

## Screening of Anti-Inflammatory and Anti-Platelet Aggregation Property Studies from *Ipomea staphylina*

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### Abstract

Inflammation is a biological reaction which is a part of non-specific immune response that occurs in a series of reaction to any type of injury, damage caused to the body. Although there has been enough progress in the usage of colossal improved anti-inflammatory drugs, they are known to make up just around half of analgesics, remedying pain by reducing inflammation as opposed to opioids, which might affect the central nervous system. Thus researches in screening up of anti-inflammatory components from the plants have been shown very good progress. Such similar property was been screened using *Ipomea staphylina*, a local weed belonging to Convolvulaceae family. Study included the mechanism by which the Convolvulaceae species *I. staphylina* inhibited inflammatory pathway key enzymes. Plant extract was randomly divided into 3 groups namely water extract, ethanol extract and methanol extract. We screened by randomized trials to estimate levels of all the three key enzymes activity involved in inflammation i.e., PLA<sub>2</sub>, COX (cyclooxygenase) & LOX (lipoxygenase) using the plant sample.

Results show that the water extract of the Plant sample inhibited secretory PLA<sub>2</sub> significantly than the other groups. PLA<sub>2</sub> was significantly inhibited as the concentration of the sample increased and this showed reproducible results. COX and LOX had inhibitory effects but not to a greater extent. These data suggest that the crude plant water extract had molecules that inhibited PLA<sub>2</sub> which is the first enzyme in the inflammatory pathway.

**Keywords:** Anti-inflammatory; Anti platelet aggregation; *Ipomea staphylina*; Phospholipase A<sub>2</sub>; Plant aqueous extract; COX; LOX

### Introduction

The potential of higher plants as source of pharmacologically important has not been much explored. The plants though have great potential in medicinal field, most of the species are yet to be discovered and most of the ones which are identified have not yet submitted to the phytochemical investigation. Thus lots of plant mysteries are open for the researchers to explore and isolate the potential active bio-components which are medically important. One of such important plant genus was *Ipomea* which had great medical values [1-7]. As we all know the second largest genus of Convolvulaceae family is *Ipomea* with around 600 species [8]. *Ipomea* species are usually found in tropic or subtropical regions which are exclusively known for their therapeutic values such as Anti-inflammatory, alkaloids are known to have shown potential anticancer properties. *Ipomea staphylina* is one of the recently identified species of Convolvulaceae family which are infected by *Caviceps purpurea*, leading to the presence of ergot alkaloids in them [9-11]. Bioactive components like low molecular weight phenolic compounds and Flavones are widely distributed among the *Ipomea* genus. Flavones are known to possess potential biochemical effects which inhibit variety of enzymes such as xanthine oxidase, lipoxygenase, etc. [12].

The beneficial effects of the Flavones are known by mechanisms that include antioxidant, anti-inflammatory, anti thrombic, anti-adhesive, anti-vasodilatory and anti-tumor properties too [13-16]. To understand its role in inflammatory pathway, it is important to understand the inflammatory mediators. As we all know that Inflammation is the biological response hosted by the body against any harmful stimuli or foreign visitors (pathogens) or damage to the cells [17,18]. The phases of inflammation could be divided into several phases based on the time period. The inflammatory mediators are usually categorized into exogenous and endogenous mediators. Exogenous

mediators are usually bacterial toxins. Endogenous mediators are again categorized into early phase mediators which include histamine, serotonin, chemo-attractants etc., that are produced by mast cells and platelets and late phase mediators include lipoxygenase, leukotrienes and cyclooxygenase [19,20].

