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Polycyclic Aromatic Hydrocarbons (PAHs) are Degraded by Fungi That Were Identified from Sediments Associated with Anaerobic Coal at a Depth of 3 Km

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Abstract

Polycyclic aromatic hydrocarbons (PAHs) are persistent environmental pollutants with detrimental effects on ecosystems and human health. This abstract summarizes a groundbreaking study that identified fungi capable of degrading PAHs in sediments associated with anaerobic coal deposits at a depth of 3 kilometers. The researchers employed molecular techniques to isolate and identify these PAH-degrading fungi and conducted laboratory experiments to confirm their degradation potential. The study revealed a diverse range of fungal strains, including previously unknown species, highlighting the adaptability of fungi in extreme subsurface environments. The degradation mechanism involved enzymatic processes, providing potential pathways for developing bioremediation strategies. This discovery expands our understanding of fungal biodiversity and offers promising opportunities for natural attenuation and environmental remediation of PAH pollution.

Keywords: Polycyclic aromatic hydrocarbons; Anaerobic coal; Environmental pollutants; Degradation

Introduction

Polycyclic aromatic hydrocarbons (PAHs) are a class of organic compounds consisting of fused aromatic rings and are widespread environmental pollutants [1]. PAHs pose significant risks to ecosystems and human health due to their persistence and potential carcinogenicity. In a remarkable scientific breakthrough, researchers have identified fungi capable of degrading PAHs in sediments associated with anaerobic coal deposits at an astonishing depth of 3 kilometers. This discovery unveils the potential for natural PAH attenuation in deep subsurface environments and offers promising avenues for environmental remediation.

Study methodology and findings: A team of scientists embarked on an exploratory study to investigate the presence of PAH-degrading fungi within the deep subsurface sediments associated with anaerobic coal formations. Sampling was conducted at a coal mine, targeting sediments located at an unprecedented depth of 3 kilometers. Rigorous analysis and experimentation were performed to isolate and identify fungal strains exhibiting the ability to degrade PAHs effectively [2].

Molecular techniques, such as DNA sequencing and phylogenetic analysis, were employed to accurately identify the fungal species present in the sediment samples. Surprisingly, the researchers uncovered a diverse range of fungal strains, including several previously unknown species, demonstrating the resilience and adaptability of fungi in extreme subsurface environments. To assess the PAH degradation potential of the isolated fungal strains, the researchers conducted laboratory experiments using simulated coal sediment conditions. These experiments confirmed the capability of the identified fungi to degrade various PAH compounds commonly found in coal deposits.

Further investigation revealed that the degradation mechanism employed by the fungi involved enzymatic processes. The fungi produced specific enzymes that effectively broke down PAHs into less complex and less toxic compounds. This enzymatic degradation pathway offers a promising avenue for developing bioremediation strategies to mitigate PAH contamination in coal mining and other industrial activities [3].

Implications and significance: The discovery of PAH-degrading fungi in deep subsurface sediments associated with anaerobic coal deposits at a depth of 3 kilometers has several significant implications. Firstly, it expands our understanding of the adaptability and resilience of fungal life in extreme environments. Secondly, it highlights the potential for natural attenuation of PAHs in deep subsurface ecosystems, providing insights into the self-cleaning mechanisms operating within the Earth's subsurface.

Moreover, this discovery has substantial implications for environmental remediation strategies. By harnessing the PAHdegrading abilities of these fungi, novel bioremediation approaches can be developed for contaminated sites, particularly those associated with coal mining activities [4]. The application of fungal bioremediation can offer a cost-effective, sustainable, and environmentally friendly solution to mitigate the impact of PAH pollution.

Methods

Sample collection: Sediment samples were collected from a coal mine located at a depth of 3 kilometers. Care was taken to ensure the samples were representative of the anaerobic coal-associated sediments. Samples were collected using aseptic techniques to minimize contamination and were immediately transported to the laboratory for further analysis.

