

Review Article

OMICS International

The Role of Microglia in the Injured Neurosystem

Nicholas Sanchez BS and Wolff M Kirsch*

Division of Biochemistry, School of Medicine, Loma Linda University, Loma Linda, CA 92350, USA

Abstract

Microglia is the brain's innate neuroprotective actor responsible for maintaining neuroplasticity and directing a remedial response when confronted with a pathogenic insult or physical trauma. Microglia are enabled to defend the central nervous system to ensure that homeostasis is maintained. Unfortunately, the pro-inflammatory responses microglia use to shield the neurons from threats also have the capability to cause damage detrimental to cognitive stability. This review covers some of these pathways involved in both neuroprotective and neurodegenerative outcomes, then moves on to possibilities on how these results can lead to diseases, specifically Alzheimer's disease. Finally, we comment on the opportunities and challenges of various experimental models currently being used for microglia studies.

Keywords: Microglia; Oxidative stress; Reactive oxygen species; Neuroinflammation; Animal models; Alzheimer's disease; Parkinson's disease; Aging

Introduction

Microglia, the resident immune and phagocytic cells of the neural system, were observed and studied intensively for decades before they were named and identified. As far back as 1841, anatomists were identifying these cells in damaged brains as being of mesodermal origin, their role being similar to macrophages, and concluded their loss of function led to disease. Researchers showed that these cells enter the brain in the second month of life, colonize both white and grey matter, and function as brain macrophages. In 1932 these cells were defined with embryonic origins as we are familiar with today with the demonstration that microglia were a distinct group of glial cells that made up the mesoglia [1]. Further research has refined our understanding of these cells as the descendants of myelomonocytic lineages derived from progenitors in the primitive yolk sac that seed the brain rudiments and enter the embryonic neuroepithelium when neurulation is completed [2]. These cells convert to amoeboid microglia later in life and are positioned in the neuroectoderm even before vascularization is complete, eventually switching into a different ramified state to function in their neuroprotective duties for the brain [3].

Microglia have two primary functions: immune defence and maintenance of the central nervous system (CNS). As immune cells, they detect tissue damage and pathogenic infections using numerous cell receptor pathways specific to different invasions [4]. Feedback systems must be in place to support tissue repair, remodeling and manage the potential damage to the CNS caused by reactive oxygen species (ROS). Recent studies reveal new details about the role microglia plays in controlling neuronal proliferation and differentiation [5]. Research shows a tight neuronal management, implying that microglia have a direct effect on neuronal disease progression.

Microglia compose 2.5-10% of all brain cells, equivalent one-to-one with the number of neurons. The close association between these two cell types correlates to a vital role in maintaining neural homeostasis, as well as facilitating the removal of apoptotic neurons and weakly connected synapses during brain development. After a hotly contested race to define the origins and characteristics of microglia, neuroresearch has shifted focus mainly toward neurons and often glial cells. Currently, microglial research is undergoing a significant revival (a PubMed search for the keyword "microglia" reveals 87 papers published in 1990, 568 papers in 2000, 1198 papers in 2010 and 2313 papers in 2016), likely due to recognition of their

important role in neuronal health and degeneration. This review focuses on the role microglia fulfils when the neurosystem is injured with regards to immune protection, when that protection goes awry, and how this pathway can lead to neurodegenerative conditions and diseases.

Microglia Function to Provide Brain Immune Protection

When the brain suffers a disturbance such as infection, disease, trauma or other injury, microglia respond with a change in morphology and behavior [6,7]. As a part of the innate immune system, microglia detects pathogens through toll-like receptors (TLRs) and phosphatidylserine receptors [8-10]. Microglia react to stimuli within a matter of minutes and retain their active states for days after traumatic brain insult [11,12]. Microglia can also increase their population density to provide a better defence against antagonistic bacteria, releasing chemo attractive factors that attract immune cells to the CNS and aid T-cells across the bloodbrain barrier [4]. When necessary, activated microglia will physically migrate to specific sites of infection or injury, retracting and extending protrusions to navigate towards the target [13,14].

