

Residual Hepatic HCV-RNA in the Native Liver of Living Donor Liver Transplant Recipients with Chronic Hepatitis C: Mini-Review

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Abstract

Chronic Hepatitis C Virus (HCV) infection is the leading risk factor for Hepatocellular Carcinoma (HCC), with an annual HCC risk of 2%-4% in cirrhotic patients. With the highly effective and safe direct-acting antiviral (DAA) therapy, HCV infection would be considered "cured and eliminated" successfully in >95% treated patients. However, even with the widespread use of direct-acting antivirals, Living Donor Liver Transplantation (LDLT) plays an important role in advanced liver disease. According to our recent observation, the pre-transplant serum HCV RNA levels may give an underestimate of the number of positive HCV RNA cases and that hepatic HCV RNA data may be more accurate.

Herein, we would like to have a mini review to detailed address issues including the significance of hepatic HCV RNA and the discrepancy between hepatic HCV RNA and HCV core antigen in native liver of chronic hepatitis C recipients undergoing LDLT, a new insight into cure and complete elimination of HCV infection and the utilization of HCV-aviremic organs into aviremic recipients in liver transplantation.

Keywords: Hepatitis C virus; Hepatocellular carcinoma; HCV RNA; Direct-acting antiviral agents; Liver transplantation

Abbreviations: AFP: Alpha Fetoprotein; Anti-HCV: Antibody for Hepatitis C Virus; BCLC: Barcelona Clinic Liver Cancer; DAA: Direct-Acting Antiviral Agents; DNA: Deoxyribonucleic Acid; HCC: Hepatocellular Carcinoma; HCV: Hepatitis C Virus; LDLT: Living Donor Liver Transplantation; LT: Liver Transplantation; RNA: Ribonucleic Acid; RT-PCR: Reverse Transcription-Polymerase Chain Reaction

Introduction

In clinical practice, despite Hepatitis C Virus (HCV)-infected patients achieved a Sustained Viral Response (SVR) with Direct-Acting Antiviral Agents (DAAs), they remained at risk of Hepatocellular Carcinoma (HCC) development [1-3]. A recent study showed unusually high rates of HCC after treatment with DAA therapy for hepatitis C-related liver cirrhosis [4]. Just as the similar phenomenon occurred in patients diagnosed to have COVID-19, the residual virus reservoirs in tissue specimens is warranted to be explored due to the potential replication and transmissibility in recovered individuals [5,6]. Hence, it is important to determine whether a SVR in patients who have underwent DAA treatment is equal to disease cured or persistent HCV infection which will progress in target organ, leading to future liver cirrhosis and HCC development.

A recent study conducted by Wang presented that HCV-RNA would persist in hepatocytes and/or PBMC in a certain of patients who achieved spontaneous or treatment-induced HCV RNA clearance from serum [7]. Furthermore, detection of occult hepatitis C Virus infection in patients with SVR to DAAs for recurrent infection after liver transplantation also has been reported in 2017 [8,9].

The clearances of HCV and treatment response to anti-viral therapy were associated with genetic variants [10]. In the setting of our liver transplant program, Chiu has investigated the association of *IL28B* SNPs rs12979860 and rs8099917 on HCV RNA status in donors and recipients of LDLT [11]. In the *IL28B* SNP rs12979860, high expression of unfavorable genotype CT in both recipient and donor led to high HCV-RNA recurrence after LDLT.

Notably, with the availability of DAA therapy, the number of liver transplants for HCV had decreased by more than one-third over the past

decade worldwide [12]; however, there still remain some controversies to be clarified. The purpose of this article is to make an evident-based mini review of current issues and findings related to HCV patients undergoing LDLT in our liver transplantation program.

Identification of residual hepatic HCV-RNA in the native liver

In recent years, the introduction of DAA has made HCV become an easily treatable disease, with high rates of SVR, which is associated with improvement of liver dysfunction, promotion of fibrosis regression and reduction the risk of HCC development [1-3].