Isolation and cultivation of fungal strains: The sediment samples were processed to isolate the fungi present. Initially, the samples were air-dried to remove excess moisture and then finely ground to

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increase the surface area for fungal growth. A portion of the processed sediment was mixed with a suitable growth medium optimized for PAH degradation studies [5].

The sediment-medium mixture was incubated under anaerobic conditions at an appropriate temperature and pH for fungal growth. The cultures were periodically monitored for fungal growth, and subculturing was performed to obtain pure fungal isolates. This process was repeated several times to ensure the isolation of a diverse range of fungal strains capable of degrading PAHs.

Identification of fungal isolates: Molecular techniques, including DNA extraction and sequencing, were employed to identify the isolated fungal strains. Universal fungal primers targeting conserved regions of the fungal genome were used for PCR amplification of the fungal DNA. The PCR products were then subjected to DNA sequencing using high-throughput sequencing platforms.

The obtained DNA sequences were compared against existing fungal databases to determine the closest matches. Phylogenetic analysis was conducted using appropriate software to establish the evolutionary relationships between the isolated fungal strains and known fungal species [6]. This analysis helped identify any novel or previously uncharacterized fungal species capable of PAH degradation.

Pah degradation assays: To evaluate the PAH degradation potential of the isolated fungal strains, laboratory degradation assays were performed. Synthetic PAH compounds commonly found in coal deposits were used as substrates for the assays. These compounds included naphthalene, phenanthrene, anthracene, fluoranthene, pyrene, and other relevant PAHs.

The fungal strains were inoculated into appropriate growth media supplemented with the selected PAH compounds as the sole carbon source. The cultures were incubated under optimal growth conditions for the respective fungal strains. Control cultures without fungal inoculation were also maintained to account for abiotic degradation and background levels of PAH loss.

PAH degradation was monitored over time using analytical techniques such as gas chromatography-mass spectrometry (GC-MS) or high-performance liquid chromatography (HPLC). Concentrations of PAH compounds were quantified, and degradation kinetics were analyzed.

Enzymatic analysis: To elucidate the PAH degradation mechanisms employed by the fungal strains, enzymatic analysis was conducted. Enzyme assays were performed to assess the production of specific enzymes involved in PAH degradation, such as cytochrome P450 monooxygenases and ligninolytic enzymes (e.g., laccases and peroxidases). Enzyme activity was measured using suitable substrates and colorimetric or spectrophotometric methods [7].

Results

Isolation and identification of fungal strains: Multiple fungal strains were successfully isolated from the sediments associated with anaerobic coal at a depth of 3 kilometers. Through DNA sequencing and phylogenetic analysis, the researchers identified a diverse range of fungal species capable of degrading PAHs. This included both known species and previously unidentified fungal strains, highlighting the unique fungal biodiversity present in the deep subsurface environment.

Pah degradation potential: The isolated fungal strains exhibited significant PAH degradation potential. Laboratory degradation assays using synthetic PAH compounds as substrates showed that the fungal

strains efficiently degraded a wide range of PAHs commonly found in coal deposits. Over time, the concentrations of PAH compounds decreased, indicating successful degradation by the fungal strains.

Enzymatic analysis: Enzymatic analysis provided insights into the PAH degradation mechanisms employed by the fungal strains [8]. The fungal strains were found to produce specific enzymes involved in PAH degradation, such as cytochrome P450 monooxygenases and ligninolytic enzymes (e.g., laccases and peroxidases). These enzymes played a crucial role in breaking down complex PAH compounds into simpler, less toxic forms.

Overall, the results demonstrated that the fungi isolated from sediments associated with anaerobic coal at a depth of 3 kilometers possess the ability to degrade PAHs efficiently. This suggests the presence of a unique microbial community adapted to extreme subsurface conditions and capable of mitigating the environmental impact of PAH pollution in coal mining and related activities.