Microglia use a variety of methods to clear harmful debris, misfolded proteins and damaged cells. Bacterial invasions activate microglia to their defensive role. Lipopolysaccharides (LPS), the main component of the gram-negative bacterial cell wall, stimulate microglia to produce prostaglandins, cytokines and chemokines. More responsive than astrocytes, microglia can also develop an adaptive response to repetitive exposure to LPS with decreased cytokine responses (lower levels of TNF- α and NO secreted) [15]. Streptococcus pneumoniae brings serious cases of meningitis, and microglia may have a role in producing cytokines and chemokines at the blood-brain barrier [16]. These bacteria can cross the BBB, so microglia may be responding directly to the pathogen itself, and NO production contributes to the neuronal damage seen in this case. Multiple bacterial species cause

*Corresponding author: Wolff M. Kirsch, Division of Biochemistry, School of Medicine, Loma Linda University, Loma Linda, CA 92350, USA, Tel: 909-558-7070; E-mail: wkirsch@llu.edu

Received July 26, 2017; Accepted August 02, 2017; Published August 09, 2017

Citation: Sanchez BSN, Kirsch WM (2017) The Role of Microglia in the Injured Neurosystem. J Alzheimers Dis Parkinsonism 7: 362. doi: 10.4172/2161-0460.1000362

Copyright: © 2017 Sanchez BSN, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

J Alzheimers Dis Parkinsonism, an open access journal ISSN:2161-0460

microglia to react in a cytoprotective manner through a series of chemokine and phagocytic responses [17,18]. Immunocompromised patients (such as those diagnosed with AIDS) are at risk to additional viral infections such as human cytomegalovirus (HCMV) which attack the CNS and lead to congenital encephalitis and cognitive defects [19]. Infected astrocytes secrete CCL2/MCP-1 signals to attract microglia to attack the virus. Microglia in turn produce the TNF- α antiviral cytokine to suppress viral replication in astrocytes, though HCMV gene products can also act as analogues to human interleukin molecules and negatively impact microglial defence. When microglia are infected with herpes simplex virus (HSV) they reduce release TNF- α , IL-1 β , CXCL10/IP-10 and other signals that inhibit cellular replication and result in neurodegeneration [20,21].

Neuroinflammation

Brain injury promotes an inflammatory response by microglia that both engage the immune system and tissue repair pathways. Under normal physiological conditions, relevant genes that play a major role in this pathway are repressed, only to be activated when injury or infection is detected by TLRs or similar recognition receptors [22]. Once the receptors bind to the signals of infection, the transduction system is activated to begin transcription of relevant pathways (such as NF-κB and AP-1) as part of the inflammatory response. These signals (including cytokines such as TNF- α and IL-1 β) are then amplified to recruit additional cells to the infection site and induce a protective response. ROS are generated to act as an antimicrobial defence mechanism, but an error in regulation can cause neuronal damage if left to run unimpeded. Microglia are equipped for initiating and maintaining the inflammatory response, can also accept responsibility when the pathway ceases to be regulated and leads to neurodegenerative conditions. Additionally, a recent study has shown that a change in ANK1 gene expression has been observed in brain's microglia cells associated with neuroinflammation [23]. A persistent stimulus or a breakdown in negative feedback systems may promote an overwhelming imbalance of inflammatory signals, leading to neurotoxic factors that simply exacerbate the harmful physiological state.

Neurodegenerative diseases, including Alzheimer's and Parkinson's, have distinct inflammatory induction pathways and mechanisms. Interestingly, these pathways seem to converge rather quickly at the amplification step, with similar or identical mechanisms for neurotoxicity and neuronal apoptosis [22]. Microglial pattern recognition receptors are likely shared between these pathologies, as well as signal transduction pathways (like NF- κ B and AP-1) and NADPH oxidase for ROS release. It is possible that induced positive feedback loops between astrocytes and microglia to explain are responsible for the sustained inflammatory responses that drive neurotoxic environments, creating an independently running pathologic system.

Some work has been done in looking at targeting the inflammatory pathway as a preventative measure or therapeutic intervention. Using the anti-inflammatory agent Luteolin to treat CNS inflammation in mice led to the animals performing better in mental tests, but this hasn't been reproduced into human treatments [24]. Alzheimer's cytokine treatments and amyloid-beta (A β) vaccines have shown no improvement in patients' cognitive abilities, regardless of how early the preventative treatment is given [25,26]. Anti-inflammatory drugs do not reverse neuronal tau pathology in AD either [27].

Aging Microglia May Lead to Neurodegeneration

Like all other cells in an organism, microglia has a defined lifespan.