In recent study, we further evaluated 80 recipients with chronic hepatitis C [13]. Before LDLT, 36.25% (29/80) were serum positive and

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63.75% (51/80) serum negative for HCV RNA (Table 1). After LDLT, we performed one-step reverse-transcription QPCR to generate a standard curve for the absolute quantification of hepatic HCV RNA. Quantification of relative expression (reported as arbitrary units (copy number)) was performed using the $2^{-\Delta\Delta Ct}$ relative quantification method. Quantitative PCR data showed that the variability coefficient of Ct was always lower than 2% of the mean values (Figure 1). After LDLT, 85% (68/80) of native livers were positive for hepatic HCV RNA, whereas 15.0% (12/80) were negative for hepatic HCV RNA (Table 1). Among 68 recipients with positive hepatic HCV RNA, 3.75% (3/80) remained serum positive for HCV RNA (Figure 2). There was a statistically significant difference in HCV RNA identification between the serum HCV RNA and hepatic HCV RNA ($p < 0.001$) before LT and between the serum HCV RNA and hepatic HCV RNA ($p < 0.001$) after LT (Table 1).

Among 68 recipients with positive hepatic HCV RNA, 39.7% (27/68) had been treated with DAA before LDLT, of which 51% (15/27) were positive for serum HCV RNA and 30.8% (12/27) were negative for serum HCV RNA. In total, 44% (3/68) cases remained positive

for serum HCV RNA after LDLT. All three positive serum HCV RNA recipients sustained viral response because of DAA treatment after LDLT. There was no significant difference in hepatic HCV RNA levels between samples from patients pretreated with DAA and serum HCV RNA levels before and after LDLT (Table 2).

Our recent study concluded that the significant underestimation of HCV RNA in the serum was identified by measuring the hepatic HCV RNA levels ($p < 0.001$) followed by their comparison with the pre-LDLT serum HCV RNA and the removed native liver hepatic HCV RNA levels.

HCV core antigen (HCV Ag) in native liver of chronic hepatitis C recipients

According to guidelines from European Association for the Study of the Liver (EASL) recommended HCV core Antigen (HCV Ag) testing to be a less expensive alternative to HCV RNA for determination of HCV viraemia when HCV RNA testing is not available [14]. In 2018, Tilborg conducted a research to support the use of HCV Ag testing

Table 1: Comparison of the HCV RNA status of the native liver and serum before/after liver transplantation in 80 recipients who underwent living donor liver transplantation.

Category	Native liver		p value
	Hepatic HCV RNA (+)	Hepatic HCV RNA (-)	
	N=68 (85%)	N=12 (15.0%)	
Serum HCV RNA before LT	29 (36.3)	51 (63.7)	<0.001
Serum HCV RNA after LT	3 (3.7)	77 (96.3)	<0.001

