

Research Article

Eco-Friendly Nano-emulsion Formulation of *Mentha piperita* Against Stored Product Pest *Sitophilus oryzae*

Magdy A Massoud¹, Manal M Adel^{2*}, Osman A Zaghloul¹, Magdy IE Mohamed³ and Khaled H Abdel-Rheim³

¹Department of Plant Protection, Faculty of Agriculture-Saba Basha, Alexandria University, Egypt

²Department of Pests and Plant Protection, National Research Center, Cairo, Egypt

³Department of Stored Products and Grains Pests, Plant Protection Research Institute, Agricultural Research Center, Sabahia, Alexandria, Egypt

*Corresponding author: Manal M Adel, Department of Pests and Plant Protection, National Research Center, Cairo, Egypt, Tel: 01224214021; E-mail: mhassanein11@hotmail.com

Rec date: October 16, 2018; Acc date: November 26, 2018; Pub date: December 04, 2018

Copyright: © 2018 Massoud MA, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

Nanotechnology is not just involved with small things. Nanotechnology is a multi-disciplinary science. In Egypt, the annual loss in wheat due to stored insects is estimated as equivalent to half a million tons of which 12% is caused by the rice weevil alone. The nanotechnology can be one of the rational solution especially with utilizing essential oils. The efficacy of nano-emulsion of *Mentha piperita* was formulated using ultrasonic emulsification and compared with its essential oil against Rice weevil, *Sitophilus oryzae*. The resulted nano-emulsion droplets were in size of 43.55 nm. Insecticidal activity of *M. piperita* nano-emulsion and the free *M. piperita* EO were evaluated by film residue contact toxicity. *M. piperita* nano-emulsion caused high mortality of insect adults with low concentrations compared to the bulk oil. Moreover, the mortality in *S. oryzae* was increased with increasing exposure time and concentration. The results also showed significant differences of adult emergence between *M. piperita* (EO) and *M. piperita* nano-emulsion after 6 weeks from treatment. The results concluded that *M. piperita* nano-emulsions may be used as an alternative for the control of other stored-product insect pests. They have the advantage of promising insecticidal activity and being eco-friendly and less toxic than synthetic pesticides.

Keywords: Rice weevil; *Sitophilus oryzae*, *Mentha piperita* L.; Nano formulation; EO nano-emulsion

Introduction

Stored-grain insects in developing countries cause huge losses of stored-grain products, amounting to 5-10% loss in temperate regions and 20-30% in the tropical regions [1]. In Egypt, the annual loss in wheat due to stored insects is estimated as equivalent to half a million tons of which 12% is caused by the rice weevil alone [2].

The rice weevil, *Sitophilus oryzae* (Coleoptera: Curculionidae) is a major primary pest, of stored grain-based products, including maize, rice and wheat, particularly in the tropic regions [3,4]. This pest can cause considerable damage to stored grains and completely destroy kernels. Moreover, the product quality is affected by presence of eggs and dead insects, and holes on the grains [5]. According to the global economic estimation, the related costs of pests to stored food stuffs could reach to about 500 million USD per year [6].

Massive applications of conventional pesticides to control these insects result in adverse effects on bene icial organisms, leaves their residues in the food leads to severe risks to human health and environment, it reduced the populations of natural enemies and developed the insect resistance to synthetic insecticides [7,8].

Recently scientists pointed to use naturally occurring insecticides. many of these compounds are secondary plant substances (allelochemicals) including alkaloids, quinones and essential oils [9]. These active substances extracted from plants are effective against wide range of insects and act as toxicants, as insect growth regulator (IGR), repellents or as phagodeterrent [10]. These properties make them suitable bioinsecticides for organic agriculture and could be an alternative to those chemical insecticides.

Owing to the fact, the most essential oils used as flavoring agents possessing insecticidal properties showed ovicidal, larvicidal, adulticidal against several insect species [11,12]. One of the most effective plants against stored pests is *Mentha piperita* L. or peppermint, a plant from the Lamiaceae family, *Mentha piperita* oil possesses diversified potential in the areas of food, cosmetics, medicines, and pest control [13]. The essential oils extracted from *M. piperita* have also been reported as a source of botanical insecticides [14].

However, the major inconvenience of the use of essential oils are their chemical instability in the presence of air, light, moisture and high temperature that can determine the rapid evaporation and degradation of some active components [15]. A method to overcome these problems is the incorporation of essential oils into a controlledrelease nano-formulation which prevents rapid evaporation and degradation, enhances stability and maintains the minimum effective dosage/application [16]. In addition, this nano-formulation compared with bulk formulation is expected to be more effective, showed less toxicity towards non-target organisms and increased persistence of the active ingredient [17,18].