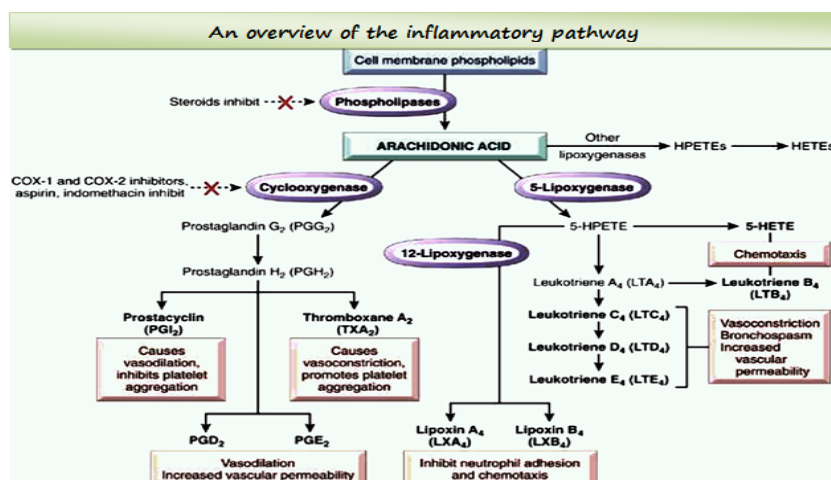
sPLA<sub>2</sub> (secretory Phospholipase A<sub>2</sub>) is known to catalyze the first step in the Arachidonic acid pathway which is keynote substrate for the enzymes Lipoxygenase and cyclooxygenase [21-24]. sPLA<sub>2</sub> are abundantly present in mammalian pancreatic secretion and in snake venom. sPLA<sub>2</sub> acts mostly on the phosphatidyl choline of the membrane lipids which are abundant in the cell membrane at C2 position giving rise to a lysophospholipid and Arachidonic acid [25,26]. This Arachidonic acid serves as substrate for COX and LOX enzymes based on whether it is taking cyclic (COX) or linear (LOX) pathway. If COX enzyme acts on the substrate then Prostaglandins are formed. Based on whether it is COX 1 or COX 2 enzyme that acts on Prostaglandin, the products formed are Prostacyclins, PGD<sub>2</sub> (Prostaglandin D<sub>2</sub>), PGE<sub>2</sub> (Prostaglandin E<sub>2</sub>) and Thromboxanes respectively. Prostacyclins are known to cause vasodilation and inhibit platelet aggregation. Thromboxanes have counter effect unlike Prostacyclins. They are known to cause vasoconstriction and promote platelet aggregation (Figure 1).

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**Figure 1:** This figure illustrates the pictorial overview of the inflammatory pathway.

Arachidonic acid forms various HPETEs (hydro-peroxy-eicosa-tetraenoic acid) like 5, 12, 15 HPETEs by 5, 12, 15 Lipoxygenases respectively [27-32]. 5 HPETEs will form leukotrienes which enhance vasoconstriction, Bronchospasm and increased vascular permeability. 12, 15 LOX will form Lipoxins and Hepoxins respectively where Lipoxins are known to have chemotactic effect and inhibit neutrophil adhesion whereas Hepoxins function still remains unexplored. In this study we have screened the anti-inflammatory and antiplatelet aggregation properties from *Ipomea staphylina*.

## Materials and Methods

### Preparation of plant extract and protein estimation of snake venom

Known amount of dried leaves of *Ipomea staphylina* were taken with little amount of water and centrifuged at 1500 rpm for 15 minutes at 4°C. Supernatant obtained was used for further experiments. Similar procedure was carried out for solvent extraction using 50% ethanol which was used for Indirect Hemolytic Assay.

Concentration of protein present in snake venom (*Daboia russelii* (Russel viper)) was estimated using BSA as standard (1-75 µg) by Lowry's method [33].

### Indirect hemolytic assay

Erythrocytes were isolated from fresh blood taken from the donor and it was added to 3% Trisodium citrate (9:1 v/v) by centrifuging at 1500 rpm for 15 min at room temperature by using Phosphate buffer saline until clear supernatant was obtained. The pellet was used for further study.

Indirect Hemolytic Assay was carried out using PLA2 standard as described by Boman and Kaletta in 1957 [34]. One ml of erythrocytes was taken to which 1 ml of egg yolk and PBS was added in the ratio of 1:8. One ml of this suspension was added into different concentration of venom and incubated for 30 minutes at 37°C. This reaction mixture was stopped by adding ice cold PBS and centrifuged at 1500 rpm, 4°C for 10 min. The supernatant was measured colorimetrically at 540 nm.