HCV: Hepatitis C virus; LT: Liver Transplantation; Fisher's exact test

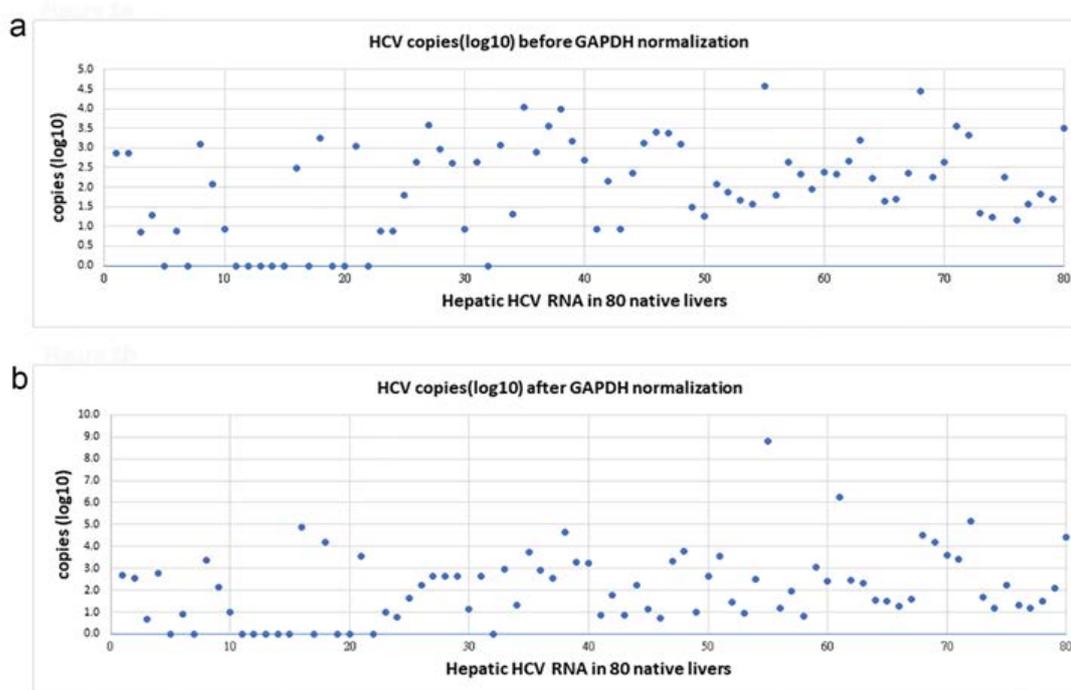


Figure 1: One step reverse-transcription QPCR for absolute quantification of HCV to determine copy numbers (1a) Normalized with human GAPDH expression (1b). **Abbreviations:** HCV: Hepatitis C Virus.

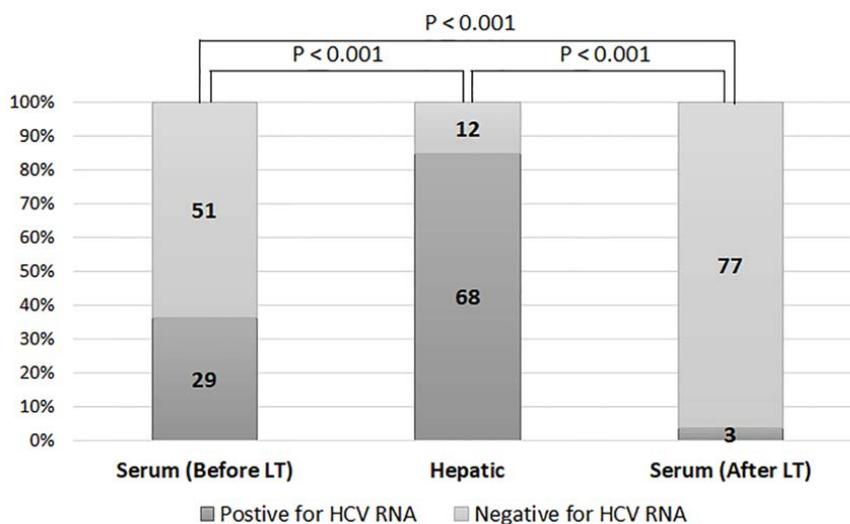


Figure 2: Hepatic HCV RNA showed higher positivity than the serum samples collected before and after living donor liver transplantation. Abbreviations: HCV: Hepatitis C Virus; LT: Liver Transplantation.

Table 2: The relationship between the hepatic HCV RNA levels in patients pre-treated with/without direct-acting antiviral agents and the outcome of serum HCV RNA after living donor liver transplantation.

Native liver	Hepatic HCV RNA (+)		Hepatic HCV RNA (-)	p value
	N=68 (%)		N=12 (%)	
Pre-Tx DAA	27 (39.7)		7 (58.3)	0.187
Serum HCV RNA before LT	Positive	12 (30.8) ^a	Negative	0.067 [*]
	Negative	29 (42.7) ^b	12 (100)	
Serum HCV RNA after LT	Positive	3 (4.4)	Negative	0.414 ^{**}
	Negative	65 (95.6)		
Post-Tx DAA	1 (3.5) ^c	2 (5.1) ^c		

^a =aa':bb'; ^b =aa':cc'; Fisher's exact test; Pre-Tx: pre-treatment; Post-Tx: Post-Treatment; DAA: Direct-Acting Antiviral Agent.

to document HCV viraemia in a cost-saving diagnostic algorithm (sensitivity 94%, 95% CI 86-98; specificity 100%, 95% CI 94-100); it also could be used for documentation of treatment adherence, but it might not be adequate to determine SVR [15]. Recently, a study in Taiwanese cohort analyzing serum samples from 110 patients treated with DAAs suggested that HCV Ag assay may be a feasible alternative to HCV RNA for the determination of SVR12 in patients treated with DAAs [16].

In our liver transplant setting, we investigated the discrepancy between hepatic HCV RNA and HCV Ag in the liver tissue by using one step reverse transcribed QPCR for identification of hepatic HCV RNA and HCV Ag assay based on the protocol provided by Human Hepatitis C Virus Core Antigen ELISA Kit (Cat. No: MBS 167758) in the explanted native liver of CHC patients undergoing LDLT. Among 80 HCV related recipients, 85% (68/80) positive HCV-RNA was significantly higher in the native liver tissues than in the serum before (29/80, 36.3%; p=0.000) and after LDLT (3/80, 4.4%; p=0.000). In contrast, hepatic HCV Ag was 100 % negative identified in all 80 removed native liver.