Regarding, the peppermint is considered a medicinal plant and its extracted essential oils have been reported as a source of botanical insecticides. Formulating the essential oils may improve its efficacy as pesticide delivery system. Moreover, the small droplet sizes of EO nano-emulsion with less organic solvents can enhance the kinetic stability and permeability as a result of low surface tension than conventional emulsifiable concentrates. So, this study aims to evaluate the efficiency of *M. piperita* (EO nano-emulsion) as possible as protecting agents of wheat grains against infestation by the rice weevil, Sitophilus oryzae.

Materials and Methods

Insect rearing

Cultures of the rice weevil, Sitophilus oryzae (L.), was maintained in (Stored Products Department, Plant Protection Research Institute, Agriculture Research Center, Dokki, Giza, Egypt.) over 5 years without exposure to insecticides and reared on sterilized whole wheat. Insect rearing and all experimental procedures were carried out at 26.1°C and 65.5% R.H. Adults used in studies was two weeks post-eclosion.

Materials

Tween 80, Acetone and Peppermint oil was purchased as pure oil from Department of Botany and microbiology, Faculty of Science, Alexandria University, Egypt.

Preparation of nano-emulsion

Preparation of nano-emulsion was conducted in National Research Center laboratory, the oil-in-water nano-emulsion was formulated using M. piperita (peppermint) essential oil, non-ionic surfactant (tween 80) and deionized water, according to Ghotbi et al. [19] and Sugumar et al. [20]. The concentration of *M. piperita* (EO) (4%, v/v) was prepared. Initially, coarse emulsion was prepared by adding water to organic phase containing oil and surfactant in ratios 1:5 (v/v) using a magnetic stirrer, which was then subjected to ultrasonic emulsification using a 20 kHz Sonicator (BANDELIN Sonopuls).

M. piperita nano-emulsion characterizations

Droplet size determined at central lab in National Research Center: The emulsion droplet size and size distribution was determined using particle size analyzer (Malvern-UK, 4700 model) Droplet size was analyzed using dynamic light scattering (DLS) technique [21]. Prior to all the experiments, the nano-emulsion oil formulations were diluted with water to get rid of the multiple scattering effects. The droplet size and the polydispersity index (PDI) of the formulated nano-emulsion oil were measured.

Morphology of *M. piperita* nano-emulsion: To visualize the shape and morphology of the formulated M. piperita nano-emulsion oil, transmission electron microscopy (TEM) at the EM Unit in the Faculty of Science, Alex. Univ. was carried out. One drop of emulsion was negatively stained with ethanol and was positioned on a copper grid. The TEM micrographs were acquired using a transmission electron microscope (JEOL JEM-1400Plus) with a tungsten source and operating at 80 kV.

Bioassay technique

Contact toxicity bioassay using thin ilm residue: The insecticidal activity of the tested essential oils against the adults of S. oryzae was determined by direct contact application [22,23]. A series of dilutions of *M. piperita* essential oils were prepared using acetone as a solvent. Aliquots of 1 ml of the dilutions were applied on the bottom of a glass Petri dish (9 cm diameter) to give different concentrations from the bulk of M. piperita and M. piperita nano-emulsion. A ter evaporation

of the solvent for 2 min, 20 adults of tested insects were separately placed into each Petri dish. Control dishes with and without solvent were used. All treatments were replicated three times. Mortality percentages were recorded after 24, 48 and 72 h of treatment and LC50 values were calculated according to Finney [24]. The toxic index (TI) of the tested EO nano-emulsion or free EO was calculated by the following equation according to Yamamoto et al. [25] and Sun [26]:

The toxic index (%) (TI) was calculated (based on LC_{50} after 72 hrs.)

$$Toxicity index (TI) = \frac{LC_{50} of EO nanoemulsion or EO}{LC_{50} of EO}$$

The compound has TI less than 1 (TI1) have high toxicity.

