-bp deletion in mtDNA, known as the "common ageing deletion", increases with age in human temporal bones [12-15]. *Polg* knockin mice, which were created by introducing a two-base substitution, lost the ability to proofread mtDNA defects and showed an early onset of AHL with severe degeneration of spiral ganglion and cochlear nucleus neurons [16,17]. These findings indicate that the accumulation of mtDNA mutations during aging leads to mitochondrial dysfunction, an associated impairment of energy metabolism, and degeneration of the cochlea and central auditory pathway.

Multiple antioxidant enzymes scavenge ROS and control their damaging effects, including superoxide dismutase, catalase, glutathione S-transferase, and glutathione peroxidase. These antioxidant defense systems may also be influenced by the genetic background including race, and the level of some antioxidant enzymes have been shown to decrease during aging. For example, Mice lacking SOD1 were shown to have accelerated age-related cochlear hair cell loss, reduced thickness of the stria vascularis, and severe degeneration of spiral ganglion neurons [16,17]. These findings indicate that an age-related decline in the cochlear antioxidant defence systems can lead to an age-related increase in ROS and also play a pivotal role in the development of AHL.

In fact, supplementation of the following antioxidants slows AHL in mice and rats: vitamin E and C plus melatonin or lazaroid; lecithin; a mixture of L-cysteine-glutathione mixed disulfide, ribose-cysteine, NW-nitro-L-arginine methyl ester, vitamin B12, folate, and ascorbic acid; α-lipoic acid and coenzyme Q_{10}; and N-acetyl-L-cysteine [8,18-20]. Further, compared to age-matched controls, C57BL/6 mice that received caloric restriction (CR) retained normal hearing and showed no obvious cochlear degeneration and a significant reduction in the number of terminal deoxynucleotidyl transferase dUTP nick end labeling-positive and cleaved caspase-3-positive cells among the spiral ganglion cells. DNA microarray analysis revealed that CR downregulated the expression of 24 apoptotic genes, including *Bak* and *Bim*, suggesting that CR could prevent apoptosis of cochlear cells [21]. Oxidative stress from paraquat induced Bak expression and apoptosis in primary cochlear cells, which was ameliorated in Bak-deficient cells [8]. These results suggested that AHL was caused by oxidative stress-induced Bak-dependent apoptosis. Furthermore, CR failed to reduce oxidative DNA damage and prevent AHL in C57B/6 mice lacking Sirt3, a mitochondrial deacetylase [22]. In response to CR, Sirt3 was deacetylated and it in turn activated mitochondrial isocitrate dehydrogenase 2, leading to increased NADPH levels and an increased ratio of reduced-to-oxidized glutathione in the mitochondria. These findings indicated that the beneficial effects of CR were mediated by ROS-antioxidant systems and that Sirt3 is essential for upregulating the cochlear mitochondrial glutathione antioxidant defence system during CR.

Taken together, the putative AHL mechanism is that the cumulative effects of oxidative stress result in the accumulation of mtDNA mutations/deletions and the decline of mitochondrial function, and that these progressively induce Bak-dependent apoptosis of cochlear cells. AHL may be delayed or prevented by supplementary antioxidants or CR. Large clinical trials are needed to verify if AHL can be delayed or prevented in humans and to clarify the molecular mechanism of AHL.

Acknowledgement

References

1. Pickles JO (2008) An Introduction to the Physiology of Hearing. Emerald Group Publishing, Bingley, UK.
2. Schuknecht HF (1955) Presbycusis. Laryngoscope 65: 402-419.

3. Van Eyken E, Van Camp G, Van Laer L (2007) The complexity of age-related hearing impairment: Contributing environmental and genetic factors. Audiol Neurootol 12:345-358.

4. Cooper JC Jr (1994) Health and Nutrition Examination Survey of 1971-75: Part I. Ear and race effects in hearing. J Am Acad Audiol 5: 30-36.

5. Cruickshanks KJ, Klein R, Klein BE, Wiley TL, Nondahl DM, et al. (1998) Cigarette smoking and hearing loss: the epidemiology of hearing loss study. JAMA 279: 1715-1719.