### Determination of platelet aggregation activity

Ardlie and Han method was employed to isolate platelets from fresh human donor [35]. Turbidometric method was employed to follow platelet aggregation where agonist platelet aggregating agent was added to platelet rich plasma the platelets aggregate and gradually becomes clear increasing light transmission. The change in turbidity is observed and plotted on a recorder.

### Polyphenols and flavones identification

Estimation of total polyphenols content was carried out by Folin-Ciocalteu method using gallic acid as standard. Similarly Flavones were also estimated by Woisky & Salatino Colorimetric method using Quercetin as standard.

These were further supported by Concentrated Sulphuric acid test and Mg-HCl test.

### Malonyl dialdehyde assay

Known amount was plasma was added to blank, control and sample tubes to which known amount of BHT and TCA was added and centrifuged at 10,000 rpm for 20 min followed by incubation in boiling water bath for 30 min. Absorbance was read at 532 nm spectrophotometrically.

### Lipoxygenase assay

Isolation of PMNLs of human blood was carried out by Ficoll-Histopaque density gradient and hypotonic lysis of erythrocytes. These PMNLs were re suspended in PBS and sonicated at 20-30s at 20 kHz to release the cytosolic 5 LO enzyme into the solution. This solution was centrifuged at 100,000g for 30 min at 4°C. The supernatant is directly used as source of enzyme. 5 lipoxygenase enzymatic assays was carried out using buffer, DTT, ATP and CaCl2 and the end product HETE was measured at 234 nm using Shimadzu spectrophotometer.

## Results and Discussion

Indirect Hemolytic Assay showed significant result. Plant aqueous extract showed almost 90% of PLA2 inhibition (Figures 2 and 3). Thus this implies that the plant is a potent inhibitor of phospholipase A2

which also implies its important role in inhibiting the first step in inflammatory pathway.

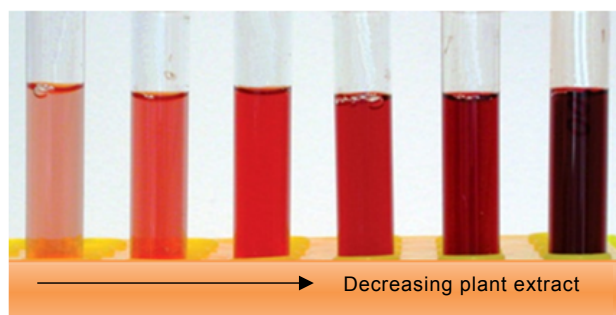
However plant extract did not show any significant effect in Platelet aggregation activity. This could be viewed by the recorder plotted graph (Figure 4). This was further confirmed by Malonyl Dialdehyde Assay.

The estimation of polyphenols and flavones showed erect graph which indicates its presence, (Figures 5 and 6). This was further supported by conc. sulphuric acid test. Mg HCl test clearly produced pink to orange effervescence see (Figure 7).

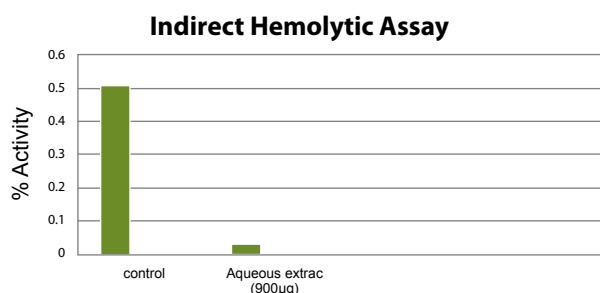
However plant extract decreased the activity of 5 Lipoxygenase enzymes significantly. This might be helpful in improving the endothelial function which has role in decreasing coronary heart disease. This also has significant role in other medical therapeutics [36-39] (Figure 8).