Toxicity increase (%)=(TI of EO-TI of EO nano-emulsion) × 100

Contact toxicity bioassay using treatment wheat grains with free M. piperita (EO) and M. piperita (EO) nano-emulsion: The essential oils were admixed with grains according to Qi and Burkholder [22]. Wheat grains were treated with both M. piperita (EO) and (EO nanoemulsion) at different concentration (0.8, 1.6, 3.3, and 13.3 ml/kg). The M. piperita (EO) and (EO nano-emulsion) were dissolved in acetone (2 ml) then mixed manually with grains (60 gm in 0.4-Litter) glass jars and were divided into three equal replicates. Notably, it has been previously elucidated that when the solvent evaporates, the nanoemulsion retains its properties [27,28]. A ter evaporation of acetone, the treated grains were infested by newly emerged adults (10 pairs). Mortality was recorded every week for two weeks. The number of progeny was recorded a ter six weeks of infestation.

Statistical analysis

Mortality rate was estimated and corrected according to formula [29] as follows:

Corrected Mortality

$$= \frac{Mortality\% of treated insects - Mortality\% of control}{(100 - Mortality\% of control)} \times 100$$

The toxicity data was analyzed using probit analysis to estimate the LC₅₀ (Ldp line).

Results

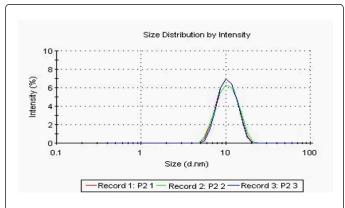
M. piperita nano-emulsion characterizations

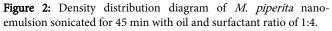
Nano-emulsion droplet size distribution and surface morphology: The nano-emulsion was obtained after sonicating coarse emulsion for 45 min Figure 1 the results showed that emulsion droplets were in the range of 43.55 nm. The average of droplets size recorded 43.55 nm with poly dispersity index (PDI) of 0.787, Figure 2 the morphology of peppermint oil *M. piperita* nano-emulsions was optically transparent or translucent appearance compared with micro-emulsion with the same formulation. The nano-emulsion was visualized using transmission electron microscopy (TEM). Figure 3 show the particles appeared round, spherical in shape, a good dispersion and narrow size distribution, when M. piperita (EO) used at 4% concentration.

Page 3 of 6



Figure 1: Nano-emulsion (O/W) of peppermint essential oil *M. piperita* (4%) obtained by (A) Before Sonication, (B) After Sonication.





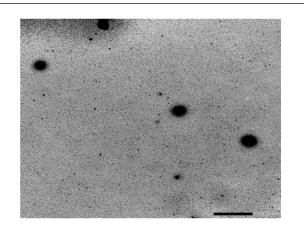


Figure 3: Round, spherical in shape, a good dispersion and narrow size distribution of peppermint oil M. piperita nano-emulsion at 4% by Transmission Electron Microscopy (TEM).

Contact toxicity bioassay using thin film residue

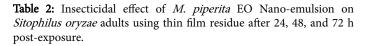
Insecticidal activity of *M. piperita* nano-emulsion and the free *M. piperita* EO were evaluated by film residue contact toxicity against *S. oryzae*. Results showed that both of them have toxic effect against *S. oryzae* after, 24, 48 and 72 h of treatment. The lethal concentration (LC_{50}) accounted 0.181, 0.147 and 0.136 µl/cm² of free *M. piperita* EO against *S. oryzae* after 24, 48 and 72 h post exposure, respectively, as show in Table 1. However, the lethal concentration (LC_{50}) of *M. piperita* nano-emulsion accounted 0.127, 48 h 0.106 and 0.095 µl/cm² after 24, 48 and 72 h of exposure, respectively, as show in Table 2.

	LC ₅₀ µl/cm ²	Confidence Limits				Toxicity
Time		Lower	Upper	Slope	X ²	Index (TI)
24 hr	0.181	0.159	0.199	3.861 ± 0.451	1.132	
48 hr	0.147	-	-	3.696 ± 0.565	12.06 5	
72 hr	0.136	0.109	0.155	4.063 ± 0.674	3.668	1

 Table 1: Insecticidal effect of *M. piperita* on *Sitophilus oryzae* adults using thin film residue after 24, 48, and 72 hr post-exposure.

M. piperita nano-emulsion caused high mortality and toxicity with less concentration compared to the bulk oil. The toxicity index (TI) accounted for *S. oryzae* 0.698 based on LC_{50} after 72 h of exposure as shown in Table 2.

