Effects of Triterpenoids on Herpes Simplex Virus Type1 (Hsv-1) In Vitro

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Abstract

Herpes simplex virus (HSV) infection is an ancient disease, which is mostly easily acquired. HSV remains a common cause of ulcerative mucocutaneous disease today in both healthy and immunocompromised hosts. Due to the increasing HIV infections, many immuno compromised persons are infected with HSV-1 and/or HSV-2. However, there is no effective treatment against the disease but only drugs to manage the infections. The aim of the study was to evaluate antiviral effects of triterpenoids. Five naturally occurring triterpenoids including lupenone, 3-oxo-20-(29)-lupen-30al, lupeol, betulin and 3-(E)-trans coumaroylbetulin were tested on the infectivity of HSV-1 in vitro using the VERO cell monolayer. The activity was observed as inhibition of cytopathic effects on a VERO cell monolayer infected with a virus, using the end-point titration method. The antiviral activity was expressed as the maximal non-toxic dose (MNTD) as reduction factor (RF). Betulin was active at a concentration of 50ug/ml against the infectivity of HSV-1. Lupenone, lupeol, and 3-(E)-trans coumaroylbetulin had no effects on infectivity but at 100 ug/ml they all reduced intracellular replication of the HSV-1 virus. Lupeol, and 3-(E)-trans coumaroylbetulin disrupted 100% of the cell monolayer at a concentration of 50 µg/ml. In conclusion, these naturally occurring triterpenoids possess some anti-HSV-1 activity. Betulin showed a pronounced antiviral activity than the rest of the compounds tested.

Introduction

Herpes simplex viruses (both types, HSV-1 and -2) are common human pathogen which causes a broad spectrum of illness ranging from symptomatic infection to disseminated diseases, such as labial herpes, genital herpes, keratitis, and encephalitis [1-4]. HSV-1 infections are very common and mostly affect adult people [5].

The primary symptoms of HSV infection include a prodromal “flu-like” syndrome, with fever, headache, malaise, diffuse myalgias, followed by local symptoms consisting of genital itching, tenderness, dysuria, lesions, and painful papules over genital regions and ulceration [6,7]. Studies have shown that reactivation of latent HSV-1 can occur as a result of physical or emotional stress [8,9]. In Scotland, it found that 40% of genital herpes was due to HSV1. HSV-1 and HSV-2 are able to infect and reactivate in the same anatomic area, although the natural history of these infections is markedly different [10]. The HSV2 recurring more frequently than HSV1, so most clinical reactivations are likely to be due to HSV2 [10].

In immunosuppressed patients, the clinical manifestation of the disease exhibits different severity and several patients always encounter recurrent attacks [11,12]. In addition, HSV is involved in several ocular diseases [13-15]. For instance, herpetic stromal keratitis (HSK) is an immunopathological disease, which is one of the leading causes of blindness in the western world [16]. Therefore, anti-HSV molecules could be of great relevance for infectious epithelial keratitis, neurotrophic keratopathy, stromal keratitis and endotheliitis [17].

A major advance in antiviral therapy has been the use of effective anti-herpes drugs, such as acyclovir, ganciclovir, valaciclovir, penciclovir, famciclovir (a prodrug of penciclovir and vidarabine). Among these acyclovir is the most commonly used drug for treatment of HSV infections, followed by penciclovir/famciclovir, acyclovir and related nucleoside analogues. However, the appearance of acyclovir and related nucleoside analogues-resistant HSV strains has become evident in immunosuppressed patients, such as organ transplant recipients and patients with acquired immunodeficiency syndrome. A study has shown resistance to acyclovir and related nucleoside analogues can occur following mutation in either HSV thymidine kinase (TK) or DNA polymerase [18]. Moreover, it should be considered that these drugs are very expensive and several patients with frequent attacks may not be able to afford the cost of long-term treatment [19]. Therefore, new antiviral agents exhibiting different mechanisms of action and affordable are urgently needed.

Interestingly, the uses of traditional natural products have increased [20-23]. It has been suggested that aqueous and ethanolic extracts from plants used in allopathic medicine are potential sources of antiviral and antitumor agents [23,24]. Furthermore, the selection of crude plant extracts for screening programs has the potential of being more successful in its initial steps than the screening of pure compounds isolated from natural products [21,25]. Therefore, Acacia mellowifer stem barks, which are commonly used in treatment of pneumonia, malaria, primary infection of syphilis, and stomachaches were analyzed in this study. The dichloromethane extract was fractionated and purified through HPLC to give six pure compounds. These compounds were examined for anti-HSV activity according to method described by [23].