## Conclusion

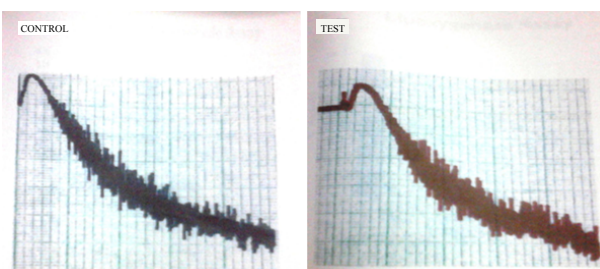
Phospholipases A2 superfamily plays a crucial role in the biological



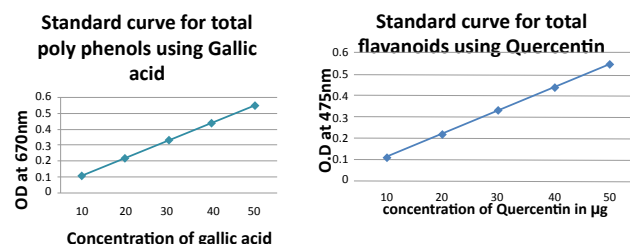
**Figure 2:** This figure shows clearly the inhibition of the PLA2 in first test tube decreasing gradually as the plant aqueous extract concentration decreased.



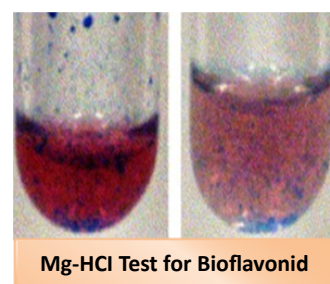
**Figure 3:** Indirect Hemolytic Assay showing almost 90% inhibition of PLA2 by plant aqueous extract.



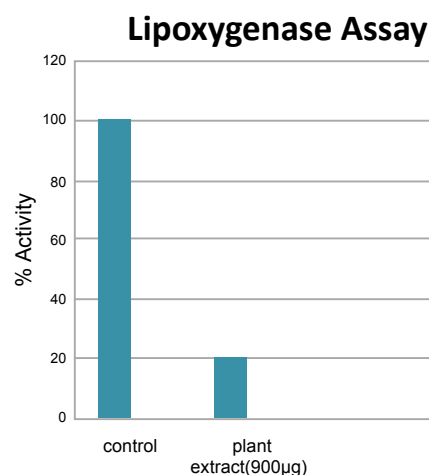
**Figure 4:** The graphs do not show any significant effect of plant extract on Platelet aggregation activity.



**Figure 5,6:** These both graphs represent the presence of Polyphenols and Flavanoids in the plant aqueous extract.



**Figure 7:** Mg-HCl Test to identify the presence of Flavonoid in the diluted plant extract.



**Figure 8:** Graphical representation of Lipoxygenase Assay. This shows that the plant aqueous extract significantly reduced the activity of the enzyme 5 LOX.

system by extending diverse cellular responses including phospholipid digestion. *Ipomea staphylina* aqueous extract are a potent biological compound capable of efficiently inhibiting PLA2 enzyme. However the biological conditions matter in its variability of the result in the inhibition. The extract did not show much significant effect on platelet aggregation which was further confirmed by Malonyl Dialdehyde assay. The plant extract however contained the medically important compounds that possessed diverse health benefits such as Flavonoid and polyphenols. The molecules were not exactly identified, thus further studies have to be carried out for the purification of the active ingredient and their effects on animal model. If the crude extract is capable of exhibiting such beneficial effects then the isolated molecules might show much significant result.