Time	LC ₅₀ µl/cm²	Confidence limits		Slope	X ²			
		Lower	Upper	olope	^	Toxicity	Toxicity	
24 hr	0.127	0.097	0.145	5.102 ± 1.043	0.001	Index (TI)	Increase (%)	
48 hr	0.106	0.031	0.123	5.224 ± 1.737	0.576	•		
72 hr	0.095	0.023	0.123	5.576 ± 2.004	1.859	0.698	30.2	



However, the mortality in *S. oryzae* was increased with increasing exposure time and concentration of *M. piperita* nano-emulsion or the bulk EO in a concentration-dependent manner. The mortality in *S. oryzae* as shown in Table 2 account 68.33, 91.67 and 100% after 24 hr and 86.67, 98.33 and 100% after 48 hr and 88.33, 100 and 100% after 72 hr of exposure to nano-emulsion at concentration 10, 15 and 30 μ l (Figure 4). In contrast, the free *M. piperita* EO showed nearly the same effect with high concentrations ranged from 10-30 μ g as shown in Table 2. These observations demonstrated that *M. piperita* nano-emulsion in lower concentrations compared with essential oil has a considerably greater effect on the adult of *S. oryzae*. The results concluded that, the toxicity effects were significantly more pronounced

Citation: Massoud MA, Adel MM, Zaghloul OA, Mohamed MIE, Abdel-Rheim KH (2018) Eco-Friendly Nano-emulsion Formulation of *Mentha* piperita Against Stored Product Pest Sitophilus oryzae. Adv Crop Sci Tech 6: 404. doi:10.4172/2329-8863.1000404

for the *M. piperita* nano-emulsion formulations compared with the bulk *M. piperita* oil (Figure 5).

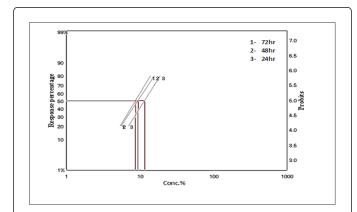


Figure 4: Efficacy of free *M. piperita* EO against *S. oryzae* adults after 24, 48 and 72 hr.

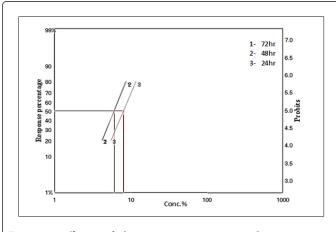


Figure 5: Efficacy of the *M. piperita* Nano-emulsion against *S. oryzae* adults after 24, 48 and 72 hr.

Contact toxicity bioassay using treatment wheat grains with *M. piperita* (EO) and (EO nano-emulsion)

Mortality of *S. oryzae* via Contact toxicity using treatment with wheat grains method, were significantly affected by the exposure time, all main effects (formulations and concentrations) and associated interactions (Table 3).

After one week of exposure, the highest mortality was recorded for *S. oryzae*, exposed to 13.3 ml/kg of *M. piperita* nano-emulsion, where all exposed adults were dead. After two weeks of exposure, 8.3 mL/kg of the tested oil nano-emulsion formulations caused 100% mortality of exposed adults (Table 3).

Among the bulk of *M. piperita* (EO) the highest mortality recorded after one week for *S. oryzae* exposed to 13.3 ml/kg and after two weeks of exposure, 1.6 ml/kg. The results also showed significant differences in the adult emergence of *S. oryzae* in *M. piperita* (EO) and *M. piperita* nano-emulsion after 6 weeks from treatment (Table 3).

Essential oil	Conc.	Mortality (%) after	Mean of emerged adults after		
Essential on	mL/kg	1 week ± SD	2 weeks ± SD	Mean	6 weeks ± SD	Mean
	1.6	0.0 ± 0.0	44 ± 3.13		1.67	0.84
M. piperita (EO nano-	3.3	88 ± 0.57	98.33 ± 0.58		1.67	
emulsion) 4%	8.3	98.33 ± 0.58	100 ± 0.0	78.58	0.0	
	13.3	100 ± 0.0	100 ± 0.0		0.0	
	0.8	82.5 ± 2.52	95 ± 1.73		2.5	0.83
M. piperita (EO)	1.6	85 ± 1.73	100 ± 0.0	94.06	2.5	
	3.3	90 ± 2.64	100 ± 0.0		0.0	
	13.3	100 ± 0.0	100 ± 0.0		0.0	
Control	0.0	0.0	0.0		115	

 Table 3: Effect of *M. piperita* (EO), (EO nano-emulsion) on mortality and emergence of *S. oryzae* adults.