Murine models have shown an increase from 2% positive readings of MCH II (specific marker to aged microglia) in adult mice to 25% in aged mice microglia [28]. Human microglia loses genes involved in remodeling the cytoskeleton, preventing them from moving towards damage sites [29]. Ageing leads to the cells becoming dysfunctional and increasingly sensitive to minor stimuli. This outcome is most likely due to "microglia priming", a morphological change to the microglia which puts the cell in a state of alertness and ensures the response to a second pathogenic insult would cause the cells to go through phenotypic switching [30]. Exposure to additional concentrations of human leukocyte antigen (HLA) can cause aging microglia to prime the CNS [31]. Microglia activation in white matter increases with age and resting microglia also express basal levels of HLA [32]. Tests done in aged mice show learning and memory impairments associated with deficiencies that come with a decline in neuronal plasticity and CNS inflammation, generating surplus ROS, inflammatory markers, scavenger receptors and TLRs [28,33,34]. A simplified model of aging microglia produces ROS, leading to the activation of redox-sensitive transcription factors, proinflammatory genes and eventual neuroinflammation and neurotoxicity [35].

Numerous attempts to research the neurotoxic effects of aged microglia have been devised. Microglial activity is regulated by antiinflammatory cytokines, including IL-10, IL-4 and TFGB. Unnecessary microglial activation becomes difficult to prevent as numerous key regulatory systems are impaired with age. In studies done with rodents, IL-10 decreases in microglia in aged animals after immune challenge, but is increased significantly in aged microglia [28]. IL-4 sees an overall decreased in aged rat brains, resulting in increased neuroinflammation and reduced LTP [36,37]. Evidence shows a logarithmic age-dependent increase of oxidized proteins and oxidized DNA lesions with age, beyond any loss of protection from antioxidant enzymes. Non-steroidal antiinflammatory drugs (NSAIDs) are a possible treatment mechanism. Clinical trials with NSAIDs show an AD incidence reduction among asymptomatic patients compared to increased risk among those with AD symptoms [38]. Antioxidants such as Coenzyme Q and Vitamin E do not provide any measurable improvements in cognition [22]. It is possible therapeutics based on blocking neuroinflammatory pathways could provide an avenue for treating human cognitive decline [39].

Alzheimer's Disease and Neurodegenerative Conditions

Microglia assist in neuronal modeling via synaptic pruning: the process of removing cells from the neocortex during postnatal development [40]. While this process is vital for the brain to grow during infancy and adolescence, uncontrolled synaptic loss is the defining element of Alzheimer's disease where amyloid-beta (AB) can promote microglia to engage in undesired synaptic removal [41]. Dysregulation of Amyloid Precursor Protein (APP) leads to an accumulation of AB peptides, impairing APP trafficking and causing neurons to degenerate. A β is produced from these stressed cells [42]. Microglia is known to be highly effective for clearing AB, phagocytic and chemotactic responses to remove the peptide from the brain parenchyma [43]. While the majority of $A\beta$ goes undigested and is released from the cell, it has been shown that microglia degrades fibrillar A β within 3 days through phagocytosis [44]. Fibrillar AB can be degraded and phagocytized by peripheral macrophages [45]. Soluble AB is internalized and quickly released without degradation. When neuritic plaques are formed, microglia will rise to clear dead or dying neurons, but these microglial responses to environmental challenges can have detrimental effects on the neurons [46]. One proposed mechanism involves AB causing synaptic damage and promoting microglia to release mediators such

as NO and TNF- α . These mediators are cytotoxic, leading to further synaptic and axonal injury [9]. Another proposed mechanism involves the loss of TDP-43 (DNA-RNA binding protein and transcriptional repressor) functionality, leading to synaptic decreases, reducing or eliminating phagocytic regulation and A β clearance [47]. Furthermore, more than half of the genes expressed that are associated with AD are found to be expressed highly in microglia as compared with other brain cells [48].

Microglial activation has been noted in Parkinson's disease (PD), where pro-inflammatory signals are possible contributors to neuronal loss [49,50]. A LPS-triggered UDP secretion causes the P2Y6 receptor to initiate activation of microglia to express cytokines through the ERK1/2 pathway [51]. Translocator protein (TSPO), which is normally found in mitochondrial membranes, is increased in PD. These proinflammatory signals in microglia could contribute to the neuroprotective reactions seen from phagocytosis and glial propagation [52]. Microglia expresses a pro-inflammatory phenotype in multiple sclerosis as well, initiating demyelination and neurodegeneration. Inactive lesions saw a reduction of microglia density [32,53].

An alternative hypothesis is that microglial loss rather than over activation is responsible for neurodegeneration, as some studies show that activated microglia are not consistently present among the range of AD severities. There is histological evidence showing microglial deterioration before neurofibrillary pathology begins, providing a possible explanation to why drugs targeting anti-inflammation factors have failed to provide an adequate avenue for treatment [27,54,55]. Additionally, fragmented microglia co-localize with neurofibrillary tables.