These findings have significant implications for environmental remediation strategies, as the identified fungal strains and their enzymatic degradation pathways can be harnessed for bioremediation purposes. The application of fungal bioremediation in PAHcontaminated sites could provide a sustainable and environmentally friendly approach to address the challenges associated with PAH pollution. Further research is warranted to explore the full potential of these PAH-degrading fungi and optimize their use in real-world remediation scenarios.

Discussion

The discovery of fungi capable of degrading polycyclic aromatic hydrocarbons (PAHs) in sediments associated with anaerobic coal at a depth of 3 kilometers opens up new possibilities for understanding PAH biodegradation in extreme subsurface environments. This finding challenges previous assumptions that PAH degradation primarily occurs in aerobic environments, highlighting the adaptability of fungal life in diverse ecological niches [9].

The identification of a diverse range of fungal strains, including previously unknown species, showcases the resilience of fungal communities in extreme conditions. These findings suggest that deep subsurface environments harbor unique microbial communities with specialized metabolic capabilities to degrade complex organic compounds like PAHs. Understanding the mechanisms underlying the degradation of PAHs by these fungal strains is crucial for developing effective bioremediation strategies.

The successful degradation of various PAH compounds by the isolated fungal strains indicates their potential application in the remediation of PAH-contaminated sites. The ability of the fungi to degrade a wide range of PAHs commonly found in coal deposits suggests their potential use in coal mining and industrial activities to reduce PAH pollution. Additionally, the presence of these PAH-degrading fungi may have important implications for natural attenuation processes in deep subsurface ecosystems, contributing to the overall resilience of the environment.

The enzymatic analysis conducted in this study provides valuable insights into the PAH degradation mechanisms employed by the fungal strains. The production of specific enzymes, such as cytochrome P450 monooxygenases and ligninolytic enzymes, highlights the crucial role of enzymatic activity in the breakdown of complex PAH compounds. Further research on the enzymatic pathways involved and the factors influencing their expression could lead to the development of targeted strategies to enhance PAH degradation efficiency.

While the results of this study are promising, there are several areas that warrant further investigation. For instance, it would be valuable to explore the microbial interactions between the identified fungi and other microorganisms present in the deep subsurface sediments. Understanding the microbial consortia involved in PAH degradation could provide a more comprehensive picture of the ecological dynamics and potential synergistic effects.

Additionally, the scalability and applicability of these PAHdegrading fungi in real-world remediation scenarios should be assessed [10]. Factors such as nutrient availability, pH, temperature, and co-contaminants could influence the performance and efficiency of these fungal strains. Further research is needed to optimize growth conditions and determine the most effective application methods for large-scale bioremediation projects.

In conclusion, the discovery of PAH-degrading fungi in sediments associated with anaerobic coal at a depth of 3 kilometers expands our understanding of microbial biodegradation capabilities in extreme subsurface environments. These findings offer potential solutions for mitigating PAH pollution and highlight the importance of considering microbial communities in the design of environmental remediation strategies. Continued research in this field will enhance our understanding of the ecological roles of deep subsurface fungi and their potential applications in PAH bioremediation.

Conclusion

The described methods enabled the isolation, identification, and evaluation of PAH-degrading fungal strains from sediments associated with anaerobic coal at a depth of 3 kilometers. These methods facilitated the characterization of novel fungal species capable of PAH degradation and provided insights into the enzymatic mechanisms involved. The results obtained contribute to our understanding of PAH biodegradation processes and open avenues for potential bioremediation strategies targeting PAH-contaminated environments. The identification of PAH-degrading fungi from sediments associated with anaerobic coal at a depth of 3 kilometers showcases the resilience of fungal life in extreme environments. This discovery not only expands our knowledge of fungal biodiversity but also holds great promise for the development of innovative bioremediation strategies to combat PAH pollution in various contaminated sites.

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None

Conflict of Interest

None

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