Discussion

Few reports are available on the insecticidal activity of *M. piperita* essential, with fast and high mortality, against stored-product pests. This study showed an increase in the insecticidal effect of M. piperita essential oil against the insect species when formulated as a nanoemulsion. In the present study, the results are in almost agreed with the results stated by earlier investigators. The differences in chemical components may be due to variations in environmental, climatic and geographical which effect on chemical composition of M. piperita. Formulating essential oils (EOs) into nano-emulsion which is transparent and can be used in food and beverage products, thereby, decreasing the amount of EOs required [30]. The TEM image of M. piperita confirmed the results that show the spherical shape and a good dispersion of droplets nano-emulsion agreed with Sugumar et al. [20] and Ostertag et al. [31] who reported that good nano-emulsion had droplets size between 20-200 nm. The tested M. piperita coarse emulsion was turbid and milky white in color due to droplet size in micrometer range. After sonication, the emulsion became optically transparent. This decrease in turbidity was due to minimized droplet diameter after sonication which results in relatively weak scattering making the emulsion system optically transparent agreed with McClements [32]; Pey et al. [33]; Abouelkassem et al. [34].

The bioassay of the nano-emulsion formulations of *M. piperita* demonstrated toxicity effects on *S. oryzae*. The insecticidal effects of the formulations varied with the insect species, concentration of the formulations, exposure time and the method of application. The results of contact toxicity in thin film residue method using glass Petri dish and contact toxicity using treatment with wheat grains method, showed that increasing mortality with increasing concentration and the exposure time for the tested insect [35-38]. The possible explanation for these results is the absorption of the toxic substance increases through insect's body by increase time and concentration. The results indicated that the *S. oryzae* very sensitive when exposed to *M. piperita* (EO) and (EO nano-emulsion) [39-44]. Results agreed with the effectiveness of *M. piperita* EO with concentration 1.0% against *S.*

Page 4 of 6

oryzae evaluated by Magdy et al. [45], which lead to about 96.6% mortality after 24 hr and 100% mortality after 84 hr of exposure using thin film residue method.

The insecticidal activity of *M. piperita* EO is due to the presence of menthol, menthone, methyl acetate, mentho-furan and 1,8-cineole that found as major components. Moreover, the minor compounds can play an important role in EO toxicity. Previous studies showed that insecticidal and biological activity of *M. piperita* EO could be due to the present of major constituents such as menthol, menthone, and menthofuran [46-50]. It has been reported that *M. piperita* EO, have insecticidal activity against many insects such as *S. oryzae* [51-54].

Among the nano-emulsion formulations, the highest and fastest toxic effect were observed with *M. piperita* (4%) nano-emulsion against *S. oryzae* via thin film residue method and treatment with wheat grains method, because of smaller particle size and increase biological activity due to increased surface area of emulsion droplets therefore more opportunity of the formulation to come in contact with the target insect. Whereas, the lower mortality caused by *M. piperita* EO with the biggest particle size indicates that the smaller the particle size, the greater the probability of higher efficacy.

This finding is consistent with the studies of Anjali et al. [17]; Nenaah [55]; Pant et al. [56]; Sugumar et al. [20]; Abouelkassem et al. [34]; Nenaah et al. [28]; Oliveira et al. [57,58]; Choupanian et al. [59]; Mossa et al. [60]; Choupanian et al. [61]. The study showed that *M. piperita* essential oil based nano-emulsion formulations were able to increase the mean mortality rate of *S. oryzae* compared to the free *M. piperita* EO.

Conclusion

The nano-emulsion formulations of *M. piperita* essential oil containing surfactants were successfully created via the high-energy emulsification method. The formulation provided a nano particle-size, with the smallest size being 43.55 nm. The *M. piperita* with the smallest particle size was found to be most effective in controlling *S. oryzae* adults. Overall, the present study proved that *M. piperita* nano-emulsions are effective in controlling *S. oryzae* adults. These nano-emulsions may be used as an alternative for the control of other stored-product insect pests. They have the advantage of promising insecticidal activity and being eco-friendly and less toxic than synthetic pesticides.