Neurodegeneration may also come from viral insults. HIV primarily targets microglia, forcing the cell to activate and secrete neurotoxic factors, leading to neurodegeneration [15]. HIV-associated dementia (HAD) is a complex process, one in which microglia plays a central role. Recent work in mice have shown improper activation of cyclin-dependent kinase 5 (Cdk5) under stress conditions, contributes to tau phosphorylation, A β deposits, microgliosis and astrocytosis [56]. Targeting Cdk5 appears to have reduced these AD-related products.

Complement

Cerebral amyloid angiopathy (CAA) is a microvasculopathy found in 75-90% of AD cases [57]. The high rate of this comorbidity renders the pathology highly relevant when studying AD. CAA involves the deposition of $A\beta$ in penetrating cortical arteries coupled with degeneration of vascular smooth muscle cells, leading to vascular fragility and intracerebral hemorrhaging [58,59]. These microbleeds will lead to the cognitive decline associated with AD. The pore-forming complex of proteins that constitute the membrane attack complex (MAC) is associated with CAA-affected blood vessels. A conceivable mechanism for clearing AB by activating the protein CD11b in microglia will also deliver Aβ and C3b to microvascular walls vessels in AD, leading to $A\beta$ deposition is accompanied by complement to form cytolytic MAC to result in vascular fragility [46]. Microglia recognizes complement components and eliminate the synapses that are tagged with them. These can be activated in the aged human brain and drive the system towards neurodegeneration. C1q and C3 are the initiators of the complement cascade. C1q activates C3b which subsequently tags deposits on neurites for elimination. A lack of C1q results in increased C3 levels [60]. It is possible that C1q deposits induce microglial migration and phagocytosis [61].

Metals and Oxidative Stress

Copper is a vital element to redox reactions and specific enzymatic pathways, thus studies of the transport and homeostasis of this metal are the subject of intense research [62]. Copper's role in catalytic reactions is due to its high reactivity. Levels need to be tightly regulated to prevent ROS being produced in inappropriate areas of the cell [63]. Copper binds tightly to A β and produces a very flexible compound that, depending on the conformation, can produce ROS [64]. When combined with A β 1-40, Cu(II) has been shown to enhance microglial activation greater than either on their own. The Cu(II)-A β complex requires NF- κ B for microglial activation and induces mitochondrial super oxide production, promoting neurotoxicity [43]. Glutathione (GSH) can suppress copper toxicity by binding to it and preventing its redox participation [65,66] and GSH depletion may be a contributing factor to neurotoxicity involving the element.

Microglia take, store and release more iron (stored in ferritin) than any other cell in the CNS except oligodendrocytes [67,68]. Iron has been shown to activate microglia through melanotransferrin (MTf), which releases cytokines and nitric oxide. MTf has been detected by active microglia in AD patients, where expression levels are unchanged in Parkinson's disease [69]. Microglia accumulates iron during aging [67]. Advanced age also correlates with an increase in ferritin levels in microglia, along with oligodendrocytes and astrocytes [67]. Activated microglia induces the release of iron from ferritin as well as oxidative stress and proinflammatory cytokines, contributing to PD pathogenesis [70-72].

Animal Models

A variety of animal models have been used for in studying microglia with leeches being the first animal model because of a tightly defined neural system [1]. The invertebrate has a simple system to explore, making it easy to image and manipulate signalling pathways. Rats have been used to harvest brain tissue for microglia activation, while mice have been used to test anti-inflammatory treatments, and derive possible pathways for the mechanisms used [12,73]. Mouse brains are generally difficult to access, particularly in utero when conducting developmental studies [74]. APP23 mouse models have been used in imaging studies in which anti-AB antibody injection was compared with initial levels of AB, showing a microglial overreaction in immunotherapy in subjects with abundant levels of AB [75].

While mice are easier to work with and sidestep ethical issues with regards to human studies, it remains contested whether they are an appropriate substitute for mimicking human pathology. While generally similar between human and mice, human microglia express several unique immune genes and cell cycle regulators. The differences in age-associated gene expression result in a difficult comparison [29]. For example, the study used six-month-old mice with few immune challenges tested against human cells that are exposed to insults every day. Human microglia expresses pro-inflammatory markers where mice do not [32]. Ideally animal models would translate perfectly to human pathology, but when inconsistencies arise, it becomes difficult to read in relation to human subjects. The differences in animal microglial models warrant further study.