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## References

1. Luís A, Domingues F, Duarte AP (2011) Bioactive compounds, RP-HPLC analysis of phenolics, and antioxidant activity of some Portuguese shrub species extracts. *Nat Prod Commun* 6: 1863-72.
2. Shojaii A, Dabaghian FH, Goushegiri A, Fard MA (2011) Antidiabetic plants of Iran. *Acta Med Iran* 49: 637-42.
3. Bruschi P, Morganti M, Mancini M, Signorini MA (2011) Traditional healers and laypeople: a qualitative and quantitative approach to local knowledge on medicinal plants in Muda (Mozambique). *J Ethnopharmacol* 138: 543-63.
4. Suksomboon N, Poolsup N, Boonkaew S, Suthisang CC (2011) Meta-analysis of the effect of herbal supplement on glycemic control in type 2 diabetes. *J Ethnopharmacol* 137: 1328-33.
5. Hamsa TP, Kuttan G (2011) Augmentation of cellular immune response by *Ipomea obscura* and *Ipobscure* alkaloid attenuates tumor growth in mice. *Can J Physiol Pharmacol* 89: 259-68.
6. León-Rivera I, Herrera-Ruiz M, Estrada-Soto S, Gutiérrez Mdel C, Martínez-Duncker I, et al. (2011) Sedative, vasorelaxant, and cytotoxic effects of convolvulin from *Ipomea tyrianthina*. *J Ethnopharmacol* 135: 434-9.
7. Ng ZX, Chai JW, Kuppusamy UR (2011) Customized cooking method improves total antioxidant activity in selected vegetables. *Int J Food Sci Nutr* 62: 158-63.
8. Austin DF, Huáman Z (1996) A synopsis of *Ipomea* (Convolvulaceae) in the Americas. *Taxon* 45: 3-38.
9. Vega J, Goecke H, Santamarina M (2011) [Retroperitoneal fibrosis associated with chronic use of ergotamine: report of one case]. *Rev Med Chil* 139: 489-94.
10. Murad MH, Miller FA, Glockner J (2011) Multi-system fibrosis and long-term use of ergotamine. *Ann Acad Med Singapore* 40: 327-8.
11. Delgado-Rosas F, Gómez R, Ferrero H, Gaytan F, Garcia-Velasco J, et al. (2011) The effects of ergot and non-ergot-derived dopamine agonists in an experimental mouse model of endometriosis. *Reproduction* 142: 745-55.
12. Echart Eich (2008) Solanaceae and Convolvulaceae - Secondary Metabolites: Biosynthesis, Chemotaxonomy, Biological and Economic Significance: a Handbook. Springer-Verlag Berlin Heidelberg, Germany.
13. Zheng CD, Li G, Li HQ, Xu XJ, Gao JM, et al. (2010) DPPH-scavenging activities and structure-activity relationships of phenolic compounds. *Nat Prod Commun* 5: 1759-65.
14. Donnelly PE, Churilla TM, Coco MG Jr, Vinson JA (2010) Vitamin enhanced waters and polyphenol rich beverages analyzed for antioxidant capacity and antioxidants/calorie. *Nutrients* 2: 1290-6.
15. Tsao R (2010) Chemistry and biochemistry of dietary polyphenols. *Nutrients* 2: 1231-46.
16. Lee JH, Kim GH (2010) Evaluation of antioxidant and inhibitory activities for different subclasses flavonoids on enzymes for rheumatoid arthritis. *J Food Sci* 75: 212-7.
17. Ferrero-Miliani L, Nielsen OH, Andersen PS, Girardin SE (2007) Chronic inflammation: importance of NOD2 and NALP3 in interleukin-1 $\beta$  generation. *Clin Exp Immunol* 147: 227-35.
18. Rovenský J, Svík K, Rovenská E, Stvrtinová V, Stancíková M (2008) Effects of purified micronized flavonoid fraction (Detralex) on prophylactic treatment of adjuvant arthritis with methotrexate in rats. *Isr Med Assoc J* 10: 377-80.
19. Vick BA, Zimmerman DC (1987) Pathways of Fatty Acid hydroperoxide metabolism in spinach leaf chloroplasts. *Plant Physiol* 85: 1073-8.S
20. Lefkowitz JB, Evers AS, Elliott WJ, Needleman P (1986) Essential fatty acid deficiency: a new look at an old problem. *Prostaglandins Leukot Med* 23: 123-7.
21. Sadashiva MP, Nataraju A, Mallesha H, Rajesh R, Vishwanath BS, et al. (2005) Synthesis and evaluation of trimethoxyphenyl isoxazolidines as inhibitors of secretory phospholipase A2 with anti-inflammatory activity. *Int J Mol Med* 16: 895-904.