References

- 1. Nakakita H (1998) Stored rice and stored product insects. Rice Inspection Technology Manual. ACE Corporation, Tokyo, Japan, pp: 49-65.
- 2. Ministry of Agriculture and Land Reclamation Report (2007) Egypt.
- 3. Howe RW (1965) Losses caused by insects and mites in stored foods and feeding stuffs. In Nutrition Abstracts and Reviews 35: 285-303.
- González JOW, Gutiérrez MM, Ferrero AA, Band BF (2014) Essential oils nano-formulations for stored-product pest control-Characterization and biological properties. Chemosphere 100: 130-138.
- Lü JH, He YQ (2010) Fumigant toxicity of Ailanthus altissima Swingle, Atractylodes lancea (Thunb.) DC and Elsholtzia stauntonii Benth extracts on three major stored-grain insects. Ind Crops Prod 32: 681-683.
- Domínguez Umpiérrez JE, Marrero Artabe L (2010) Catalog of the entomofauna associated with food stores in the province of Matanzas. Fitosanidad 14: 75-82.
- 7. Sharma RP, Yadav RR (2001) Susceptibility Status of Helicoverpa armigera Hub. to some Synthetic Insecticides and Neem Seed Kernel

Extract as In luenced by Host Plant. Pesticide Research Journal 13: 152-159.

- Saleem MA, Ahmad M, Ahmad M, Aslam M, Sayyed AH (2008) Resistance to selected organochlorin, organophosphate, carbamate and pyrethroid, in Spodoptera litura (Lepidoptera: Noctuidae) from Pakistan. J Econ Entomol 101: 1667-1675.
- Appel AG, Gehret MJ, Tanley MJ (2001) Repellency and toxicity of mint oil to American and German cockroaches (Dictyoptera: Blattidae and Blattellidae). J Agric Urban Entomol 18: 149-156.
- 10. Bur ield T, Reekie SL (2005) Mosquitoes, malaria and essential oils. International Journal of Aromatherapy 15: 30-41.
- 11. Souguir S, Chaieb I, Cheikh ZB, Laarif A (2013) Insecticidal activities of essential oils from some cultivated aromatic plants against Spodoptera littoralis (Boisd). J Plant Prot Res 53: 388-391.
- 12. Adel MM, Atwa WA, Hassan ML, Salem NY, Farghaly DS, et al. (2014) Biological activity and field persistence of Pelargonium graveolens (Geraniales: Geraniaceae) loaded solid lipid nanoparticles (SLNs) on Phthorimaea operculella (Zeller) (PTM) (Lepidoptera: Gelechiidae). International Journal of Science and Research 4: 514-520.
- 13. Shah PP, Mello PMD (2004) A review of medicinal uses and pharmacological effects of Mentha piperita.
- 14. Aflatuni A (2005) The yield and essential oil content of mint (Mentha spp.) in Northern Ostrobothnia. PhD Thesis, University of Oulu.
- Regnault-Roger C, Vincent C, Arnason JT (2012) Essential oils in insect control: low-risk products in a high-stakes world. Annu Rev Entomol 57: 405-424.
- Ghormade V, Deshpande MV, Paknikar KM (2011) Perspectives for nano-biotechnology enabled protection and nutrition of plants. Biotechnology Advances 29: 792-803.
- Anjali CH, Sharma Y, Mukherjee A, Chandrasekaran N (2012) Neem oil (Azadirachta indica) nano-emulsion-a potent larvicidal agent against Culex quinquefasciatus. Pest Manag Sci 68: 158-163.
- Devi N, Maji TK (2011) Study of complex coacervation of gelatin a with sodium carboxymethyl cellulose: microencapsulation of neem (Azadirachta indica A. Juss.) seed oil (NSO). International Journal of Polymeric Materials 60: 1091-1105.
- Ghotbi RS, Khatibzadeh M, Kordbacheh S (2014) Preparation of neem seed oil nano-emulsion. In Proceedings of the 5th International Conference on Nanotechnology: Fundamentals and Applications, Prague, Czech Republic, pp: 11-13.
- Sugumar S, Clarke SK, Nirmala MJ, Tyagi BK, Mukherjee A, et al. (2014) Nano-emulsion of eucalyptus oil and its larvicidal activity against Culex quinquefasciatus. Bull Entomol Res 104: 393-402.
- Fernandes CP, Mascarenhas MP, Zibetti FM, Lima BG, Oliveira RP, et al. (2013) HLB value, an important parameter for the development of essential oil phytopharmaceuticals. Rev bras farmacogn 23: 108-114.
- 22. Qi YT, Burkholder WE (1981) Protection of stored wheat from the granary weevil by vegetable oils. J Econ Entomol 74: 502-505.
- Broussalis AM, Ferraro GE, Martino VS, Pinzón R, Coussio JD, et al. (1999) Argentine plants as potential source of insecticidal compounds. J Ethnopharmacol 67: 219-223.
- 24. Finney DJ (1971) Probit analysis. Cambridge University Press, London, p: 318.
- 25. Yamamoto I, Casida JE (1999) Nicotinoid insecticides and the nicotinic acetylcholine receptor.
- Sun YP (1950) Toxicity Index-an improved Method of comparing the relative 378 Toxicity of Insecticides. J Econ Entomol.
- 27. Da Costa JT, Forim MR, Costa ES, De Souza JR, Mondego JM, et al. (2014) Effects of different formulations of neem oil-based products on control Zabrotes subfasciatus (Boheman, 1833) (Coleoptera: Bruchidae) on beans. J Stored Prod Res 56: 49-53.
- Nenaah GE, Ibrahim SI, Al-Assiuty BA (2015) Chemical composition, insecticidal activity and persistence of three Asteraceae essential oils and