Materials and Methods

Isolation of compounds

Grounded stem bark of Acacia mellifera (2.25 kg) was extracted at room temperature, successively with dichloromethane (CH2Cl2) and methanol (MeOH) for 24 hours each. The crude extract was evaporated under reduced pressure in vacuo to yield CH2Cl2 extract...
and MeOH extract. The crude extract of dichloromethane (CH₂Cl₂) was initially fractionated using silica gel column chromatography. Elution was started with n-hexane, and the polarity of solvents was increased in a stepwise manner in the following sequence: n-hexane–ethyl acetate, followed by ethyl acetate, then ethyl acetate–methanol and finally methanol. Fractions of 50 ml each were collected and examined on silica TLC plate. Identical fractions were pooled together. Further purification was done by using HPLC. The structures of pure compounds were elucidated using ¹H NMR and ¹³C NMR.

### Antiviral assays

**End-point titration technique (EPTT)** - The technique described by [23] with a few modifications, was used. Briefly, confluent monolayer Vero cells were grown in 96-well flat-bottomed plates. Two-fold dilutions of the extracts in maintenance medium, identical to growth medium except for FBS, which was 2%, were added 1 h before viral infection. Cells were infected with 0.1 ml of serial ten-fold dilutions of the previously titrated virus suspension and incubated again at 37°C in humidified 5% CO₂ atmosphere for 48 h. Controls consisted of infected cells with HSV-1 serial ten-fold dilutions in the absence of the extracts, treated non-infected and untreated non-infected cells. Furthermore all tests were compared with positive controls (acyclovir and 3-methylquercetin) tested simultaneously under identical conditions. The antiviral activity is expressed as the maximal non-toxic dose (MNTD) of the test extract needed to obtain the reduction virus titer. The reduction in virus titer was determined as the reduction factor (RF) of the virus titer, i.e. the ratio of the virus titer in the absence over virus titer in the presence of the extract. Three assays were carried out in duplicate with at least 5 concentrations. The results are expressed as the mean obtained from 3 different assays. The RF value of Acyclovir and 3-methylquercetin was 10⁴ at MNTD of 0.1 µg/ml. Therefore, the extracts with RF values of 1x10⁻² to 1x10⁻⁴ indicated a pronounced antiviral activity.

### Results and Discussion

As far as studies on isolated compounds, different research groups have recently reported some compounds, which exhibit potent antiviral activity against HSV-1 [23,26,27] has also reported and explained that extracts with RFs of the virus titer of > 1x10³ show relevant antiviral activity. Therefore looking at the results of triterpenoids tested against the HSV-1 virus in Table 1, betulin exhibited the moderately anti-HSV-1 activity at 50 µg/ml and lower anti-HSV-1 activity between 1-10 µg/m. But lupenone, 3-oxo-20 (29)-lupen-30al, lupeol and 3-(E)-trans coumaroylbetulin lacked anti-HSV-1 activity. Although, Madureira et al. (2003) had reported lupenone isolated from the whole

<table>
<thead>
<tr>
<th>Sample name</th>
<th>Conc (µg/ml):</th>
<th>RF</th>
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<tbody>
<tr>
<td>Lupenone</td>
<td>100; N 1-100</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>50; T/2 1-25</td>
<td>1</td>
</tr>
<tr>
<td>3-oxo-20 (29)-lupen-30-al</td>
<td>100; T 25-100</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>T/2 1-25</td>
<td>1</td>
</tr>
<tr>
<td>Lupeol</td>
<td>100; N 25-100</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>T/2 1-25</td>
<td>1</td>
</tr>
<tr>
<td>Betulin</td>
<td>100; N 100; T</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>T/2 50; T/4 25</td>
<td>10⁻²</td>
</tr>
<tr>
<td></td>
<td>10-1</td>
<td>1</td>
</tr>
<tr>
<td>3-(E)-trans coumaroylbetulin</td>
<td>100; N 50-100</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>T/2 1-25</td>
<td>1</td>
</tr>
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</table>

**Table 1**: Results of the antiviral activity screening of triterpenoids.

![Figure 1](image-url)
References