22. Narendra Sharath Chandra JN, Ponnappa KC, Sadashiva CT, Priya BS, Nanda BL, et al. (2007) Chemistry and structural evaluation of different phospholipase A2 inhibitors in arachidonic acid pathway mediated inflammation and snake venom toxicity. *Curr Top Med Chem* 7: 787-800.
23. Yoshida K, Shinohara H, Haneji T, Nagata T (2007) Arachidonic acid inhibits osteoblast differentiation through cytosolic phospholipase A2-dependent pathway. *Oral Dis* 13: 32-9.
24. Trevisi L, Bova S, Cargnelli G, Ceolotto G, Luciani S (2002) Endothelin-1-induced arachidonic acid release by cytosolic phospholipase A2 activation in rat vascular smooth muscle via extracellular signal-regulated kinases pathway. *Biochem Pharmacol* 64: 425-31.
25. Cohen F, Jaffe BM (1973) Production of prostaglandins by cells in vitro: radioimmunoassay measurement of the conversion of arachidonic acid to PGE2 and PGF2  $\alpha$ . *Biochem Biophys Res Commun* 55: 724-9.
26. Silver MJ, Smith JB, Ingberman C, Kocsis JJ (1973) Arachidonic acid-induced human platelet aggregation and prostaglandin formation. *Prostaglandins* 4: 863-75.
27. Poyser NL (1973) The physiology of prostaglandins. *Clin Endocrinol Metab* 2: 393-410.
28. Bormann BJ, Walenga RW, Showel HJ, Becker EL (1981) Mediated uptake of arachidonic acid by rabbit neutrophils. *FEBS Lett* 136: 293-7.
29. Capdevila J, Yadagiri P, Manna S, Falck JR (1986) Absolute configuration of the hydroxyicosatetraenoic acids (HETEs) formed during catalytic oxygenation of arachidonic acid by microsomal cytochrome P-450. *Biochem Biophys Res Commun* 141: 1007-11.
30. Jacinová V, Nosál R, Danihelová E (1999) Decreased arachidonic acid liberation participates in the anti-aggregatory effect of the histamine H(1)-receptor antagonist Bromadryl. *Platelets* 10: 391-8.
31. Horrobin DF (2001) Phospholipid metabolism and depression: the possible roles of phospholipase A2 and coenzyme A-independent transacylase. *Hum Psychopharmacol* 16: 45-52.
32. Onguru O, Casey MB, Kajita S, Nakamura N, Lloyd RV (2005) -2 and thromboxane synthase in non-endocrine and endocrine tumors: a review. *Endocr Pathol* 16: 253-77.
33. LOWRY OH, ROSEBROUGH NJ, FARR AL, RANDALL RJ (1951) Protein measurement with the Folin phenol reagent. 193: 265-75.
34. BOMAN HG, KALETTA U (1957) Chromatography of rattlesnake venom; a separation of three phosphodiesterases. *Biochim Biophys Acta* 24: 619-31.
35. Ardlie NG, Han P (1974) Enzymatic basis for platelet aggregation and release: the significance of the 'platelet atmosphere' and the relationship between platelet function and blood coagulation. *Br J Haematol* 26: 331-56.
36. Li L, Berthelette C, Chateaufneuf A, Ouellet M, Sturino CF, et al. (2010) Potent and selective 5-LO inhibitor bearing benzothiophene pharmacophore: discovery of MK-5286. *Bioorg Med Chem Lett* 20: 7440-3.
37. Konatė K, Souza A, Coulibaly AY, Meda NT, Kiendrebeogo M, et al. (2010) In vitro antioxidant, lipoygenase and xanthine oxidase inhibitory activities of fractions from *Cienfuegosiadigitata* Cav., *Sida alba* L. and *Sidaacuta* Burn f. (Malvaceae). *Pak J BiolSci* 13: 1092-8.
38. Czubowicz K, Czapski GA, Cieřlik M, Strosznajder RP (2010) Lipoygenase inhibitors protect brain cortex macromolecules against oxidation evoked by nitrosative stress. *Folia Neuropathol* 48: 283-92.
39. Raghavendra RH, Diwakar BT, Lokesh BR, Naidu KA (2006) Eugenol- the active principle from Cloves inhibits 5-Lipoygenase activity and Leukotriene C4 in human PMNLs. *Journal of Prostaglandins, Leukotrienes and Essential Fatty Acids* 74: 23-27.