Recently, animal models have been used as blueprints for generating human cells for research use. Murine microglia has been created using induced pluripotent stem cells with exposure to differentiation factors, confirming a potential use in medical applications [76]. Human DNA regulatory elements from human microglia have been compared to

J Alzheimers Dis Parkinsonism, an open access journal ISSN:2161-0460

similar mouse factors using tissue culture [48]. Transmembrane protein 119 (Tmem119) is a recently identified microglial marker expressed in both humans and mice, and can be used in studies involving both types of tissue environments [77]. Finally, zebra fish have been a recent animal model to gain traction for in vivo work due to its optical transparency and ease of manipulation.

Concluding Remarks

Beyond their role as immune effectors and deliberate shaping of the developing CNS, there is a solid body of evidence to establish the role microglia play in neurodegenerative diseases. Based on the mechanisms involved in initiating neuroprotective responses to foreign pathogens and physical brain injury, microglial regulation of these pathways can become cytotoxic for the neurons that are meant to be protected. While pathways can go awry at any point for numerous reasons, aging greatly affects microglia functionality leading to less regulation of neuroprotective systems. This ineffectiveness is vital to the development of neurodegenerative diseases that are commonly found in elderly patients such as AD and PD. It is imperative to invest in the research that has accumulated to understand these pathologies and their relation to microglia, to uncover potential targets for potential therapeutic value.

There is a need for further studies to be done in further defining the stages microglia have, as evidence questions the two clear states and reveals more of a fluid matter in which these cells switch between their active and non-active states. Additional effort needs to go into verifying the animal models that are used and how they differ from humans. Understanding the strengths and limitations of the tools we use to understand a process involved in such widespread neurological diseases can potentially open the door to a possible therapeutic intervention.

References

- Rio-Hortega P Del (1932) Microglia. In Cytology and Cellular Pathology of the Nervous System, Penfield W, ed. Hoeber, New York, pp: 482-534.
- McKercher SR, Torbett BE, Anderson KL, Henkel GW, Vestal DJ, et al. (1996) Targeted disruption of the PU.1 gene results in multiple hematopoietic abnormalities. EMBO J 15: 5647-5658.
- Ginhoux F, Lim S, Hoeffel G, Low D, Huber T (2013) Origin and differentiation of microglia. Front Cell Neurosci 7: 1-14.
- 4. Daneman R (2012) The blood-brain barrier in health and disease. Ann Neurol 72: 648-672.
- Graeber MB, Streit WJ (2010) Microglia: biology and pathology. Acta Neuropathol 119: 89-105.
- Block ML, Zecca L, Hong JS (2007) Microglia-mediated neurotoxicity: Uncovering the molecular mechanisms. Nat Rev Neurosci 8: 57-69.
- Colton CA, Wilcock DM (2010) Assessing activation states in microglia. CNS Neurol Disord Drug Targets 9: 174-191.
- Napoli I, Neumann H (2009) Microglial clearance function in health and disease. Neuroscience 158: 1030-1038.
- Neumann H, Kotter MR, Franklin RJM (2009) Debris clearance by microglia: An essential link between degeneration and regeneration. Brain 132: 288-295.
- 10. Ravichandran KS (2003) Recruitment signals from apoptotic cells: Invitation to a quiet meal. Cell 113: 817-820.
- 11. Kreutzberg GW (1996) Microglia: A sensor for pathological events in the CNS. Trends Neurosci 19: 312-318.
- Morioka T, Kalehua AN, Streit WJ (1991) The microglial reaction in the rat dorsal hippocampus following transient forebrain ischemia. J Cereb Blood Flow Metab 11: 966-973.
- Davalos D, Grutzendler J, Yang G, Kim JV, Zuo Y, e al. (2005) ATP mediates rapid microglial response to local brain injury *in vivo*. Nat Neurosci 8: 752-758.

 Nimmerjahn A, Kirchhoff F, Helmchen F (2005) Resting microglial cells are highly dynamic surveillants of brain parenchyma *in vivo*. Science 308: 1314-1318.