6. Cruickshanks KJ, Tweed TS, Wiley TL, Klein BE, Klein R, et al. (2003) The 5-year incidence and progression of hearing loss: the epidemiology of hearing loss study. Arch Otolaryngol Head Neck Surg 129: 1041-1046.

7. Cheng AG, Cunningham LL, Rubel EW (2005) Mechanisms of hair cell death and protection. Curr Opin Otolaryngol Head Neck Surg 13: 343-348.

8. Someya S, Xu J, Kondo K, Ding D, Salvi RJ, et al. (2009) Age-related hearing loss in C57BL/6J mice is mediated by Bak-dependent mitochondrial apoptosis. Proc Natl Acad Sci U S A 106: 19432-19437.

9. Balaban RS, Nemoto S, Finkel T (2005) Mitochondria, oxidants, and aging. Cell 120: 483-495.

10. Lin MT, Beal MF (2006) Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. Nature 443: 787-795.

11. Wallace DC (2005) A mitochondrial paradigm of metabolic and degenerative diseases, aging, and cancer: a dawn for evolutionary medicine. Annu Rev Genet 39: 359-407.

12. Bai U, Seidman MD, Hinojosa R, Quirk WS (1997) Mitochondrial DNA deletions associated with aging and possibly presbycusis: a human archival temporal bone study. Am J Otol 18: 449-453.

13. Fischel-Ghodsian N, Bykhovskaya Y, Taylor K, Kahen T, Cantor R, et al. (1997) Temporal bone analysis of patients with presbycusis reveals high frequency of mitochondrial mutations. Hear Res 110: 147-154.

14. Markaryan A, Nelson EG, Hinojosa R (2009) Quantification of the mitochondrial DNA common deletion in presbycusis. Laryngoscope 119: 1184-1189.

15. Markaryan A, Nelson EG, Hinojosa R (2010) Major arc mitochondrial DNA deletions in cytochrome c oxidase-deficient human cochlear spiral ganglion cells. Acta Otolaryngol 130: 780-787.

16. Someya S, Yamasoba T, Kujoth GC, Pugh TD, Weindruch R, et al. (2008) The role of mtDNA mutations in the pathogenesis of age-related hearing loss in mice carrying a mutator DNA polymerase gamma. Neurobiol Aging 29: 1080-1092.

17. Niu X, Trifunovic A, Larsson NG, Canlon B (2007) Somatic mtDNA mutations cause progressive hearing loss in the mouse. Exp Cell Res 313: 3924-3934.

18. Keithley EM, Canto C, Zheng QY, Wang X, Fischel-Ghodsian N, et al. (2005) Cu/Zn superoxide dismutase and age-related hearing loss. Hear Res 209: 76-85.

19. McFadden SL, Ding D, Reaume AG, Flood DG, Salvi RJ (1999) Age-related cochlear hair cell loss is enhanced in mice lacking copper/zinc superoxide dismutase. Neurobiol Aging 20:1-8.

20. Heman-Ackah SE, Juhn SK, Huang TC, Wiedmann TS (2010) A combination antioxidant therapy prevents age-related hearing loss in C57BL/6 mice. Otolaryngol Head Neck Surg 143: 429-434.

21. Seidman MD (2000) Effects of dietary restriction and antioxidants on presbyacusis. Laryngoscope 110: 727-738.

22. Seidman MD, Khan MJ, Tang WX, Quirk WS (2002) Influence of lecithin on mitochondrial DNA and age-related hearing loss. Otolaryngol Head Neck Surg 127: 138-144.

23. Someya S, Yamasoba T, Weindruch R, Prolla TA, Tanokura M (2007) Caloric restriction suppresses apoptotic cell death in the mammalian cochlea and leads to prevention of presbycusis. Neurobiol Aging 28: 1613-1622.

24. Someya S, Yu W, Hallows WC, Xu J, Vann JM, et al. (2010) Sirt3 mediates reduction of oxidative damage and prevention of age-related hearing loss under caloric restriction. Cell 143: 802-812.