Based on our research, we suggested HCV Ag assay may have lack of sensitivity and negative predictive value in liver tissues. In contrast to the serum HCV RNA and HCV Ag, a great discrepancy might be described between hepatic HCV RNA and HCV Ag in the liver tissue.

New insights into cure of HCV infection

Similar to the mechanism of antiviral treatment for hepatitis B infection, patients with negative serum HCV RNA may not have HCV clearance in the liver. In contrast, the high percentage of positive hepatic HCV RNA represents that HCV returns to the hepatocyte as well as a target organ for its replication. It may not be beneficial for the long-term outcomes of chronic hepatitis C infection. Recent reports have shown unusually high rates of HCC after treatment with DAA for hepatitis C-related liver cirrhosis [17,18]. Therefore, high-viral carriers target organs, advanced cirrhosis and HCC are completely cured simultaneously through liver transplantation. Obtaining safe results is a real benefit for patients. In our study, although there were no statistically significant differences in clinical liver function before and after LDLT between patients positive and negative for hepatic HCV RNA, 4.4% of recipients with positive hepatic HCV RNA remained positive for serum HCV RNA after LDLT. All three recipients had already been treated with DAA after LDLT and had a sustained viral response in the serum. The outcome of DAA treatment after LDLT may be different from that of the other 77 recipients, as HCV is driven back to a new liver graft. The high load of HCV RNA in a liver graft may be an entirely new concern. Owing to hepatocyte infection with HCV, cell surface levels of sulfate proteins increase and subsequently, the virus enters the liver cells by regulating SMAD6 and SMAD7 [19]. In contrast, DAA does not inhibit

HCV in liver cells. The new liver graft may become a target organ for HCV infection after DAA treatment. In our recent studies, we showed that HCV enhanced expression of micro-RNA 122, micro-RNA 92b and PNPLA3 variants, as well as increased the risk of developing HCC; these form the clinical evidence of the recurrence of HCC [20-22]. In addition to environmental factors such as alcohol and tobacco [23,24], hepatic HCV RNA has been reported to play a role in the severity of HCV infection and the response to antiviral treatment [25]. We suggest that the chronic liver injury concomitant residual hepatic HCV-RNA and long-term outcomes with DAA therapy could still remain an issue after SVR [7].

HCV-positive donors to HCV-negative recipients

Liver transplantation has been hampered by the shortage of available donors in relation to the number of people on the waiting list. To decrease the number of deaths on the waiting list and overcome the problems of lack of donors, many transplant centers aggressively use marginal donors or even HCV seropositive, non-aviremic donors to HCV-seronegative recipients [26]. Recent research from Kapila showed a real-world experience of the transplantation of HCV-aviremic organs into aviremic recipients and they stated that in carefully selected patients, the use of HCV-aviremic grafts in the DAA era appears to be efficacious and well tolerated [27].

However, based on our study, hepatic HCV RNA might still be found after LDLT even in patients with pre-transplant negative serum HCV RNA. Serum HCV-RNA load might represent an underrated estimation to entire HCV viral loads in patients receiving anti-viral therapy. Nowadays, there is no strong evidence published yet to correlate high *de novo* HCC occurrence or recurrence after LT in hepatic HCV RNA patients, but given the constant exposure to immunosuppressant in this organ-transplant cohort in the real-world setting, the oncologic safety of aviremic/viraemia HCV-infected organ donor may need longer duration of observation, even in the DAA era.

Conclusion

A complete eradication of HCV in patients treated with antiviral agents after clinical diagnosis of SVR seems unlikely despite of substantially improved anti-HCV efficacy. Based on the results of our study, the importance of pre-transplant antiviral therapy to attain negative HCV RNA needs to be emphasized here; it has proven to be far more beneficial than post-transplant antiviral therapy.

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Authors Contributions

Study concept and design: King-Wah Chiu and Chih-Chi Wang; data collection: Shu-Hsien Lin and Chih-Che Lin; data analysis and interpretation: King-Wah Chiu and Chih-Chi Wang; performed experiments: Kuang-Den Chen, Kuang-Tzu Huang and Li-Wen Hsu; and manuscript drafting and critical revisions: Shu-Hsien Lin, King-Wah Chiu.

Conflicts of Interest

The authors declare that they have no competing interests.

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