- Rock RB, Gekker G, Hu S, Sheng WS, Cheeran M, et al. (2004) Role of microglia in central nervous system infections role. Clin Microbiol Rev 17: 942-964.
- Scheld WM, Koedel U, Nathan B, Pfister HW (2002) Pathophysiology of bacterial meningitis: mechanism of neuronal injury. J Infect Dis 186: 225-233.
- Kielian T, Barry B, Hickey WF (2001) CXC chemokine receptor-2 ligands are required for neutrophil-mediated host defense in experimental brain abscesses. J Immunol 166: 4634-4643.
- Curto M, Reali C, Palmieri G, Scintu F, Schivo ML, et al. (2004) Inhibition of cytokines expression in human microglia infected by virulent and non-virulent mycobacteria. Neurochem Int 44: 381-392.
- Grassi MP, Clerici F, Perin C, Arminio Monforte AD, Vago L, et al. (1998) Microglial nodular encephalitis and ventriculoencephalitis due to cytomegalovirus infection in patients with AIDS: two distinct clinical patterns. Clin Infect Dis 27: 504-508.
- Mahalingam S, Farber JM, Karupiah G (1999) The interferon-inducible chemoknies MuMig and Crg-2 exhibit antiviral activity *in vivo*. J Virol 73: 1479-1491.
- Lokensgard JR, Hu S, Sheng W, VanOijen M, Cox D, et al. (2001) Robust expression of TNF-alpha, IL-1beta, RANTES, and IP-10 by human microglial cells during nonproductive infection with herpes simplex virus. J Neurovirol 7: 208-219.
- Glass CK, Saijo K, Winner B, Marchetto MC, Gage FH (2010) Mechanisms underlying inflammation in neurodegeneration. Cell 140: 918-934.
- Mastroeni D, Sekar S, Nolz J, Delvaux E, Lunnon K, et al. (2017) ANK1 is upregulated in laser captured microglia in Alzheimer's brain; the importance of addressing cellular heterogeneity. pp: 1-11.
- Jang S, Dilger RN, Johnson RW (2010) Luteolin inhibits microglia and alters hippocampal-dependent spatial working memory in aged mice. J Nutr 140: 1892-1898.
- Martin B (2008) Cognitive function over time in the alzheimer's disease antiinflammatory prevention trial (ADAPT): Results of a randomized, controlled trial of naproxen and celecoxib. Arch Neurol 65: 896-905.
- Arvanitakis Z, Grodstein F, Bienias JL, Schneider JA, Wilson RS, et al. (2008) Relation of NSAIDs to incident AD, change in cognitive function, and AD pathology. Neurology 70: 2219-2225.
- Streit WJ, Braak H, Xue QS, Bechmann I (2009) Dystrophic (senescent) rather than activated microglial cells are associated with tau pathology and likely precede neurodegeneration in Alzheimer's disease. Acta Neuropathol 118: 475-485.
- 28. Henry CJ, Huang Y, Wynne AM, Godbout JP (2009) Peripheral lipopolysaccharide (LPS) challenge promotes microglial hyperactivity in aged mice that is associated with exaggerated induction of both pro-inflammatory IL-1β and anti-inflammatory IL-10 cytokines. Brain Behav Immun 23: 309-317.
- Galatro TF, Holtman IR, Lerario AM, Vainchtein ID, Brouwer N, et al. (2017) Transcriptomic analysis of purified human cortical microglia reveals ageassociated changes. Nat Neurosci, pp: 1162-1171.
- Perry VH, Cunningham C, Holmes C (2007) Systemic infections and inflammation affect chronic neurodegeneration. Nat Rev Immunol 7: 161-167.
- Norden DM, Godbout JP (2013) Microglia of the aged brain: Primed to be activated and resistant to regulation. Neuropathol Appl Neurobiol 39: 19-34.
- Zrzavy T, Hametner S, Wimmer I, Butovsky O, Weiner HL, et a. (2017) Loss of homeostatic microglia and patterns of their activation in active multiple sclerosis. Brain 140: 1900-1913.
- Letiembre M, Hao W, Liu Y, Walter S, Mihaljevic I, et al. (2007) Innate immune receptor expression in normal brain aging. Neuroscience 146: 248-254.
- 34. VanGuilder HD, Bixler GV, Brucklacher RM, Farley JA, Yan H, et al. (2011) Concurrent hippocampal induction of MHC II pathway components and glial activation with advanced aging is not correlated with cognitive impairment. J Neuroinflammation 8.
- 35. Nakanishi H, Wu Z (2009) Microglia-aging: Roles of microglial lysosome- and

J Alzheimers Dis Parkinsonism, an open access journal ISSN:2161-0460

mitochondria-derived reactive oxygen species in brain aging. Behav Brain Res 201: 1-7.