Page 5 of 6

their nano-emulsions against Callosobruchus maculatus (F.). J Stored Prod Res 61: 9-16.

- 29. Abbott WS (1925) A method of computing the effectiveness of an insecticide. J Econ Entomol 18: 265-267.
- 30. Qian C, Decker EA, Xiao H, McClements DJ (2012) Physical and chemical stability of β -carotene-enriched nano-emulsions: Influence of pH, ionic strength, temperature, and emulsifier type. Food Chemistry 132: 1221-1229.
- 31. Ostertag F, Weiss J, McClements DJ (2012) Low-energy formation of edible nano-emulsions: factors in luencing droplet size produced by emulsion phase inversion. J Colloid Interface Sci 388: 95-102.
- McClements DJ (2002) Colloidal basis of emulsion color. Curr Opin Colloid Interface Sci. 7: 451-455.
- 33. Pey CM, Maestro A, Solé I, González C, Solans C, et al. (2006) Optimization of nano-emulsions prepared by low-energy emulsi ication methods at constant temperature using a factorial design study. Colloids Surf A Physicochem Eng Asp 288: 144-150.
- Sh A, Abdelrazeik AB, Rakha OM (2015) Nano-emulsion of jojoba oil, preparation, characterization and insecticidal activity against Sitophilus oryzae (Coleoptera: Curculionidae) on wheat.
- 35. Scott IM, Jensen H, Scott JG, Isman MB, Arnason JT, et al. (2003) Botanical insecticides for controlling agricultural pests: piperamides and the Colorado potato beetle Leptinotarsa decemlineata Say (Coleoptera: Chrysomelidae). Arch Insect Biochem Physiol 54: 212-225.
- 36. Upadhyay RK, Jaiswal G (2007) Evaluation of biological activities of Piper nigrum oil against Tribolium castaneum. Bull Insectology 60: 57.
- 37. Arabi M, Frankenberger JR, Engel BA, Arnold JG (2008) Representation of agricultural conservation practices with SWAT. Hydrological Processes: An International Journal 22: 3042-3055.
- Kraikrathok C, Ngamsaengi S, Bullangpoti V, Pluempanupat W, Koul O (2013) Bio efficacy of some piperaceae plant extracts against Plutella xylostella L.(Lepidoptera: Plutellidae). Communications in Agricultural and Applied Biological Sciences 78: 305-309.
- Abo-Arab RB, Helal RMY, Nadia AE (1998) Bioresidul activity of certain oils and plant extracts on some stored grain insects in relation with quality of wheat grain. J Agric Sci Mansoura Univ 23: 5641-5653.
- 40. Negahban M, Moharramipour S, Sefidkon F (2007) Fumigant toxicity of essential oil from Artemisia sieberi Besser against three stored-product insects. J Stored Prod Res 43: 123-128.
- 41. Sahaf BZ, Moharramipour S, Meshkatalsadat MH (2007) Chemical constituents and fumigant toxicity of essential oil from Carum copticum against two stored product beetles. Insect Sci 14: 213-218.
- Sahaf BZ, Moharramipour S, Meshkatalsadat MH (2008) Fumigant toxicity of essential oil from Vitex pseudo-negundo against Tribolium castaneum (Herbst) and Sitophilus oryzae (L.). J Asia Pac Entomol 11: 175-179.
- 43. Ogendo JO, Kostyukovsky M, Ravid U, Matasyoh JC, Deng AL, et al. (2008) Bioactivity of Ocimum gratissimum L. oil and two of its constituents against ive insect pests attacking stored food products. J Stored Prod Res 44: 328-334.
- 44. Saroukolai AT, Moharramipour S, Meshkatalsadat MH (2010) Insecticidal properties of Thymus persicus essential oil against Tribolium castaneum and Sitophilus oryzae. Journal of Pest Science 83: 3-8.
- 45. El-Disouky M (2002) Efficacy of insecticide formulations and alternative methods against certain insects. Doctoral Dissertation, PhD, Thesis in Chemistry of Pesticide, Fac Agric Alex Univ, Egypt.
- 46. İşcan G, Kirimer N, Kürkcüoğlu M, Başer HC, Demirci F (2002) Antimicrobial screening of Mentha piperita essential oils. J Agric Food Chem 50: 3943-3946.