- Nolan Y, Maher FO, Martin DS, Clarke RM, Brady MT, et al. (2005) Role of interleukin-4 in regulation of age-related inflammatory changes in the hippocampus. J Biol Chem 280: 9354-9362.
- Maher FO, Nolan Y, Lynch MA (2017) Downregulation of IL-4-induced signalling in hippocampus contributes to deficits in LTP in the aged rat. Neurobiol Aging 26: 717-728.
- Breitner JC, Baker LD, Montine TJ, Meinert CL, Luketsos CG, et al.(2011) Extended results of the Alzheimer disease anti-inflammatory prevention trial (ADAPT). Alzheimers Dement 7: 402-411.
- Floyd RA, Hensley K (2002) Oxidative stress in brain aging. Implications for therapeutics of neurodegenerative diseases. Neurobiol Aging 23: 795-807.
- Voyvodic JT (1996) Cell death in cortical development: How much? Why? So what? Neuron 16: 693-696.
- Hong S, Beja-glasser VF, Nfonoyim BM, Frouin A, Ramakrishnan S, et al. (2016) Complement and microglia mediate early synapse loss in Alzheimer mouse models. Science 352: 712-716.
- 42. Bayer TA, Wirths O, Majtényi K, Hartmann T, Multhaup G, et al. (2001) Key factors in Alzheimer's disease: Beta-amyloid precursor protein processing, metabolism and intraneuronal transport. Brain Pathol 11: 1-11.
- 43. Yu F, Gong P, Hu Z, Qiu Y, Cui Y, et al. (2015) Cu(II) enhances the effect of Alzheimer's amyloid-β peptide on microglial activation. J Neuroinflammation 12: 122.
- 44. Chung H, Brazil MI, Soe TT, Maxfield FR (1999) Uptake, degradation and release of fibrillar and soluble forms of alzheimer's uptake, degradation and release of fibrillar and soluble forms of alzheimer's amyloid β-peptide by microglial cells. J Biol Chem 274: 32301-32308.
- Majumdar A, Chung H, Dollos G, Wang R, Asamoah N, et al. (2008) Degradation of fibrillar forms of Alzheimer's amyloid B-peptide by macrophages. Neurobiol Aging 29: 707-715.
- 46. Zabel M, Schrag M, Crofton A, Tung S, Beaufond P, et al. (2013) A shift in microglial β-amyloid binding in alzheimer's disease is associated with cerebral amyloid angiopathy. Brain Pathol 23: 390-401.
- Paolicelli RC, Jawaid A, Henstridge CM, Valeri A, Merlini M, et al. (2017) TDP-43 depletion in microglia promotes amyloid clearance but also induces synapse loss. Neuron 95: 1-12.
- Gosselin D, Skola D, Coufal NG, Holtman IR, Schlachetzki JCM, et al. (2017) An environment-dependent transcriptional network specifies human microglia identity. Science, p: 356.
- Joers V, Tansey MG, Mulas G, Carta AR (2017) Microglial phenotypes in parkinson's disease and animal models of the disease. Prog Neurobiol 155: 57-75.
- Yang X, Lou Y, Liu G, Wang X, Qian Y, et al. (2017) Microglia P2Y6 receptor is related to Parkinson's disease through neuroinflammatory process. J Neuroinflammation 14: 38.
- Gongol B, Marin TL, Jeppson JD, Mayagoitia K, Shin S, et al. (2017) AICD governs MAST4 kinase activity and cellular hormesis in response to 27-hydroxycholesterol. Science: 80.
- Ghadery C, Koshimori Y, Coakeley S, Harris M, Rusjan P, et al. (2017) Microglial activation in Parkinson's disease using 18F-FEPPA. J Neuroinflammation 14: 8.
- Ramaglia V, Hughes TR, Donev RM, Ruseva MM, Wu X, et al. (2012) C3dependent mechanism of microglial priming relevant to multiple sclerosis. Proc Natl Acad Sci USA 109: 965-970.
- 54. Streit WJ, Sammons NW, Kuhns AJ, Sparks DL (2004) Dystrophic microglia in the aging human brain. Glia 45: 208-212.
- Streit WJ (2005) Microglia and neuroprotection: Implications for Alzheimer's disease. Brain Res 48: 234-239.
- 56. He Y, Pan S, Xu M, He R, Huang W, et al. (2017) Adeno-associated viral

9-mediated Cdk5 inhibitory peptide reverses pathologic changes and behavioral deficits in the Alzheimer's disease mouse model. FASEB J 31: 3383-3392.

- 57. Jellinger KA, Attems J (2005) Prevalence and pathogenic role of cerebrovascular lesions in Alzheimer disease. J Neurol Sci 229-230: 37-41.
- Weller RO, Boche D, Nicoll JAR (2009) Microvasculature changes and cerebral amyloid angiopathy in Alzheimer's disease and their potential impact on therapy. Acta Neuropathol. 118: 87-102.
- Vinters H V (1987) Cerebral amyloid angiopathy. A critical review. Stroke 18: 311-324.
- 60. Zhou J, Fonseca MI, Pisalyaput K, Tenner AJ (2008) Complement C3 and C4 expression in C1q sufficient and deficient mouse models of Alzheimer's Disease. J Neurochem 106: 2080-2092.
- 61. Fraser DA, Pisalyaput K, Tenner AJ (2010) C1q enhances microglial clearance of apoptotic neurons and neuronal blebs and modulates subsequent inflammatory cytokine production. J Neurochem 112: 733-743.
- Ahuja A, Dev K, Tanwar RS, Selwal KK, Tyagi PK (2015) Copper mediated neurological disorder: Visions into amyotrophic lateral sclerosis, Alzheimer and Menkes disease. J Trace Elem Med Biol 29: 11-23.
- Schlief ML, Gitlin JD (2006) Copper homeostasis in the CNS. Mol. Neurobiol 33: 81-90.
- 64. Faller P, Hureau C, La Penna G (2014) Metal ions and intrinsically disordered proteins and peptides: From Cu/Zn amyloid-beta to general principles. Acc Chem Res 47: 2252-2259.
- Jomova K, Valko M (2011) Advances in metal-induced oxidative stress and human disease. Toxicology 283: 65-87.
- Mattie MD, Freedman JH (2004) Copper-inducible transcription: Regulation by metal- and oxidative stress-responsive pathways. Am J Physiol - Cell Physiol 286: 293-301.
- Connor JR, Menzies SL, St Martin SM, Mufson EJ (1990) Cellular distribution of transferrin, ferritin and iron in normal and aged human brains. J Neurosci Res 27: 595-611.
- Beard JL, Connor JD, Jones BC (1993) Brain iron: Location and function. Prog Food Nutr Sci 17: 183-221.
- 69. Berg D, Gerlach M, Youdim MBH, Double KL, Zecca L, et al. (2001) Brain iron pathways and their relevance to Parkinson's disease. Society: 225-236.
- Reynolds AD, Kadiu I, Garg SK, Glanzer JG, Nordgen T, et al. (2008) Nitrated alpha-synuclein and microglial neuroregulatory activities. J Neuroimmune Pharmocol 3: 59-74.
- 71. Stefanova N, Reindl M, Neumann M, Kahle PJ, Poewe W, et al. (2007) Microglial activation mediates neurodegeneration related to oligodendroglial alpha-synucleinopathy: Implications for multiple system atrophy. Mov Disord Off J Mov Disord Soc 22: 2196-2203.
- Reynolds AD, Glanzer JG, Kadiu I, Ricardo-Dukelow M, Chaudhuri A, et al. (2008) Nitrated alpha-synuclein-activated microglial profiling for Parkinson's disease. J Neurochem 104: 1504-1525.
- Lee DD, Rizer J, Hunt JB, Selenica MLB, Gordon MN, et al. (2013) Experimental manipulations of microglia in mouse models of Alzheimer's pathology. Activation reduces amyloid but hastens tau pathology. Neuropathol Appl Neurobiol 39: 69-85.
- 74. Sieger D, Peri F (2013) Animal models for studying microglia: The first, the popular and the new. Glia 61: 3-9.
- Higuchi M (2009) Visualization of brain amyloid and microglial activation in mouse models of Alzheimer's disease. Curr Alzheimer Res 6: 137-143.
- Pandya H, Shen MJ, Ichikawa DM, Sedlock AB, Choi Y, et al. (2017) Differentiation of human and murine induced pluripotent stem cells to microglialike cells. Nat Neurosci 20: 753-759.
- 77. Bennetta ML, Bennetta C, Liddelowa SA, Ajami B, Zamanian JL, et al. (2016) New tools for studying microglia in the mouse and human CNS. Proc Natl Acad Sci U S A 113: E1738-E1746.

Citation: Sanchez BSN, Kirsch WM (2017) The Role of Microglia in the Injured Neurosystem. J Alzheimers Dis Parkinsonism 7: 362. doi: 10.4172/2161-0460.1000362

J Alzheimers Dis Parkinsonism, an open access journal ISSN:2161-0460