- 47. Behnam S, Farzaneh M, Ahmadzadeh M, Tehrani AS (2006) Composition and antifungal activity of essential oils of Mentha piperita and Lavendula angustifolia on post-harvest phytopathogens. Communications in Agricultural and Applied Biological Sciences 71: 1321-1326.
- Soković MD, Vukojević J, Marin PD, Brkić DD, Vajs V, et al. (2009) Chemical composition of essential oils of thymus and mentha species and their antifungal activities. Molecules 14: 238-249.
- 49. Hayes JR, Stavanja MS, Lawrence BM (2007) Biological and toxicological properties of mint oils and their major isolates: safety assessment. Mint: Genus Mentha. Taylor & Francis Group, Boca Raton, FL.
- Hussain AI, Anwar F, Nigam PS, Ashraf M, Gilani AH (2010) Seasonal variation in content, chemical composition and antimicrobial and cytotoxic activities of essential oils from four Mentha species. J Sci Food Agric 90: 1827-1836.
- Kordali S, Cakir A, Mavi A, Kilic H, Yildirim A (2005) Screening of chemical composition and antifungal and antioxidant activities of the essential oils from three Turkish Artemisia species. J Agric Food Chem 53: 1408-1416.
- Michaelraj S, Sharma RK (2006) Toxicity of essential oils against rice moth, Corcyra cephalonica Stainton in stored maize. Journal of Entomological Research 30: 251-254.
- Michaelraj S, Sharma K, Sharma RK (2007) Fumigation toxicity of some phyto essential oils against stored insect pests of maize. Pesticide Research Journal 19: 9-14.
- Varma J, Dubey NK (2001) Efficacy of essential oils of Caesulia axillaris and Mentha arvensis against some storage pests causing biodeterioration of food commodities. Int J Food Microbiol 68: 207-210.
- 55. Nenaah GE (2014) Chemical composition, toxicity and growth inhibitory activities of essential oils of three Achillea species and their nanoemulsions against Tribolium castaneum (Herbst). Ind Crops Prod 53: 252-260.
- 56. Pant M, Dubey S, Patanjali PK, Naik SN, Sharma S (2014) Insecticidal activity of eucalyptus oil nano-emulsion with karanja and jatropha aqueous iltrates. Int Biodeterior Biodegradation 91: 119-127.
- 57. Oliveira AE, Duarte JL, Amado JR, Cruz RA, Rocha CF, et al. (2016) Development of a larvicidal nano-emulsion with Pterodon emarginatus Vogel Oil. PLOS One 11: e0145835.
- 58. Oliveira AP, Santana AS, Santana ED, Lima APS, Faro RR, et al. (2017) Nano-formulation prototype of the essential oil of Lippia sidoides and thymol to population management of Sitophilus zeamais (Coleoptera: Curculionidae). Ind Crops Prod 107: 198-205.
- Choupanian M, Omar D, Basri M, Asib N (2017) Preparation and characterization of neem oil nano-emulsion formulations against Sitophilus oryzae and Tribolium castaneum adults. Journal of Pesticide Science 42: 158-165.
- 60. Mossa ATH, Abdelfattah NAH, Mohafrash SMM (2017) Nano-emulsion of Camphor (Eucalyptus globulus) Essential Oil, Formulation, Characterization and Insecticidal Activity against Wheat Weevil, Sitophilus granarius. Asian Journal of Crop Science 9: 50-62.
- Choupanian M, Dzolkhifli O (2018) Cytophilus oryzae (L., 1763) (Coleoptera: Curculionidae) and Tribolium castaneum (Herbst, 1797) (Coleoptera: Tenebrionidae) under the control of neem oil nano emulsions and physicochemical identification and formulation. Turkey Journal of Entomology 42: 127-139.